

Thesis Title Proteomic Analysis of Lipase Producing Soil-Isolated Bacteria

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ABSTRACT

Forty strain bacteria were isolated from oil-contaminated soil. These bacteria were screened for lipase production on selective medium using neutral red as an indicator. Nine bacterial isolates could form red area around their colonies. These bacteria were further measured for lipase activity by using *p*-NPL (*p*-nitrophenyl laurate) as substrate. The isolate CFS 14 could produce the highest enzyme activity of 163.51 U/ml. Thus this bacterium was chosen for this experiment. By 16s rDNA sequencing technique, the bacterial isolate CFS 14 was identified to be *Psuedomonas xinjiangensis* with 99.6% similarity. The optimal condition for lipase production of this bacterium was determined. The optimum temperature and pH were 37°C and 8.0, respectively. Among various divalent cations tested, MgCl₂ was found to be a good enhancer for this bacterium to produce lipase. At 0.5% MgCl₂ the highest activity was observed at 270.27 U/ml which was higher than control. Confirmed by native-polyacrylamide gel electrophoresis, lipase protein band specifically stained as red by FastRed/TR Salt was more intensified in bacteria grown in 0.5% MgCl₂ than without MgCl₂. Due to the property to enhance lipase production of Mg²⁺, this condition was chosen for proteomic analysis using 2DE technique coupled with MALDI-ToF/MS to identify differentially expressed proteins. *Psuedomonas xinjiangensis* strain CFS 14 was cultured in 2 different media containing 0% and 0.5% MgCl₂. The proteome analysis revealed 6 highly expressed proteins, 1 lower expressed protein and 8 protein spots expressed only in 0.5% MgCl₂ condition. Five spots were chosen to annotate by MALDI-ToF/MS. Only 2 protein spots could be identified which were likely to be lipoprotein and hypothetical protein cdivTM_18888.

