CHAPTER 2

BIODIVERSITY AND TAXONOMY

2.1. Introduction

There have been relatively few studies on the fungi associated with palms in Thailand (Aramsiriujiwet, 1996; Hyde *et al.*, 2002; McKenzie *et al.*, 2002; Pinnoi *et al.*, 2004, 2006; Pinruan *et al.*, 2002, 2004, 2007, 2008; Sarapat, 2003; Hidayat *et al.*, 2006), and most of the published data were carried out in southern Thailand where species of palms at freshwater to brackish area ecosystems are widely distributed. The information of palmicolous fungi in this area was reviewed in the chapter 1. In northern Thailand, the publication on the palmicolous fungi is relatively lacking. The first report on palmicolous fungi in this area was probably reported by Techa (2001), however, the data were from unpublished thesis. Therefore, the report by Hidayat *et al.* (2006) regarding the *Oxydothis* species associated with Thai Dwarf Fishtail Palm (*Wallichia siamensis*) is arguably the first publication of fungi from northern Thailand.

Although many new species of several palmicolous genera have been reported in Thailand during the last approximately 10 years, however, information on fungi associated with necrotic leaflet of palms are few, as the previous studies mainly focused on decaying fronds (Aramsiriujiwet 1996; Hidayat *et al.*, 2006; Hyde *et al.*, 2002; McKenzie *et al.*, 2002; Pinnoi *et al.*, 2004, 2006; Pinruan *et al.*, 2002, 2004; Sarapat 2003). Furthermore, study on fungi potentially causing necrotic spots on economically used palms e.g. areca palm (*Areca catechu*), sago palm (*Metroxylon sagu*) and coconut (*Cocos nucifera*), are particularly important. It is well-known that

some fungi generally cause local and general necrosis of plant tissues, and they often cause reduced growth (stunting) of plant organs or entire plants (Agrios, 2005). Leaf spot, blight, blotch, and anthracnose are common necrosis symptoms caused by fungi on plants. Necrosis on palm leaflets, such as leaf spot symptoms, can be caused by species of *Oxydothis*, *Astrosphaeriella*, *Guignardia*, *Maculatipalma* and *Mycosphaerella* (Fröhlich, 1992; Fröhlich and Hyde, 1994, 1995a, b, c, 1998, 2000; hyde, 1995b; Hyde and Fröhlich, 1995a; Hyde, *et al.*, 1997).

The diversity of fungi associated with palms, particularly W. siamensis, and other terrestrial palms collected from Chiang Mai province and other locations are presented in this chapter. This includes endophytes, pathogens and saprobes. Specimens which are considered as new species or interesting taxonomically are described and illustrated. A total 181 species of fungi in 128 genera from 19 palm species have been investigated and examined (table 2.1). These include 89 Ascomycetes species in 52 genera and 29 families. Two Zygomycetes genera were recorded, both from family Mucoraceae. Two Basidiomycetes genera from two families, Agricostilbaceae and Schizophyllaceae, were also recorded. The remaining fungal genera comprised 23 species of Coelomycetes from 17 genera, and 65 Hyphomycetes species from 55 genera. Twenty-four species are new to science of which 7 species have been validly published or are in the process of publication (Hidayat et al., 2006, 2007; To-anun et al., 2009). Sixty-two species are new records to Thailand. Of the total species recorded, 92 species are found on W. siamensis. The most common families found include Botryosphaeriaceae, Mycosphaerellaceae, Xylariaceae, Pleosporaceae and Amphisphaeriaceae. Anthostomella, Astrosphaeriella and Oxydothis are more common on palms

Table 2.1 List of palmicolous fungi found on *Wallichia siamensis* and other palms during the present study.

Herbaria number (FIH)	Fungal species	Family	Host	References
332	Acremonium alternatum Link	Incertae sedis	Wallichia siamensis Becc.	Magazin Ges. naturf. Freunde, Berlin 3: 15 (1809)
325	Acrodictys bambusicola M.B. Ellis	Incertae sedis	Wallichia siamensis Becc.	<i>Mycol. Pap.</i> 79 : 6 (1961)
185	Acrogenospora sphaerocephala (Berk. & Broome) M.B. Ellis	Hysteriaceae	Wallichia siamensis Becc.	Dematiaceous Hyphomycetes: 114 (1971)
329	Agaricostilbum palmicola J.E. Wright	Agaricostilbaceae (Basidiomycota)	Borassus flabellifer L.	<i>Mycologia</i> 62 (4):
044	Alternaria alternata (Fr.) Keissl.	Pleosporaceae	Borassus flabellifer L.	Beih. bot. Zbl., Abt. 1 29 (2): 434 (1912)
335	Alternaria citri Ellis & N. Pierce	Pleosporaceae	Borassus flabellifer L.	Bot. Gaz. 33: 234 (1902)
143	Alternaria tenuissima (Kunze) Wiltshire	Pleosporaceae	Borassus flabellifer L.	Trans. Br. mycol. Soc. 18: 157 (1933)
115	Amphisphaeria umbrina (Fr.) De Not.	Amphisphaeriaceae	Caryota mitis Lour.	Sfer. Ital.: 69 (1863)
208	Anthostomella frondicola K.D. Hyde, J. Fröhl. & Joanne E. Taylor	Xylariaceae	Daemonorops sp.	Sydowia 50 : 71 (1998)
056, 073	Anthostomella leptospora (Sacc.) S.M. Francis	Xylariaceae	Borassus flabellifer L.	<i>Mycol. Pap.</i> 139 : 24 (1975)
125 Yrig	Anthostomella limitata Sacc.	Xylariaceae	Wallichia siamensis Becc.	Atti Accademia Scientifica VenetoTrentino- Istriana 4: 101
028	Anthostomella nitidissima (Durieu & Mont.) Sacc.	Xylariaceae	Cocos nucifera L.	(1875) Syll. Fung. 1: 279 (1882)
146	Anthostomella punctata (Roberge) Sacc.	Xylariacaeae	Wallichia siamensis Becc.	Sylloge Fungorum 1: 278 (1882)

Herbaria number (FIH)	Fungal species	Family	Host	References
324; 146	Anthostomella puiggarii Speg.	Xylariaceae	Livistona chinensis R.Br.; Wallichia siamensis Becc.	Boln Acad. nac. Cienc. Córdoba 23 : 467 (1919)
155	Anthostomella zongluensis K.D. Hyde	Xylariaceae	Borassus flabellifer L.	<i>Nova Hedwigia</i> 62 : 333 (1996)
123, 353	Apiospora siamicola Hidayat & To-anun, sp. nov.	Apiosporaceae	Wallichia siamensis Becc.	In this study
339; 356	Apiosporina rhapisicola Hidayat & To-anun, sp. nov.	Venturiaceae	Rhapis sp.; Wallichia siamensis Becc.	In this study
124, 354	Arecomyces foliicola Hidayat & To-anun, sp. nov.	Hyponectriaceae	Wallichia siamensis Becc.	In this study
366	Arecomyces frondicola K.D. Hyde	Hyponectriaceae	Wallichia siamensis Becc.	<i>Sydowia</i> 48 : 232 (1996)
066; 109; 426	Arthrinium phaeospermum (Corda) M.B. Ellis	Apiosporaceae	Areca catechu L.; Borassodendron sp.; Wallichia siamensis Becc.	Mycol. Pap. 103 : 8 (1965)
183	Aspergillus niger Tiegh.	Trichocomaceae	Borassus flabellifer L.	Annls Sci. Nat., Bot., sér. 5 8: 240 (1867)
320	Astrosphaeriella caryotae Hidayat & To-anun, sp. nov.	Melanommataceae	Caryota mitis Lour.	In this study
376	Astrosphaeriella fronsicola J. Fröhl. & K.D. Hyde	Melanommataceae	Wallichia siamensis Becc.	<i>Mycol. Res.</i> 99 (4): 453 (1995)
345	Astrosphaeriella nypae K.D. Hyde	Melanommataceae	Latania lontaroides (Gaertn.) H.E.Moore	J. Linn. Soc., Bot. 110: 96 (1992)
084	Astrosphaeriella palmicola Hidayat & To-anun, sp. nov.	Melanommataceae	Areca catechu L.	In this study

Herbaria number (FIH)	Fungal species	Family	Host	References
377	Beltrania rhombica Penz.	Incertae sedis	Wallichia siamensis Becc.	Michelia 2 (no. 8): 474 (1882)
378	Beltraniella portoricensis (F. Stevens) Piroz. & S.D. Patil	Hyponectriaceae	Wallichia siamensis Becc.	Can. J. Bot. 48(3): 575 (1970)
043	Bionectria ochroleuca (Schwein.) Schroers & Samuels	Bionectriaceae	Borassus flabellifer L.	Z. Mykol. 63 (2): 151 (1997)
037	Bionectria sp.1	Bionectriaceae	Cocos nucifera L.	
414	Bionectria sp.2	Bionectriaceae	Chamaedorea costaricana Oerst.	
379	Bionectria sp.3	Bionectriaceae	Wallichia siamensis Becc.	500
064; 103	Botryosphaeria cocogena Subileau, Renard & Lacoste	Botryosphaeriaceae	Cocos nucifera L.; Areca catechu L.	Mycotaxon 51: 8 (1994)
020	Botryosphaeria obtusa (Schwein.) Shoemaker	Botryosphaeriaceae	Wallichia siamensis Becc.	Canadian Journal of Botany 42 : 1298 (1964)
004	Byssosphaeria schiedermayeriana (Fuckel) Barr	Melanommataceae	Cocos nucifera L.	Mycotaxon 20: 34 (1984)
187	Botryotrichum sp.	Chaetomiaceae	Wallichia siamensis Becc.	
380	Camposporium antennatum Harkn.	Incertae sedis	Wallichia siamensis Becc.	Bull. Calif. Acad. Sci. 1: 37 (1884)
322	Catabotrys deciduum (Berk. & Broome) Seaver & Waterston	Catabotrydaceae	Areca catechu L.	Mycologia 38: 184 (1946)
330	Cercospora arecacearum Hidayat and Meeboon, sp. nov.	Mycosphaerelaceae	Areca catechu L.	<i>Mycol. Prog.</i> 8 : 116 (2009)
381	Ceuthospora palmicola Joanne E. Taylor, K.D. Hyde & E.B.G. Jones	Phacidiaceae	Wallichia siamensis Becc.	Fungal Diversity Res. Ser. 12: 250 (2003)
182	Chaetomium sp.	Chaetomiaceae	Wallichia siamensis Becc.	

Herbaria number (FIH)	Fungal species	Family	Host	References
029	Chaetospermum chaetosporum (Pat.) Smith & Ramsbottom	Incertae sedis	Wallichia siamensis Becc.	Trans. Br. Mycol. Soc. 4: 328 (1914)
021	Chaetosphaeria fusiformis W. Gams & HolJech.	Chaetosphaeriaceae	Wallichia siamensis Becc.	Mycotaxon 13: 257 (1981)
382	Chalara cylindrosperma (Corda) S. Hughes	Incertae sedis	Wallichia siamensis Becc.	Can. J. Bot. 36: 747 (1958)
383	Cladosporium cladosporioides (Fresen.) G.A. de Vries	Davidiellaceae	Wallichia siamensis Becc.	Contrib. Knowledge of the Genus Cladosporium Link ex Fries: 57 (1952)
404	Cladosporium oxysporum Berk. & M.A. Curtis	Davidiellaceae	Chamaedorea metallica O.F.Cook ex H.E.Moore	J. Linn. Soc., Bot. 10 : 362 (1868)
164; 100	Colletotrichum acutatum J.H. Simmonds	Glomerellacea	Borassus flabellifer L.; Borassodendron sp.	Queensland J. agric. Anim. Sci. 25: 178A (1968)
038, 122, 139, 145; 092, 093, 096; 401	Colletotrichum gloeosporioides (Penz.) Penz. & Sacc.	Glomerellaceae	Wallichia siamensis Becc.; Areca catechu L.; Chamaedorea metallica O.F.Cook ex H.E.Moore	Sér. 6 2 : 670 (1884)
384	Conioscypha lignicola Höhn.	Incertae sedis	Wallichia siamensis Becc.	Annales Mycologici 2 : 58 (1904)
407, 413, 417	Curvularia affinis Boedijn	Pleosporaceae	Bismarckia nobilis Hildebr. & H.Wendl.	Bull. Jard. Bot. Buitenz, 3 Sér. 13: 130 (1933)
099; 160	Curvularia lunata (Wakker) Boedijn	Pleosporaceae	Borassodendron sp.; Wallichia siamensis Becc.	Bull. Jard. Bot. Buitenz, 3 Sér. 13: 127 (1933)
161	Curvularia senegalensis (Speg.) Subram.	Pleosporaceae	Borassus flabellifer L.	J. Indian Bot. Soc. 35 : 467 (1956)
106, 112	Cylindrosporium sp.	Incertae sedis	Borassodendron	
157	Dactylaria sp.	Incertae sedis	sp. Wallichia siamensis Becc.	
334	Diatrype chlorosarca Berk. & Broome	Diatrypaceae	Wallichia siamensis Becc.	J. Linn. Soc., Bot. 14(2): 123 (1875)

Herbaria number (FIH)	Fungal species	Family	Host	References
385	Diaporthe palmarum Joanne E. Taylor, K.D. Hyde & E.B.G. Jones	Diaporthaceae	Wallichia siamensis Becc.	Fungal Diversity Res. Ser. 12: 207 (2003)
386	Diatrype chlorosarca Berk. & Broome	Diatrypaceae	Wallichia siamensis Becc.	J. Linn. Soc., Bot. 14: 123 (1875)
154	Dictyochaeta wallichianensis Hidayat & To-anun	Chaetosphaeriaceae	Wallichia siamensis Becc.	In this study (chapter 5)
387	Dictyochaetopsis apicalis (Berk. & M.A. Curtis) Aramb. & Cabello	Chaetosphaeriacea	Wallichia siamensis Becc.	Mycotaxon 38: 12 (1990)
388	Dictyosporium heptasporum (Garov.) Damon	Incertae sedis	Wallichia siamensis Becc.	Lloydia 15: 118 (1952)
046; 088	Didymella sp.	Incertae sedis	Borassus flabellifer L.; Areca catechu L.	
389	Didymosphaeria calamicola Aptroot, J. Fröhl. & K.D. Hyde	Didymosphaeriaceae	Wallichia siamensis Becc.	Nova Hedwigia 69 : 453 (1999)
390	Dinemasporium graminum Lév.	Incertae sedis	Wallichia siamensis Becc.	Annls Sci. Nat., Bot., sér. 3, 5 : 274 (1846)
057	Diplodia sp.	Botryosphaeriaceae	Borassus flabellifer L.	
391	Ellisembia sp.	Incertae sedis	Wallichia siamensis Becc.	
149	Eriosporella calami (Niessl) Höhn.	Incertae sedis	Wallichia siamensis Becc.	Sber. Akad. Wiss. Wien, Mathnaturw. Kl., Abt. 1, 125 : 109 (1916)
420	Exosporium stilbaceum (Moreau) M.B. Ellis	Incertae sedis	Licuala peltata Roxb.	<i>Mycol. Pap.</i> 82 : 38 (1961)
147	Fasciatispora petrakii (Mhaskar & V.G. Rao) K.D. Hyde	Incertae sedis	Wallichia siamensis Becc.	Nova Hedwigia 61 : 255 (1995)
214	Fasciatispora ujungkulonensis Hidayat	Incertae sedis	Caryota sp.	Mycotaxon 102: 350 (2007)
405	Fusarium sansainensis Hidayat & To-anun, sp. nov.	Nectriaceae	Chamaedorea metallica O.F.Cook ex H.E.Moore	In this study (chapter 4)

Herbaria number (FIH)	Fungal species	Family	Host	References
152	Gliocladium penicillioides Corda	Hyphocreaceae	Wallichia siamensis Becc.	Icon. Fung. 4: 31 (1840)
050, 144; 418	Glomerella cingulata (Stoneman) Spald. & H. Schrenk	Glomerellaceae	Wallichia siamensis Becc.; Livistona fulva Rodd	Science, N. S. 17: 751 (1903)
407; 327	Guignardia arengae, Hidayat & To-anun, sp. nov.	Botryosphaeriaceae	Arenga hookeriana (Becc.) Whitmore; Livistona chinensis R.Br.	In this study
075, 076; 098	Guignardia calami (Syd.) Arx & E. Müll.	Botryosphaeriaceae	Areca catechu L.; Borassodendro n sp.	Beitr. Kryptfl. Schweiz 11: 55 (1954)
008, 062; 110	Guignardia cocöes (Petch) K.D. Hyde	Botryosphaeriaceae	Cocos nucifera L.; Borassodendro n sp.	Sydowia 47 : 188 (1995)
016, 314	Guignardia uniappendiculatum Hidayat & To-anun, sp. nov.	Botryosphaeriaceae	Wallichia siamensis Becc.	In this study
392	Gyrothrix circinata (Berk. & M.A. Curtis) S. Hughes	Incertae sedis	Wallichia siamensis Becc.	Can. J. Bot. 36: 771 (1958)
393	Helicomyces lilliputeus R.T. Moore	Tubeufiaceae	Wallichia siamensis Becc.	<i>Mycologia</i> 49 : 583 (1957)
086; 394	Hermatomyces tucumanensis Speg.	Incertae sedis	Areca catechu L.; Wallichia siamensis Becc.	Anal. Mus. nac. Hist. nat. B. Aires 13: 446 (1911)
048	Lacellina graminicola (Berk. & Broome) Petch	Incertae sedis	Borassus flabellifer L.	Ann. R. bot. Gdns Peradeniya 9 : 171 (1924)
070	Lacellinopsis sacchari Subram.	Incertae sedis	Borassus flabellifer L.	Proc. natn. Acad. Sci. India, Sect. B, Biol. Sci. 37: 104 (1953)
006; 077, 095	Lachnum palmae (Kanouse) Spooner	Hyaloscyphaceae	Wallichia siamensis Becc.; Areca catechu L.	Biblthca. Mycol. 116: 484 (1987)
058	Lachnum sp.	Hyaloscyphaceae	Cocos nucifera L.	

Herbaria number (FIH)	Fungal species	Family	Host	References
063; 395	Lasiodiplodia theobromae (Pat.) Griffon & Maubl.	Botryosphaeriaceae	Cocos nucifera L.; Wallichia siamensis Becc.	Bull. Soc. mycol. Fr. 25 : 57 (1909)
291	Lasiosphaeria sp	Lasiosphaeriaceae	Caryota mitis Lour.	
091	Lepteutypa sabalicola (Ellis & G. Martin) M.E. Barr	Amphisphaeriaceae	Areca catechu L.	Mycotaxon 46 : 57 (1993)
181	Leptodothiorella sp.	Botryosphaeriaceae	Wallichia siamensis Becc.	2
083, 087	Linocarpon nonappendiculatum Hidayat & To-anun, sp. nov.	Incertae sedis	Areca catechu L	In this study
323	Lophiostoma livistonicola Hidayat & To-anun, sp. nov.	Lophiostomataceae	Livistona chinensis R.Br.	In this study
396	Lophiostoma macrostomum (Tode) Ces. & De Not.	Lophiostomataceae	Wallichia siamensis Becc.	Symbolae Mycologicae: 7 (1870)
035	Lophodermium arundinaceum (Schrad.) Chevall.	Rhytismataceae	Cocos nucifera L.	Fl. gén. env. Paris 1: 435 (1826)
397	Massarina palmicola K.D. Hyde & Aptroot	Massarinaceae	Wallichia siamensis Becc.	Nova Hedwigia 64 : 499 (1997)
415; 344	Melanographium citri (Gonz. Frag. & Cif.) M.B. Ellis	Incertae sedis	Copernicia rigida Britton & P.Wilson; Latania lontaroides (Gaertn.) H.E.Moore	Mycol. Pap. 93 : 21 (1963)
053, 068	Microthyrium elatum Rehm	Microthyriaceae	Borassus flabellifer L.	Saccardo's Syll. fung. XXIV: 426 (1926)
009	Microthyrium fagi J.P. Ellis	Microthyriaceae	Cocos nucifera L.	Trans. Br. mycol. Soc. 67 : 383 (1977) [1976]
188	Mitteriella zizyphina Syd.	Englerulaceae	Wallichia siamensis Becc.	Annales Mycologici 31: 95 (1933)
398	Monodictys putredinis (Wallr.) S. Hughes	Incertae sedis	Wallichia siamensis Becc.	Can. J. Bot. 36 : 785 (1958)

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399	Montagnula sp.	Montagnulaceae	Wallichia siamensis Becc.	
357	Morenoina palmicola J. Fröhl., K.D. Hyde & Joanne E. Taylor	Asterinaceae	Wallichia siamensis Becc.	Fungal Diversity Research Series 3: 89 (2000)
156	Mucor sp.	Mucoraceae (Zygomycota)	Wallichia siamensis Becc.	
271	Muyocopron tectum (G. Winter) Arx & E. Müll.	Microthyriaceae	Borassus flabellifer L.	Beiträge zur Kryptogamenflora der Schweiz 11 : 91 (1954)
102	Mycosphaerella arecacearum Hidayat & To-anun, sp. nov.	Mycosphaerellaceae	Areca catechu L.	In this study
059	Mycosphaerella borassi Hidayat and To-anun, sp. nov.	Mycosphaerellaceae	Borassus flabellifer L.	In this study
015, 030; 089	Mycosphaerella palmicola Chaudhury & P.N. Rao	Mycosphaerellaceae	Cocos nucifera L.; Areca catechu L.	
326	Mycosphaerella sp.	Mycosphaerellaceae	Livistona chinensis R.Br.	
013	Mycosphaerella wallichiae Hidayat and To-anun, sp. nov.	Mycosphaerellaceae	Wallichia siamensis Becc.	In this study
007; 025; 108	Myelosperma tumidum Syd. & P. Syd.	Myelospermataceae	Cocos nucifera L.; Wallichia siamensis Becc.; Borassodendron sp.	Mycologici 13: 38
097	Myrothecium sp.	Incertae sedis	Borassodendron sp.	OOTHE
003	Nectria foliicola Berk. & M.A. Curtis	Nectriaceae	Cocos nucifera L.	J. Linn. Soc., Bot. 10 : 379 (1868)
111	Nectria sp.1	Nectriaceae	Borassodendron sp.	rvo
116	Nectria sp.2	Nectriaceae	Caryota mitis Lour.	. , ,
126, 354	Neorehmia arecae (Syd.) Samuels & M.E. Barr	Trichosphaeriaceae	Wallichia siamensis Becc.	Canadian Journal of Botany 75 : 2165-2176 (1997)

Herbaria number (FIH)	Fungal species	Family	Host	References
047; 400	Nigrospora oryzae (Berk. & Broome) Petch	Incertae sedis	Borassus flabellifer L.; Wallichia siamensis Becc.	J. Indian bot. Soc. 4: 24 (1924)
412	Nigrospora sphaerica (Sacc.) E.W. Mason	Incertae sedis	Bismarckia nobilis Hildebr. & H.Wendl.	Trans. Br. mycol. Soc. 12: 158 (1927)
159	Nodulisporium acervatum (Massee) Deighton	Xylariaceae	Wallichia siamensis Becc.	<i>Trans. Br. mycol. Soc.</i> 85 (3): 391 (1985)
127	Oedocephalum formosus Hidayat & To-anun, sp. nov.	Incertae sedis	Borassus flabellifer L.	In this study
338	Ophioceras dolichostomum (Berk. & M.A. Curtis) Sacc.	Magnaporthaceae	Rhapis sp.	Syll. fung. (Abellini) 2: 358 (1883)
078, 094	Ophioceras guttulatum K.M. Tsui, H.Y.M. Leung, K.D. Hyde & Hodgkiss	Magnaporthaceae	Areca catechu L.	Mycoscience 42 (4): 321 (2001)
011; 034	Ophioceras tenuisporum Shearer, J.L. Crane & W. Chen	Magnaphortaceae	Wallichia siamensis Becc.; Cocos nucifera L.	Mycologia 91 : 149 (1999)
151	Oxydothis cyrtostachicola Hidayat & To-anun	Incertae sedis	Cyrtostachys renda Blume	Fungal Diversity 23 : 164 (2006)
019, 141	Oxydothis daemonoropsicola J. Fröhl. & K.D. Hyde	Incertae sedis	Wallichia siamensis Becc.	Fungal Diversity Research Series 3: 183(2000)
018		Incertae sedis	Wallichia siamensis Becc.	Fungal Diversity 23: 165 (2006)
119/18	Oxydothis linospadicis J. Fröhl. & K.D. Hyde	Incertae sedis	Caryota mitis Lour.	<i>Mycol. Res.</i> 98 : 215 (1994)
001	Oxydothis oedema (Mont.) K.D. Hyde	Incertae sedis	Cocos nucifera L.	<i>Sydowia</i> 46 : 300 (1994)
419	Oxydothis oraniopsis J. Fröhl. & K.D. Hyde	Incertae sedis	Chrysalidocar pus lutescens H.Wendl.	Mycol. Res. 98 : 215 (1994)
114	Oxydothis parvula (Syd.) Petr.	Incertae sedis	Borassodendro n sp.	<i>Sydowia</i> 6 : 314 (1952)

Herbaria number (FIH)	Fungal species	Family	Host	References
010	Oxydothis wallichianensis Hidayat, To-Anun & K.D. Hyde	Incertae sedis	Wallichia siamensis Becc.	Fungal Diversity 23: 167 (2006)
184	Paecilomyces variotii Bainier	Trichocomaceae	Wallichia siamensis Becc.	Bull. Soc. Mycol. Fr. 23: 27 (1907)
162	Passalora clematidis (R.K. Verma & Kamal) U. Braun & Crou	Mycosphaerellaceae	Borassus flabellifer L.	CBS Diversity Ser. 1: 448 (2003)
005	Pemphidium rattanicola J.Fröhl. & K.D. Hyde	Incertae sedis	Wallichia siamensis Becc.	Fungal Diversity Research Series 3: 225 (2000)
153	Penicillium sp.	Trichocomaceae	Wallichia siamensis Becc.	53
039, 061; 331	Periconia byssoides Pers.	Incertae sedis	Borassus flabellifer L.; Areca catechu L.	Syn. meth. fung. 1: 18 (1801)
069, 072; 417	Periconia cookei E.W. Mason & M.B. Ellis	Incertae sedis	Borassus flabellifer L.; Copernicia rigida Britton & P.Wilson	<i>Mycol. Pap.</i> 56 : 72 (1953)
411; 402	Periconia digitata (Cooke) Sacc.	Incertae sedis	Bismarckia nobilis Hildebr. & H.Wendl.; Chamaedorea metallica O.F.Cook ex H.E.Moore	Syll. fung. (Abellini) 4: 274 (1886)
190	Periconiella cocoës M.B. Ellis	Mycosphaerellaceae	Wallichia siamensis Becc.	Mycol. Pap. 111: 23 (1967)
027) Y 1 8	Pestalosphaeria elaeidis (C. Booth & J.S. Robertson) Aa	Amphisphaeriaceae	Wallichia siamensis Becc.	Proc. K. Ned. Akad. Wet., Ser.C, Biol. Med. Sci. 67 : 87 (1976)
319; 342	Pestalotiopsis eusora (Sacc.) J. Xiang Zhang & T. Xu	Amphisphaeriaceae	Caryota mitis Lour.; Pinanga sp.	Mycotaxon 85 : 93 (2003)

Herbaria number (FIH)	Fungal species	Family	Host	References
032	Pestalotiopsis guepinii (Desm.) Steyaert	Amphisphaeriaceae	Wallichia siamensis Becc.	Bulletin du Jardin Botanique de l'État à Bruxelles 19: 312 (1949)
054; 060; 410; 406; 421; 134	Pestalotiopsis palmarum (Cooke) Steyaert	Amphisphaeriaceae	Borassus flabellifer L.; Cocos nucifera L.; Bismarckia nobilisHildebr. & H.Wendl.; Chamaedorea metallica O.F.Cook ex H.E.Moore; Licuala peltata Roxb.; Livistona chinensis R.Br.	Bull. Jard. Bot. État Brux. 19: 322 (1949)
359	Petrakia echinata (Peglion) Syd. & P. Syd.	Incertae sedis	Wallichia siamensis Becc.	Annales Mycologici 11: 407 (1913)
328	Phaeosphaeria livistonae J. Fröhl. & K.D. Hyde	Phaeosphaeriaceae	Livistona chinensis R.Br.	Fungal Diversity Research Series 3: 273 (2000)
272	Phaeophleospora striae Joanne E. Taylor, K.D. Hyde & E.B.G. Jones	Mycosphaerellaceae	Borassus flabellifer L.	Fungal Diversity Res. Ser. 12: 276 (2003)
045	Phoma sp.1	Incertae sedis	Cocos nucifera L.	
360	Phoma sp.2	Incertae sedis	Wallichia siamensis Becc.	
113, 315	Phomopsis caryotae-urentis Petr. & Cif.	Diaporthaceae	Wallichia siamensis Becc.; Caryota mitis Lour.	Annales Mycologici 28: 413 (1930)
105; 409, 336	Phomopsis elaeidis Punith.	Diaporthaceae	Borassodendron sp.; Arenga hookeriana (Becc.) Whitmore; Borassus flabellifer L.	Trans. Br. Mycol. Soc. 63 : 229 (1974)
131	Phomopsis pittospori S.A. Archer	Diaporthaceae	Livistona chinensis R.Br.	Trans. Br. mycol. Soc. 61 : 221 (1973)

Herbaria number (FIH)	Fungal species	Family	Host	References
317, 321	Phyllosticta caryotae C.I. Chen	Botryosphaeriaceae	Caryota mitis Lour.	Contrib. Biol. Lab. Sci. Soc. China, Bot. Ser. 8: 157 (1932)
038; 403	Pithomyces sacchari (Speg.) M.B. Ellis	Pleosporaceae	Borassus flabellifer L.; Chamaedorea metallica O.F.Cook ex H.E.Moore	Mycol. Pap. 76 : 17 (1960)
040	Pleiochaeta setosa (Kirchn.) S. Hughes	Incertae sedis	Cocos nucifera L.	<i>Mycol. Pap.</i> 36 : 39 (1951)
265	Pleospora herbarum (Pers.) Rabenh.	Pleosporaceae	Borassus flabellifer L.	Herb. myc. 2 : 547 (1854)
361	Pleurophragmium sp.	Incertae sedis	Wallichia siamensis Becc.	
002; 362	Protocreopsis pertusa (Pat.) Samuels & Rossman	Bionectriaceae	Cocos nucifera L.; Wallichia siamensis Becc.	Stud. Mycol. 42 : 66 (1999)
049	Pseudogibellula formicarum (Mains) Samson & H.C. Evans	Cordycipitaceae	Borassus flabellifer L.	Acta bot. neerl. 22: 524 (1973)
142	Pseudohalonectria palmicola K. D. Hyde, Joanne E. Taylor & J. Fröhl.	Magnaporthaceae	Wallichia siamensis Becc.	<i>Mycologia</i> 91 : 522 (1999)
041	Rabenhorstia tiliae (Fr.) Fr.	Incertae sedis	Borassus flabellifer L.	Summa veg. Scand., Section Post. (Stockholm): 410 (1849)
140	Rachidicola obclavatum Hidayat	Incertae sedis	Wallichia siamensis	In this study
186	& To-anun, sp. nov. Rhizopus sp.	Mucoraceae (Zygomycota)	Becc. Wallichia siamensis Becc.	BOTH
026	Rosellinia victoriae Syd. & P. Syd.	Xylariaceae	Wallichia siamensis Becc.	Annales Mycologici 6: 483 (1908)
363	Roussoëlla palmicola J. Fröhl., K.D. Hyde & Aptroot	Didymosphaeriaceae	Wallichia siamensis Becc.	Nova Hedwigia 69 : 468 (1999)
042	Schizophyllum fasciatum Pat.	Schizophyllaceae (Basidiomycota)	Borassus flabellifer L.	J. Bot. Morot 1: 170 (1887)

Herbaria number (FIH)	Fungal species	Family	Host	References
268	Spegazzinia deightonii (S. Hughes) Subram.	มยนติ	Borassus flabellifer L.	J. Indian bot. Soc. 35 : 78 (1956)
364	Spiropes penicillium (Speg.) M.B. Ellis	Incertae sedis	Wallichia siamensis Becc.	<i>Mycol. Pap.</i> 114 : 23 (1968)
365	Sporidesmium sp.	Incertae sedis	Wallichia siamensis Becc.	
189	Sporoschisma saccardoi E.W. Mason & S. Hughes	Chaetosphaeriaceae	Wallichia siamensis Becc.	Mycol. Pap. 31 : 20 (1949)
368	Sporormiella minimoides S.I. Ahmed & Cain	Sporormiaceae	Wallichia siamensis Becc.	Can. J. Bot. 50 : 450 (1972)
197	Stachybotrys kampalensis Hansf.	Incertae sedis	Wallichia siamensis Becc.	Proc. Linn. Soc. London 155: 45 (1943) 45
169	Stilbella sp.	Incertae sedis	Borassus flabellifer L.	
017	Submersisphaeria suthepensis Hidayat & To-anun, sp. nov.	Annulatascaceae	Wallichia siamensis Becc.	In this study
150	Terriera brevis (Berk.) P.R. Johnst.	Rhytismataceae	Wallichia siamensis Becc.	<i>Mycol. Pap.</i> 176 : 98 (2001)
133	Terriera livistonae Hidayat & To-anun, sp. nov.	Rhytismataceae	Livistona chinensis R.Br.	In this study
085	Tetraploa aristata Berk. & Broome	Massarinaceae	Areca catechu L.	Ann. Mag. nat. Hist., Ser. 2 5: 459 (1850)
071; 367	Torula herbarum (Pers.) Link	Incertae sedis	Borassus flabellifer L.; Wallichia siamensis Becc.	Magazin Ges. naturf. Freunde, Berlin 3 : 19 (1809)
370	Trichothyrina alpestris (Sacc.) Petr.	Microthyriaceae	Wallichia siamensis Becc.	Sydowia 4 : 168 (1950)
082	Venturia frondicola Hidayat & To-anun, sp. nov.	Venturiaceae	Areca catechi L.	<i>u</i> In this study
343	Veronaea botryosa Cif. & Montemart.	Incertae sedis	Wallichia siamensis Becc.	Atti Ist. bot. Univ. Lab. crittog. Pavia, Sér. 5 15 : 68 (1957)

Herbaria number (FIH)	Fungal species	Family	Host	References
374	Verticillium sp.	Plectosphaerellaceae	Wallichia siamensis Becc.	
054; 372	Wiesneriomyces laurinus (Tassi) P.M. Kirk	Incertae sedis	Borassus flabellifer L.; Wallichia siamensis Becc.	Trans. Br. mycol. Soc. 82 : 748 (1984)
408; 416	Zygosporium echinosporum Bunting & E.W. Mason	Incertae sedis	Arenga hookeriana (Becc.) Whitmore; Copernicia rigida Britton & P.Wilson	Mycol. Pap. 5: 135 (1941)
051	Zygosporium gibbum (Sacc., M. Rousseau & E. Bommer) S. Hughes	Incertae sedis	Borassus flabellifer L.	Can. J. Bot. 36: 825 (1958)
333; 158	Zygosporium oscheoides Mont.	Incertae sedis	Caryota mitis Lour.; Wallichia siamensis Becc.	Annls Sci. Nat., Bot., sér. 2, 17 : 121 (1842)

2.2. Materials and Methods

A. Collecting Protocols and Site Description

Specimens of palm's necrotic leaflets and decaying fronds were collected from several natural and planted sites in Chiang Mai province and other locations, including Indonesia. Collections of *W.* siamensis fronds were carried out at Huay Kog Ma, Doi Suthep-Pui National Park, Chiang Mai, Thailand. The National Park is a typical tropical forest in northern Thailand as strongly seasonal. There are three distinct seasons, cool dry season (November-March), hot dry season (March-May), and warm wet season (May-November). The annual rainfall varies from 1100-1500

mm, with over 80% of annual rainfall within the 6 months of rainy season, whilst the months of December, January, and February are virtually without rain. The average temperature of the area ranged from 20°-23°C. In the dry season (December-February), the temperature at high elevation is low (Gardner *et al.*, 2000).

The amount of material collected at each site is standardized for each palm species depending on the bulkiness of the material. One to three of 11.5" x 16.5" resealable bags were used for each palm species. The different palm parts were collected in various states of decay. Living leaves with spots were collected if present. Collecting bags are sealed and labeled with the following information: *Name of the palm, Collecting site, Collector/s*, and *Date*. On returning to the laboratory, the material is incubated for a week and either studied immediately or air dried and stored for studying at a later date. Air drying enabled single spore isolations to be made, which would not be possible with oven dried specimens.

B. Examination of Materials

The decaying and senescent materials were examined for saprobic Ascomycetes, Coelomycetes, Hyphomycetes, and Basidiomycetes. The living leaves materials with necrosis symptoms were subjected for the observation of potential plant pathogenic fungi occur on the leaves symptom. The materials were examined using an Olympus SZ H10 dissecting microscope to determine the presence of the fungal fruiting structures. Once a group of fruiting structures is encountered, a marker is placed firmly in the material, or the area is marked with permanent pen. Each newly encountered fungus at each site is given an identity number and detail records are

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made, as outlined below. Once fully examined, the piece of material with all of its markers is placed in a resealable envelope with the following information:

Herbarium number

Fungus name

Host name

Collection site

Collector/s

Date

The materials were stored in labeled boxes in a dry environment, and naphthalene balls were used to keep insect infestation at a minimum. Unused materials were destroyed. Dried herbarium specimens were deposited at Mushroom Research Center Herbarium (MRC), CMU Herbarium (CMU), Faculty of Science, Chiang Mai University, Chiang Mai, Thailand, and Laboratory of Plant Pathology, Department of Plant Pathology, Faculty of Agriculture, Chiang Mai University.

Ascomycetes and Coelomycetes

A sharp one sided razor blade or a pair of Inox 5 fine forceps were used to carefully remove the top of the fruiting body. The specimens were rehydrated if the contents are dry or crystalline by using distilled water or potassium hydroxide (KOH) 3% before extraction. The contents were then placed in a drop of distilled water on a slide and covered with an 18 x 18 mm coverslip. The specimens were observed using an Olympus BX 50 photomicroscope system with differential interference contrast microscopy. Water was used for all examinations, spore measurements, and most of the photographs/line drawings.

Specific reagents were used when necessary as follows (Hawksworth *et al.*, 1995): Melzer's reagent was used to investigate any amyloid reactions at the apical ring of ascus of Ascomycetes. Indian ink (10%) was used to highlight the presence of mucilaginous sheaths of spores. Lactophenol cotton blue was used to highlight the ascus wall layers of bitunicate Ascomycetes. Potassium hydroxide (3%) and lactic acid was used in the identification of *Nectria* and allied genera. In addition, to aid the identification of the fungi, thin sections (± 10 µm) of rehydrated fruiting structures were made with a Micron HM505E cryostat microtome or by using hand section. Lactophenol was added to the slides for permanent fixation. The slides were heated to remove air bubbles in the lactophenol and the edges of the coverslip sealed with two layers of clear nail varnish. The slides were then labeled with the number of specimen it occurred on.

Hyphomycetes

Hyphomycetes materials examination was prepared by hand sectioning or by using a pair of Inox 5 fine forceps. Water was used as a medium for all examination. Lactophenol was used for permanent fixation. Detailed observations of morphological characters were carried out by means of an Olympus CX31 light microscope using oil immersion (1000Δ).

C. Single Spore Isolation

Single spore isolation of each encountered new fungus was referred to Choi *et al.* (1999) with a modification. A glass container or glass slide is sterilized with ethanol and wiped with a towel on which ethanol (70%) has been sprayed. A

sterilized pipette is then used to transfer a few drops of sterilized water into the glass container or onto the glass slide. The suspension is then prepared by mixing spores or conidia with a sterilized water on the glass container/glass slide. Sixteen squares or triangle shape are marked on the bottom of the water agar plate. The prepared homogenous spore suspension is then transferred with a sterilized pipette or loop, onto the surface of the water agar plate. A small drop of the suspension should be used at this stage to make a permanent slide and to check that the correct fungus has been selected.

The unsealed plate is incubated at 25°C for 12-24 hours. It is not sealed as this allows some of the surface water to dry out. If the plate is sealed with parafilm, water would accumulate on the surface of the agar and it would increase the chance of contamination. The spores/conidia are checked every 24 hours to establish germination. Once the spores/conidia have germinated, a sterilized glass needle is used to pick up a small piece of agar containing a spore/conidium. If the spores/conidia do not germinate after 24 hours, the plate is sealed with parafilm and periodically examined. About ten germinated spores/conidia are transferred and distributed evenly onto PDA plate and incubated at 25°C until their colony reached about 1 to 2 cm. A small piece of mycelium with agar can then be cut and transferred to another PDA plate and the culture is checked after few days, if there is no contamination, a pure culture has been obtained. Cultures can then be stored on the desired media.

There are many different groups of fungi, e.g. Ascomycetes, Coelomycetes, Basidiomycetes and Hyphomycetes and each has different types of fruiting bodies.

The methods to isolate each of them are therefore different. Fungi with closed fruiting

bodies (e.g. Ascomycetes with cleisthotechia, perithecia, and Coelomycetes with pycnidia) can be removed from the substrate surface using fine forceps and broken in sterilized water in order to provide spore/conidia suspension. If the fruiting body is submerged in the substrate, a razor blade is used to cut the fruiting body to expose the internal contents. The contents can then be transferred to a drop of sterilized water on a small glass container or slide in order to provide a spore/conidia suspension. Fungi with cup-shaped bodies (Ascomycetes with apothecia, Coelomycetes with acervuli) can be transferred directly, by removing the whole fruiting body. This can be placed is sterilized water and squashed with fine forceps, in order to provide a spore/conidia suspension. Basidiomycetes with gills can produce numerous spores. Spores suspension can be obtained by removing a few segments from the gills and agitating them in sterilized water. Isolation can also be carried out from sterile tissue within the cap. A few pieces of sterile tissue can be aseptically torn from the split fruiting structure and placed on water agar. Conidia of Hyphomycetes can be picked up directly from the substrate using fine forceps or a needle. It is better to pick up conidia only (not conidiophores), as this reduces the chance of contamination. The conidia are placed in sterilized water and agitated in order to provide conidia suspension.

D. Identification Procedures

Basic Identification

The following texts are consulted for basic identification:

(1) Ascomycetes: Müller and Arx (1962), Ainsworth *et al.* (1973), Arx and Müller (1954), Arx (1975; 1987), Wehmeyer (1975), Dennis (1981), Sivanesan (1984), Barr

(1987; 1990), Hanlin (1990, 1998a, 1998b), Fröhlich and Hyde (2000), Hyde *et al.* (2000), Taylor and Hyde (2003), and Cai *et al.* (2006).

- (2) Coelomycetes: Nag Raj (1993) and Sutton (1980).
- (3) Hyphomycetes: Ellis (1971, 1976), Carmichael *et al.* (1980), and Matsushima (1971, 1975, 1980, 1981, 1983, 1985, 1987, 1989, 1993, 1995, 1996).

In most cases the specimen could be identified to generic name with the above references. Further identification required examination of the relevant literatures. Sources are often suggested in the above references and the Dictionary of Fungi (Kirk *et al.*, 2008) is found particularly up to date and useful, as is the Bibliography of Systematic Mycology on CD ROM (Produced by I.M.I., C.A.B. International Wallingford, UK). The 'searchable' Database of Fungi on the internet is also invaluable. The following fungal databases websites are also useful in fungal identification:

(1) Index Fungorum

(http://www.indexfungorum.org/Names/Names.asp)

(2) USDA fungus-host database

(http://nt.ars-grin.gov/fungaldatabases/fungushost/fungushost.cfm)

(3) Home of the Xylariaceae

(http://mycology.sinica.edu.tw/Xylariaceae/)

(4) Fungi associated with Phragmites australis

(http://intramar.urgent.be/nemys/fungi/web/)

Description of New Taxa

In a case of the fungus could not be identified by consulting all of the relevant literature and seeking the advice of other mycological taxonomist, it is considered novel. Thorough description are made on standardized records sheets and usually 30 ascospores/conidia, 25 asci or conidiophores/conidiogenous cells and 10 fruiting bodies, apical apparatuses, ascospores sheats or appendages and paraphyses/pseudoparaphyses are measured. Microtome/hand sections are made of one or two fruiting bodies and 10 measurements are made of the whole ascomata or conidiomata and the components including the peridium, ostiole and where necessary the clypeus and/or stroma.

Presentation of Results

All fungi recorded in this study are presented in alphabetical order of the genera and subsequently species. "Host substrate" with information from the literature listed first and the relevant references cited, after which data from this study is listed. "Material examined" with information of country/ies, collecting site/s, host/s, dates, collector/s, and Herbarium/a number/s. For interesting species or genera, and new species or genera, a full description is presented, with the etymology of the specific epithet given and the holotype indicated in the "Material examined". Several unusual or rare species, or those which are poorly documented in the literature are illustrated and/or a description given.

Taxonomy

Fungi associated with Wallichia siamensis

Anthostomella limitata Sacc., Atti Accademia Scientifica VenetoTrentino-Istriana 4: 101. 1875

- = Anthostomella limitatum (Sacc.) Cooke, Grevillea 17: 90 (1889).
- = Anthostomella argentinensis (Speg.) Petr. and Syd., Annales Mycologici 23: 213 (1925).
- = Phaeophomatospora argentinensis Speg., Anales Museo Nacional Historia natural de Buenos Aires, Ser. 3, 12: 339 (1909).
- = Anthostomella gracilis Tassi, Bulletino del Laboratorio Ortor Botanico Siena 3: 53 (1900).
- = Anthostomella melanoderma Rehm, Österreichische Botanische Zeitschrift **54**: 82 (1904).

(Fig. 2.1)

Ascomata immersed, visible as blackened, raised, dome-shaped areas, dark-brown, coriaceous, clypeate, solitary, in section perithecium, globose to subglobose, 110.7-159.9 μm diam., 73.8-147.6 μm high. *Peridium* 12.5-20 μm wide, outside comprising 3-4 layers of brown-walled and compressed cells, inside comprising 1-2 layers of hyaline and compressed cells. *Paraphyses* septate, hyphal-like, up to 5 μm width. *Asci* cylindrical, 8-spored, unitunicate, ascal ring wedge shaped, J+ sub apical

ring, pedicellate, 85-112.5 \times 5-7.5 μ m. *Ascal ring* 1-1.5 μ m high, 3-4 μ m diam. *Ascospores* ellipsoid to ovoid to fusiform, 1-celled, 9-16 \times 4-5 μ m, dark brown, with a spiral germ slit (ca 5 μ m long) at the middle part of cell.

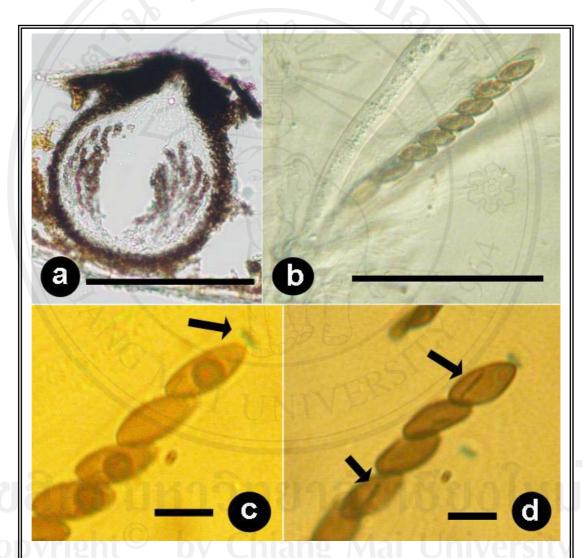


Fig. 2.1 Micrographs of *Anthostomella limitata* on *Wallichia siamensis*. a. Vertical section of ascoma. b. Ascus. c. Ascus ring (arrow). d. Ascospores with germ slit (arrows). Bars: $a = 125 \mu m$; $b = 100 \mu m$; $c, d = 5 \mu m$.

Material examined: THAILAND, Chiang Mai Province, Suthep-Pui National Park, on living leaflets of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat, (FIH 125) (CMU 27949).

Host: Callistemon, Carex, Conium, Daemonorops, Geranium, Galium, Kigelia, Rosa, Rubus, Sorbus, Typha (Lu and Hyde, 2000).

Distribution: Argentina, Azores, Brunei, Finland, Germany, India, Italy, UK (Lu and Hyde, 2000).

Notes: This species is characteristic of *Anthostomella limitata* as it has a J+ apical apparatus and ellipsoidal ascospores with a spiral germ slit, lacking a dwarf cell and being less than 15 μ m length. *Anthostomella limitata* on *W. siamensis* is only different from the type in having longer asci (85-112.5 × 5-7.5 μ m vs. 72.5-100 × 6.5-8 μ m) and longer ascospores (9-16 × 4-5 μ m vs. 9.5-12.5 × 4.5-5.5 μ m). This is the first report of *A. limitata* from Thailand and also *W. siamensis* as the host of the fungus.

Anthostomella puiggarii Speg., Anales de la Sociedad Cientifica Argentina 12: 106 (1881); non reliquiae Mycologicae Tropicae, Imprenta y Casa editora 'Coni' (1919)

= Anthostomella baileyi S. M. Francis, Transactions of the British Mycological Society 75: 201 (1980).

(Fig. 2.2)

Ascomata immersed, solitary, clypeate, visible as blackened, raised, shiny, dome-shaped areas, in section 156-252.5 μm diam., 125-205 μm high, globose to subglobose, perithecium, with a central ostiole. *Peridium* 10-12.5μm thick, outside comprising 2-3 layers of compressed brown and irregular cells, inside comprising 2-3 layers of compressed hyaline and ellipsoidal cells. *Paraphyses* 1.5-2 μm wide, numerous, filamentous, hyaline, septate, tapering to the apex, embedded in a gelatinous matrix. *Asci* 70-87.5 × 6-7.5 μm, unitunicate, 8-spored, cylindrical, short and tapering pedicellate, apically rounded, with a J+, discoid apical ring, apical ring size 1.5-2 μm wide, 1-1.75 μm high. *Ascospores* 5-6.5 × 7.5-12.5 μm, overlapping uniseriate, ellipsoidal, 1-celled, dark brown, thick-walled, with longitudinal germ slit through the spores, mucilaginous sheath lacking, often with oil drops structures near the ends.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on primary rachis of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 146); on leaflets of *Livistona* sp. (Arecaceae), 26 October 2006, Iman Hidayat (FIH 324).

Host: *Bambusa*, *Elaeis*, *Freycinetia*, *Livistona*, *Pandanus*, *Sararanga* (Liu and Hyde, 2000).

Distribution: Australia, Brazil, New Zealand, Nigeria, Philippines (Liu and Hyde, 2000).

Notes: *Anthostomella puiggarii* is commonly associated with monocotyledonous plants. This specimen is the first record of *A. puiggarii* in Thailand, and *W. siamensis* is reported here as a new host.

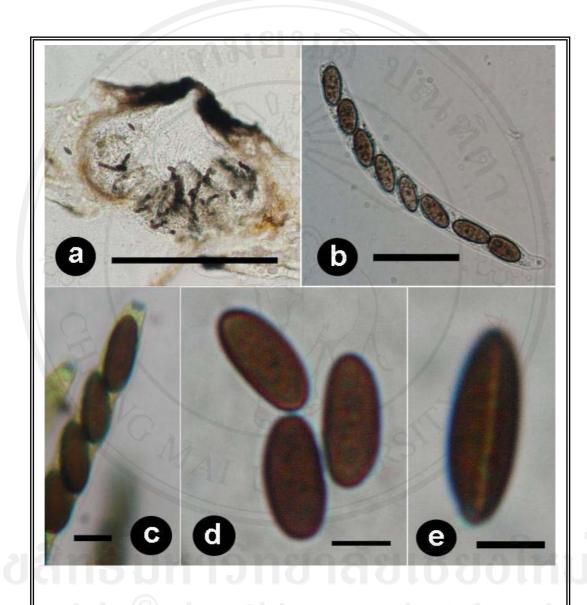


Fig. 2.2 Micrographs of *Anthostomella puiggarii* on *Wallichia siamensis*. a. Vertical section of ascoma. b. Ascus. c. J+ of ascus ring. d-e. Ascospores. Bars: $a = 200 \ \mu m$; $b = 30 \ \mu m$; $c, d, e = 10 \ \mu m$.

Apiospora siamicola Hidayat & To-anun, sp. nov.

(Fig. 2.3)

Macula epiphylla, irregulare, 2-13 mm diam. × 1-3 mm alta, brunneae cum marginum fuscae. Ascomata 200-237.5 μm diam. × 112-125 μm alta, subglobosa, medium ostiolatum, solitaria, immersa. Peridium 7-13.5 μm latum. Paraphysae admodum 5 μm latae, hyalinae, septatae. Asci 62.5-77.5 × 10-17.5 μm, 8-spori, unitunicati, clavati, apedicellati, apicis rotundis, apicale apparatus J+, discoidae, 0.5-1 μm high × 2.5-3 μm diam. Ascosporae apiosporae, 1-septatae, 20-25 × 5-6.3 μm, hyalinae, tunica gelatinosa praediti, basi cellulari amphori 1/5 collossi cellulari.

Etymology: The species name refers to Thailand (Siam) where the specimen was collected.

Leaf spots epiphyllous, irregular in shape, 2-13 mm diam., 1-3 mm high, brownish throughout with black margin, often overlaps among the spots, fruiting bodies 4-35 inside the margins, scattered. Ascomata visible as slightly raised, brown, ellipsoidal and irregular areas, with a central thin ring-like structure surrounding ostiole, in section 200-237.5 μm diam., 112.5-125 μm high, immersed, subglobose with flattened base. Peridium 7-13.5 μm wide, comprising 3-4 layers of compressed, brown-walled cells. Paraphyses hyaline, 3.8-5 μm wide, septate, hypha-like. Asci 62.5-77.5 × 10-17.5 μm ($\bar{x} = 72.6 \times 14$, n = 20), 8-spored, unitunicate, persistent, clavate, apedicellate, apically rounded, with a J+, 0.5-1 μm high × 2.5-3 μm diam.,

discoid, refractive subapical ring. *Ascospores* 20-25 × 5-6.3 μ m (\bar{x} = 21.7 × 2.2, n = 30), usually overlapping biseriate or occasionally multiseriate, apiosporous, 1-septate, hyaline, surrounded by thin persistent mucilaginous sheath, basal cell ca 1/5 the size of a larger cell.

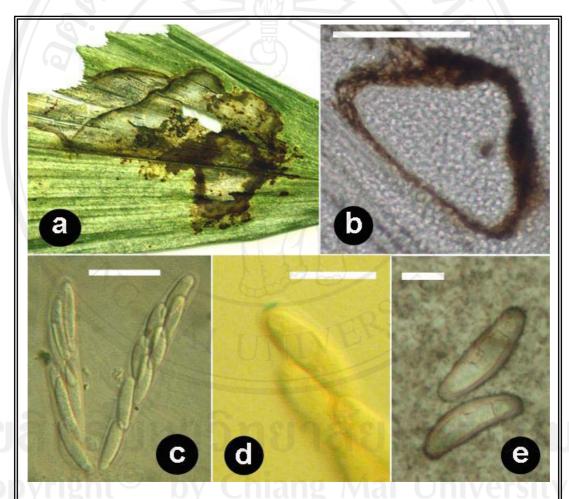


Fig. 2.3 Micrographs of *Apiospora siamicola* **sp. nov.** (from holotype). a. Symptom. b. Vertical section of ascoma. c. Asci. d. Ascus ring. e. Ascospores. Bars: $b = 100 \ \mu m$; $c = 20 \ \mu m$; $d = 15 \ \mu m$; $e = 5 \ \mu m$.

Material examined: THAILAND, Chiang Mai Province, Suthep-Pui National Park, Huay Kog Ma, on living leaflets of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 123) (Holotype: MRC 00001). Isotype: *ibid.*, 25 October 2006, Iman Hidayat and Jamjan Meeboon (FIH 353) (CMU 27948).

Host: On living leaflets of *W. siamensis*.

Distribution: This species is only known from the type locality.

Notes: By following the key to genera of ascomycetes with apiospores (Hyde *et al.*, 1998), this species is identified as *Apiospora* Sacc. based on ascomata being immersed with a slightly raised central ostiole, apedicellate and clavate asci, and hyaline ascospores.

Apiospora montagnei Sacc. (type species) and A. sinensis K. D. Hyde, J. Fröhl. & E. Taylor are the only species of Apiospora that recorded from palms. This species differs from A. montagnei and A. sinensis in having positive amyloid reaction of apical ring and lacking of stroma. In detail, this species differs from A. montagnei in having ascomata appearances on the host are scattered within black margin (ascomata appearances of A. montagnei are gregarious in linear rows), shorter asci (62.5-77.5 × 10-17.5 μm vs 75-120 × 14-18 μm of A. montagnei) and ascospores are surrounded by thin persistent mucilaginous sheath (in A. montagnei, ascospores lack of mucilaginous sheath). Furthermore, this species differs from A. sinensis in having shorter asci (62.5-77.5 × 10-17.5 μm vs 84-116 × (13-)18-25 μm of A. sinensis) and ascospores surrounded by thin persistent mucilaginous sheath (in A. sinensis, ascospores are surrounded by irregular and wide mucilaginous sheath.

The distinct character of *Apiospora siamicola* is the present of a positive amyloid reaction of the apical ring. The character is specific to *A. siamicola* as most

of the members of *Apiospora* are characterized with an amyloid negative apical ring (Hyde *et al.*, 1998). The amyloid positive apical ring can be confused with species of *Apioclypea* K. D. Hyde. However, in *Apioclypea*, the ascus is cylindrical and pedicellate, and the arrangement of ascospores in the asci is overlapping uniseriate (Hyde *et al.*, 1998).

Arecomyces foliicola Hidayat & To-anun, sp. nov.

(Fig. 2.4)

Macula epiphylla, apparui brunnei et irregulari, marginae fuscae. Ascomata 112.5-162.5 μm diam. × 100-137.5 μm alta, subglobosa, sepultum subtus pseudostromatum, parvus papillatus, medium ostiolatum. Peridium 4-12 μm latum. Paraphysae admodum 3 μm prolixae, filamentum. Asci 50-70 × 5-6.3 μm, 2(-4)-spori, unitunicati, lati cylindrici, pedicellati, ad apicem truncata, apparatu apicale J-, discoida praedita. Ascosporae 11-17 × 3-4.5 μm, uniseriatae, oblongus-ellipsoidus, aseptatae, hyalinae, laevae, tunica gelatinosa praedita.

Etymology: The species name refers to leaf (Latin: folium), a substrate where the fungus grew.

Leaf spot epiphyllous, visible as brown irregular area, darkened at the edges, fruiting bodies scattered throughout the brown area. Ascomata in section 112.5-162.5 μ m diam., 100-137.5 μ m high ($\bar{x} = 125 \mu$ m diam. × 106.3 μ m high, n = 10),

subglobose to horizontally ellipsoidal, immersed under pseudostroma, short papillate, ostiole central, slightly protruding through the leaf cuticle, visible as darkened irregular discs on the host surface, intraepidermal. *Peridium* 4-12 μ m wide, outer comprising 2-3 layers of compressed brown-walled cells, inner comprising 1-2 layers of compressed hyaline cells. *Paraphyses* up to 3 μ m wide, septate, hyaline, hyphalike, filamentous, wider distally, not embedded in gelatinous matrix, anastomosing between asci. *Asci* 50-70 \times 5-6.3 μ m ($\bar{x} = 59.4 \times 5.6$, n = 20), 2(-4)-spored, unitunicate, broadly cylindrical, long pedicellate, persistent, slightly truncate at the apex, broadest in the middle, with a J-, discoid, refractive sub apical ring. *Ascospores* 11-17 \times 3-4.5 μ m ($\bar{x} = 13.6 \times 3.8$, n = 30), uniseriate sometimes overlapping, oblong-ellipsoidal with rounded apices, aseptate, hyaline, smooth, and surrounded by thin persistent sheath.

Material examined: THAILAND, Chiang Mai Province, Suthep-Pui National Park, on living leaves of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 124) (Holotype: MRC 00003). Isotype: *ibid.*, 25 October 2006, Iman Hidayat & Jamjan Meeboon (FIH 354) (CMU 27947).

Host: On living leaflets of *W. siamensis*.

Distribution: This species is only known from the type locality.

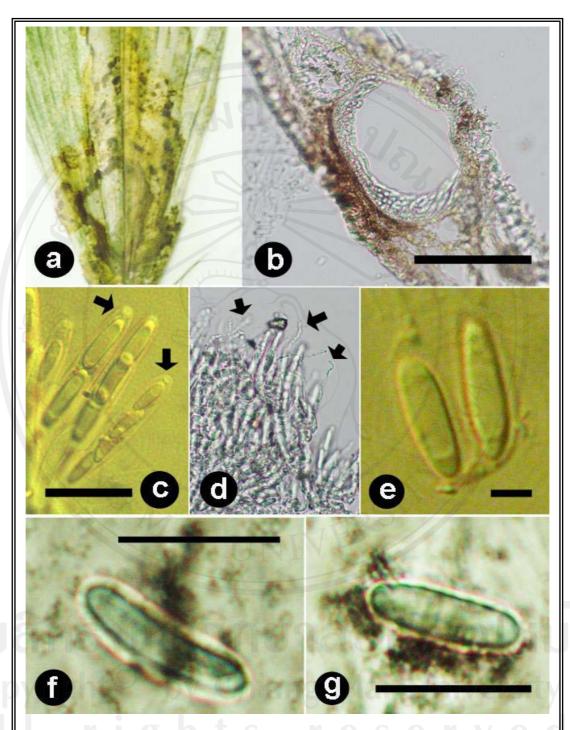


Fig. 2.4 Micrographs of *Arecomyces foliicola* **sp. nov.** (from holotype). a. Symptom. b. Vertical section of ascoma. c. Asci (ascus ring showed by arrows). d. Paraphyses (arrows). e-g. Ascospores. Bars: $a=125~\mu m$; $c=25~\mu m$; $e=2.5~\mu m$; $e=2.5~\mu m$; $e=2.5~\mu m$.

Notes: The general morphology of this species is most similar to *Arecomyces* K. D. Hyde and *Physalospora* Niessl, as ascomata are immersed, the apical ring is nonamyloid, and the ellipsoidal ascospores are hyaline (Hanlin, 1990; Hyde, 1996). However, this species differs from *Physalospora* in having ascomata immersed under pseudostroma, asci are broadly cylindrical with distinct apical ring and ascospores are less than 20 μ m (12-17 \times 3-4 μ m).

Arecomyces tetrasporus K. D. Hyde is the only member of Arecomyces having 2-4 ascospores in the ascus (Hyde, 1996). The species is characterized by ascomata immersed under clypeus; ellipsoidal and echinulose ascospores that surrounded by a wide mucilaginous sheath. However, A. foliicola differs to A. tetrasporus in having ascomata immersed under pseudostroma; smaller asci (48-64 \times 5-6 μ m vs 62-75 \times 8-12 μ m of A. tetrasporus); and the ascospores are smooth, oblong-ellipsoidal in shape, and surrounded by thin persistent sheath. The mode of life of A. foliicola is also weakly parasitic due to its association with leaf spot, but A. tetrasporus is an obligate saprobe.

Key to Arecomyces species (modified from Hyde, 1996)

1a. Asci (2-)4-spored 2

1b. Asci 8-spored 3

- 2a. Ascomata clypeate, ascospores ellipsoidal, on rachis A. tetrasporus
- 2b. Ascomata immersed under pseudostroma, ascospores oblong-ellipsoidal with rounded apices, associated with leaf spots *A. foliicola*

3a. Ascospores asymmetric, $12.5\text{-}15 \times 5.7.5 \, \mu\text{m}$, lacking a mucilaginous sheath, developing under a blackened pseudostroma *A. sekoyae*

3b. Ascospores symmetric 4

4a. Ascospores lenticular, $15-19 \times 6.5-8 \mu \text{m} \dots A$. bruneiensis

4b. Ascospores oblong ellipsoidal, lenticular or ovoid, mostly less than 15 μm long 5

5a. Ascospores mostly 12-15 μm long 6

5b. Ascospores mostly less than 12 μm long 7

6a. Asci 94-120 \times 11-15 μ m, ascospores sheath regular in outline A. frondicola

6b. Asci 82-92 × 7-8 μm, ascospores sheath wavy in outline A. epigeni

7a. Ascospores oblong-ellipsoidal, 7.5-10 × 4.5-6 μm, clypeate A. dicksonii

7b. Ascospores ovoid, $8.5\text{-}12.5 \times 5\text{-}6.5 \mu m$, developing under a reddish-brown pseudostroma *A. hedgerii*

ลบสทธมหาวทยาลยเชียงไหม Copyright[©] by Chiang Mai University All rights reserved Botryosphaeria obtusa (Schwein.) Shoemaker, Canadian Journal of Botany 42: 1298 (1964)

- = Sphaeria obtusa Schwein., Transactions of the American Philosophical Society Philadelphia N.S. 4: 220 (1832).
- Physalospora obtusa (Schwein.) Cooke, Grevillea 20: 86 (1892).(Fig. 2.5)

Stromata embedded within the cortical tissue, 510 mm wide, multilocular, scattered, solitary, botryose, dark brown to black, with ostioles darker around the neck region. *Pseudoparaphyses* 2.5-5 μm wide, hypha-like, septate, hyaline. *Asci* 140-195 × 10-15 μm, bitunicate, cylindric-clavate, long pedicellate, 8-spored. *Ascospores* 20-25 × 9.4-10 μm, irregularly biseriate, fusiform, wider around the mid region, aseptate, verrucose, hyaline.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on decaying rachis of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 020).

Host: Commonly diseased as well as saprobes on *Malus* spp. and *Vitis* spp. (Arx, 1987)

Distribution: Widespread in Europe, Southern Africa, North and South America, India, Japan, New Guinea, Western Australia, Tasmania, New Zealand (Arx, 1987).

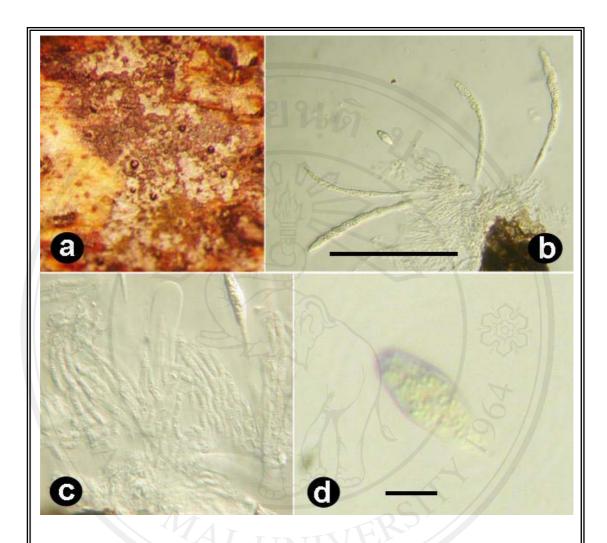


Fig. 2.5 Micrographs of *Botryosphaeria obtusa* on *Wallichia siamensis*. a. Appearances of multilocus ascomata with stroma on the host. b. Asci. c. pseudoparaphyses. d. Ascospore. Bars: $b=150~\mu m$; $d=10~\mu m$.

Notes: This species is identified as *Botryosphaeria obtusa* (Schwein.) Shoemaker due to its clavate asci with long pedicellate and fusiform ascospores (Shoemaker, 1964). This specimen is the first record of *B. obtusa* from Thailand and *W. siamensis* is reported here as a new host.

Chaetospermum chaetosporum (Pat.) Smith & Ramsbottom, Trans. Br. Mycol. Soc. 4: 328 (1914)

Tubercularia chaetospora Pat., Bulletin de la Société Mycologique de France
 4: 40 (1888).

(Fig. 2.6)

Conidiomata 300-600 μm diam., pycnidioid, globose, innate-erumpent under white and hairy stroma, initially closed, ultimately opening by an irregular split in the apical wall, gelatinous, unilocular. Conidiophores loosely aggregated, sparingly branched and septate at the base, colourless, smooth, invested in gel. Conidiogenous cells holoblastic, discrete, cylindrical to subcylindrical or irregular, colourless, smooth, bearing a single terminal conidium or an apical cluster of up to four conidia. Conidia 20-35 × 3-6.3 μm, broadly ellipsoidal to cylindrical with obtuse ends, unicellular, colourless, smooth, thin-walled, bearing appendages which are not separated from the conidium body by septa, polar or subpolar, occasionally lateral as well, unbranehed, filiform, or narrow and attenuated, flexuous, often collapsing and ribbon-like with age, 2.5-12 μm long.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on secondary rachis of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 029).

Host: Alnus glutinosa, Cupressus macrocarpa, Dahlia sp., Elaeis guineensis, Epilobium hirsutum, Glyceria maxima, Prunus serotina, Theobroma cacao, triticum sp., Valeriana officinalis (Sutton, 1980).

Distribution: India, Kenya, Tanzania, U.S.A., New Hebrides, U. K. (Sutton, 1980).

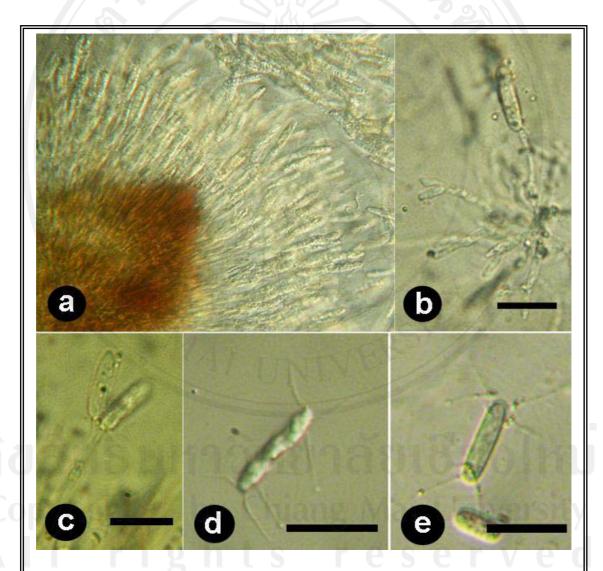


Fig. 2.6 Micrographs of *Chaetospermum chaetosporum* on *Wallichia siamensis*. a-b. Conidiophores. c. Conidiogenesis. c-d. Conidia. Bars: b, $c=20~\mu m$; d, $e=30~\mu m$.

Notes: Recently only three species belong to genus *Chaetospermum*, viz, *Chaetospermum carneum* Tassi, *C. chaetosporum* and *C. gelatinosum* Petch (Sutton, 1980; Kirk *et.al*, 2008). According to the key of Sutton (1980), this species is much closed to *C. chaetosporum* in having similar ascospores size and appendages (Sutton, 1980). This specimen is the first record of *C. chaetosporum* from Thailand, and *W. siamensis* is reported here as a new host.

Chaetosphaeria fusiformis W. Gams & Hol.-Jech., Mycotaxon 13: 257 (1981)

= Chaetosphaeria fusispora W. Gams & Hol.-Jech., Stud. Mycol. 13: 45 (1976). (Fig. 2.7)

Ascomata superficial, black, smooth-walled, shining, subglobose, ostiole conical, without a basal stroma, 110-160 mm diam., 125-190 mm high, consisting of numerous layers of thick-walled, pigmented cells. *Paraphyses* 1.3-2.5 μm, hyphallike, hyaline. *Asci* 100-165 × 7.5-12.5 μm, cylindrical-clavate, thin-walled, apedicellate, with a J-, cylindrical apical ring. *Ascospores* 47.5-82.5 × 2.5-3.8 μm, long fusiform to filiform, hyaline,smooth, with a somewhat rounded upper end, multiseptate,

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on decaying petioles of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 021).

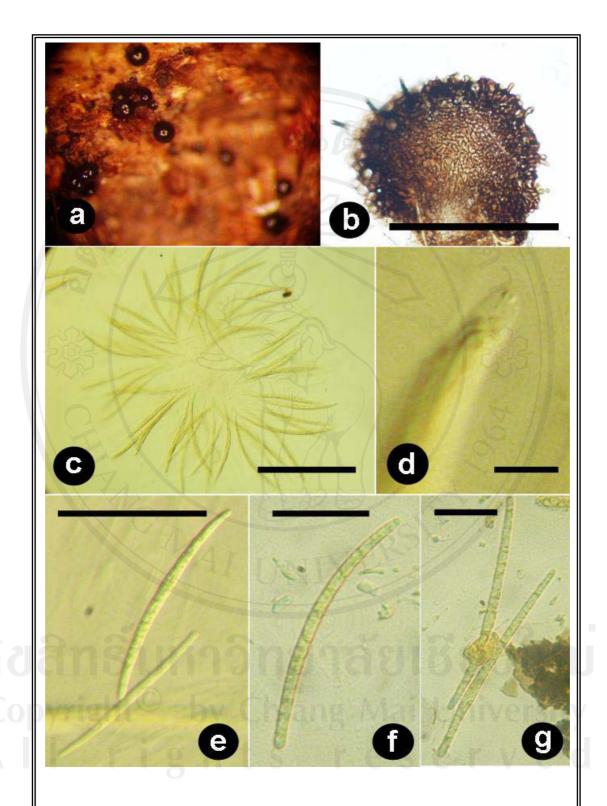


Fig. 2.7 Micrographs of *Chaetosphaeria fusiformis* on *Wallichia siamensis*. a-b. Ascomata. c. Asci. d. Ascus ring. e-g. Ascospores. Bars: b=150 mm; c=150 μ m; d=7.5 μ m; e=50 μ m; f=20 μ m, g=10 μ m.

Host: *Abie alba* (Gams and Holubová-Jechová, 1981), Gramineae (particularly bamboos genera) (Hino, 1961).

Distribution: Cezh Republic (Gams and Holubová-Jechová, 1981), Japan (Hino, 1961).

Notes: Based on the dichotomous key to species within genus *Chaetospaheria* by Reblova *et. al.* (1999), this specimen is much closed to *C. fusiformis* in almost all morphological appearances.

Fasciatispora petrakii (Mhaskar & V. G. Rao) K. D. Hyde, Nova Hedwigia 61: 255 (1995b)

= Amphisphaerella petrakii Mhaskar & V. G. Rao, Current Science 42: 28 (1973).

(Fig. 2.8)

Ascomata immersed in pseudostroma, black, solitary, in vertical section globose to subglobose, 400-500 μm diam., 230-425 μm high. *Peridium* 2.5-7.5 μm wide, composed of elongate or angular cells. *Paraphyses* 1-2.5 μm wide, aseptate, hyaline, embedded in agelatinous matrix. *Asci* 83.5-100 × 9.5-11 μm, unitunicate, cylindrical, 8-spored, J+ subapical ring, short pedicellate, apically rounded. *Ascal ring* 1-1.5 μm in high, 2-3 μm in diameter. *Ascospores* 11.6-16.7 × 5.1-6.5 μm, ellipsoid, 1-celled, hyaline at the middle part and dark brown at the 1/3 ends, surrounded by mucilaginous sheath.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on laeflets of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 147).

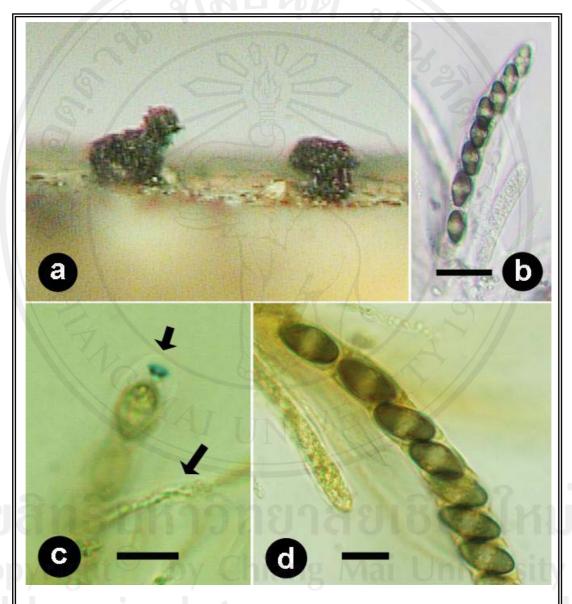


Fig. 2.8 Micrographs of *Fasciatispora petrakii* on *Wallichia siamensis*. A. Ascomata appearance on the host surface. b. Ascus. C. Ascus ring and paraphyses (arrows). d. Uniseriate ascospores. Bars: $b=15~\mu m$; $c=7.5~\mu m$; $d=5~\mu m$.

Host: Calamus spp., Cyrtostachys spp., Daemonorops spp., Elaeis spp., Livistona spp., Metroxylon spp., Oncosperma spp., Sabal spp. (Hyde, 1995b), Archontophoenix alexandrae, Trachycarpus fortunei, Cocos nucifera (Taylor and Hyde, 2003).

Distribution: Brunei, India, malaysia, papua New Guinea, Philippines, U. S. A. (Hyde, 1995b), Australia, China, Hong Kong, Singapore (Taylor and Hyde, 2003).

Notes: The first report of this species from Thailand was carried out by Hidayat *et al.* (2007).

Glomerella cingulata (Stoneman) Spauld. & H. Schrenk, Science, N. S. 17: 751 (1903)

= Gnomoniopsis cingulata Stoneman, Botanical Gazette **26**:101 (1898). (Fig. 2.9)

Ascomata 175-450 \times 125 - 300 μ m, immersed on the host surface, obpyriform, black, setae, solitary or in small groups. *Paraphyses* 2-5 μ m wide, numerous, septate, hyphal-like, hyaline, cellular. *Asci* 65.5-72 \times 5-12.5 μ m, unitunicate, 4-8-spored, cylindric to pyriform, thick at the apex, short pedicellate, apically truncate, with a refractive J-, subapical ring, *Ascospores* 18-24.5 \times 7-7.5 μ m, biseriate to multiseriate, ellipsoid to reniform to ovoid, 1-celled, hyaline.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 16

October 2005, Iman Hidayat (FIH 137); *ibid*. (FIH 144); Queen Srikit Botanical Garden, on leaflets of *Livistona fulva* Rodd (Arecaceae), 25 November 2006, Tatiya Takaew (FIH 418).

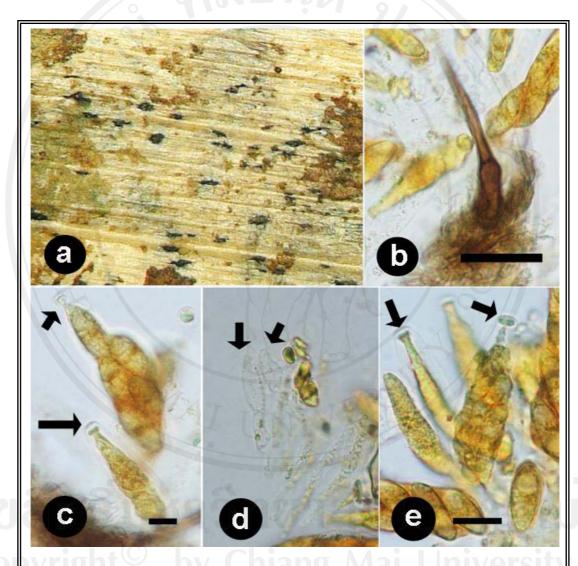


Fig. 2.9 Micrographs of *Glomerella cingulata* on *Wallichia siamensis*. a. Apperance of ascomata on the host surface. b. Setae. c. Asci with a J- apical ring (arrows). d. Paraphyses. e. Ascal ring (showed by arrows). Bars: $b=50~\mu m$; $c=7.5~\mu m$; $e=15~\mu m$.

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Host: Associated with a wide range of plants (USDA fungal-host database).

Distribution: Worldwide, from temeperate to tropical regions (Arx, 1987).

Notes: Glomerella cingulata is a common pathogen all over the world on leaves of various plants. It is the agent of bitter rot and anthracnose of Citrus, Maltus and Pyrus, and also occurs on berries of Prunus, Coffea, Sambucus and many other plants. The infection is often latent and the respective plants usually show a poor growth. Ascomata sometimes also develop in pure culture. Germinating ascospores form pigmented appressoria. The common anamorph is known as Colletotrichum gloeosporioides (Penz.) Penz. & Sacc. (Arx, 1987).

Guignardia uniappendiculatum Hidayat & To-anun, sp. nov.

(Fig. 2.10)

Differt a Guignardia cocogena et G. candeloflamma asci decretus (47.5-65 \times 15-22.5 μ m), et ascospori uniappendicum.

Etymology: Named after its single appendage of the ascospores.

Stromata lacking. *Ascomata* immersed, black, developing under slightly raised regions, in vertical section 380-825 μ m diam., 75-145 μ m high, subglobose, with a central ostiole. *Peridium* composed of brown-walled textura globulosa, inwardly compressed. *Pseudoparaphyses* up to 4.5 μ m wide, cellular, forming chains-like structure, hyaline, tapering to the apex. *Asci* 47.5-65 \times 15-22.5 μ m, bitunicate, 8-

spored, clavate, thick-walled, short-pedicellate, with ocular chamber near the apex, apex subtruncate. *Ascospores* 15-21.3 \times 5-7.5 μ m, overlapping 2-3 seriate, hyaline-greenish, ovoid, lacking germ pores, with broader part above the middle part of the spores, 1-celled, not smooth, surrounded by a thin mucilagenous sheath, with a distinct appendage at the basal end.

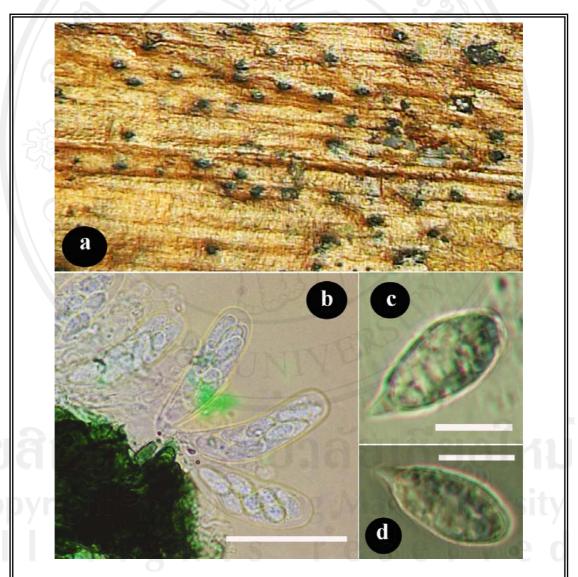


Fig. 2.10 Micrographs of *Guignardia uniappendiculatum* **sp. nov.** (from holotype). a. Ascomata appearance on the host. b. Asci. c-d. Ascospores with a basal appendage. Bars: $b = 50 \mu m$; $c, d = 7 \mu m$.

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Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park,

Huay Kog Ma, on leaflets of Wallichia siamensis Becc. (Arecaceae), 30 April 2007,

Iman Hidayat and Jamjan Meeboon (Holotype: FIH 016). Paratype: ibid., 16

November 2006, Tatiya Takaew (FIH 314).

Host: Leaflets of *W. siamensis*.

Distribution: This species is only known from the type locality.

Notes: Two species of palmicolous Guignardia species, viz, G. cocogena

(Cooke) Puntih. on Cocos nucifera and G. candeloflamma J. Fröhl. & K.D. Hyde on

Pinanga sp., are the most similar species to the present novel species as having

ornamented ascospores with polar appendages (Hyde, 1995a). However, Guignardia

uniappendiculatum is distinct from the previous two species due to only having single

polar appendage at the basal end of ascospores.

Mycosphaerella wallichiae Hidayat and To-anun, sp. nov.

(Fig. 2.11)

Differt a Mycosphaerella frenumbensis et M. serrulata asci obclavata, 40-

× 3.8-6 μm., et ascospori ornamentum.

Etymology: Named after its genus name of host, Wallichia.

Leaf spots amphigenous, irregular, lesions brown. Ascomata pseudothecial,

amphigenous, subepidermal, immersed to erumpent on the host surface, black, globose

to subglobose, 50-75 μ m diam., 60-90 μ m high, apical ostiole 5-10 μ m diam. **Peridium** wall of 2-3 layers of medium brown textura angularis. **Paraphyses** absent. **Asci** 40-50 \times 3.8-6 μ m, bitunicate, 8-spored, obclavate. **Ascospores** 10-12 \times 2-3.8 μ m, biseriate overlapping, 1-septate, slightly constricted at the septum, unequally bicelled, fusoid-ellipsoidal with obtuse ends, hyaline, thin-walled, straight to slightly curved, tapering towards the lower end, surrounding by a thin mucilagenous sheath, often with mucilagenous pad at the narrow base.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (**Holotype**: FIH 013).

Host: Leaflets of Wallichia siamensis.

Distribution: This species is only known from the type locality.

Notes: According to the key of *Mycosphaerella* on palms by Fröhlich and Hyde (1998), this specimen is much closed to *M. frenumbensis* (Speg.) Tomilin and *M. serrulatae* (Ellis & Everh.) Diehl by having obclavate to cylindrical asci with unequal bicelled ascospores. However, this specimen is distinct to the previous two species by having ascospores surrounded by mucilagenous sheath and ornamentation at the basal end.

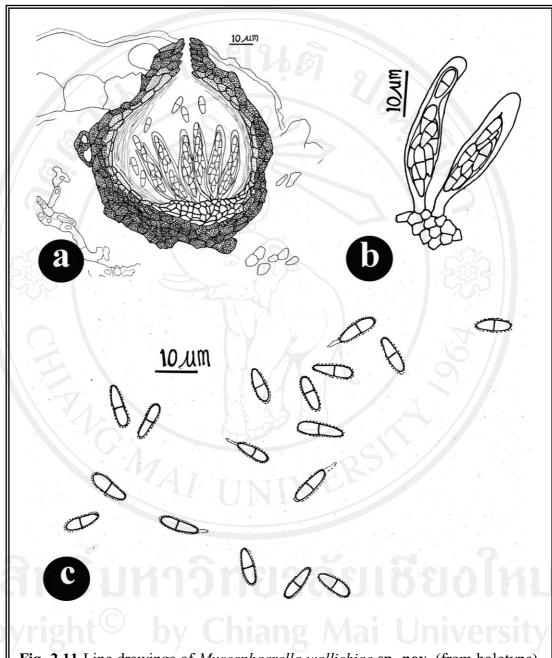


Fig. 2.11 Line drawings of *Mycosphaerella wallichiae* **sp. nov.** (from holotype). a. Vertical section through ascoma. b. Asci. c. Ascospores with mucilagenous pad at one end.

Myelosperma tumidum Syd. & P. Syd., *Annal. Mycol.* **13**: 38 (1915) **(Fig. 2-12)**

Ascomata immersed, darkened brown, in groups with 3 ascomata (1 large and 2 small) with a common central pore, in vertical section 300-550 μm diam., 150-375 μm high, subglobose. *Peridium* 5-10 μm wide, comprising layers of brown-walled elongate cells, inwardly hyaline and outwardly textura intricata. *Paraphyses* 5-7.5 μm wide, hyphal-like, septate, hyaline. *Asci* 145-217.5 × 12.5-20 μm, unitunicate, cylindrical, long pedicellate, 8-spored, with refractive, J-, discoid sub apical ring. *Ascospores* 15-27.5 × 6.3-10 μm, uniseriate, fusiform to reniform, smooth, aseptate but old ascospores has a septa-like morphology(1-3 septate-like), hyaline, surrounded by mucilaginous sheats.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 025); Chiang Mai University, Palm garden, on leaflets of *Borassodendron* sp. (Arecaceae), 29 October 2006, Iman Hidayat (FIH 108).

Host: Calamus tetradactylus, Calamus thysanolepis, Calamus walkerii, Calamus sp., Cocos nucifera, Daemonorops margaritae, Licuala sp., Livistona chinensis, Livistona sp. (Fröhlich and Hyde, 2000).

Distribution: Brunei Darussalam, Hong Kong, Indonesia, Papua New Guinea, Sri Lanka (Fröhlich and Hyde, 2000).

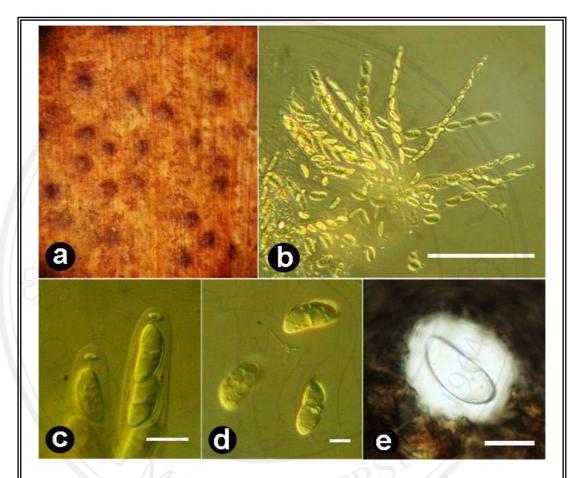


Fig. 2.12 Micrographs of *Myelosperma tumidum* on *Wallichia siamensis*. a. Appearance of ascomata on the host surface. b. Asci. c. Ascus ring. d. Ascospores. e. Mucilagenous sheath appearances using iodine solution. Bars: $b=175~\mu m$; $c=15~\mu m$; $d, e=7.5~\mu m$.

Notes: This specimen is the first record of *Myelosperma tumidum* from Thailand, and *W. siamensis* is reported as a new host of this fungus.

Neorehmia arecae (Syd.) Samuels & M. E. Barr, Canadian Journal of Botany 75: 2165-2176 (1997)

Oplothecium arecae Syd., Annales Mycologici 21: 97 (1923).(Fig. 2.13)

Leaf spots amphigenous, rounded, brown outwardly and dark-brown at the middle, fruiting bodies hypophyllous, 6-13 mm diam., 3.5-9 mm high. Ascomata in section 70-91 μm diam., 49-69 μm high, globose, non-papillate, solitary or caespitose, superficial, seated on a thin, hyphal cushion. Setae 5-7 μm wide, 10-17 μm high, stout-cylindrical, erect, 3 to 4 branches at the apex, dark brown, arising as outgrowth of cells at the wall surface. Peridium 7-12 μm wide, comprising 3 to 4 light to dark brown ellipsoidal layers. Paraphyses numerous, narrow, delicate, interspersed among the asci. Asci 24-36 × 3-4 μm, 8-spored, unitunicate, cylindrical to clavate, apedicellate, apically rounded, with a J-, apical ring. Ascospores 6-7 × 5-6 μm (part spores 3-3.5 × 2.5-3 μm), overlapping biseriate, 1-septate, ellipsoidal but then disarticulating into two equal parts subglobose with germ pore at the center, hyaline.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on living leaflets of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 126) (MRC 00202); *ibid.*, 25 October 2006, Iman Hidayat & Jamjan Meeboon (FIH 354) (CMU 27950).

Host: Areca catechu (Müller and Dennis, 1965), Wallichia siamensis (Toanun et al., 2009).

Distribution: Philippines, Venezuela (Müller & Dennis, 1965), Thailand (Toanun *et al.*, 2009).

Notes: The species collected in this study is identified as *Neorehmia* based on setae which are branched at the tips, the partspores which are subglobose and smooth walled. Three species are known in *Neorehmia*, *N. ceratophora* Höhn (type species), *N. arecae* (syn. *Oplothecium arecae*, *T. arecae*), *N. aurea* (Höhn.) Munk (≡ *Bresadolella aurea* Höhn.); however, Rossman *et al.* (1999) noted that *N. aurea* is a synonym of *T. decipiens*, the type species of *Trichosphaerella*. Therefore, *N. aurea* is an invalid name and only two *Neorehmia* species have been accepted in recent publication (Rossman *et al.*, 1999).

The type species of *Trichosphaerella* is *T. decipiens* E. Bommer, M. Rousseau & Sacc. and is characterized by short, dark and simple setae. Samuels and Barr (1997) transferred *Trichosphaerella arecae* to *Neorehmia arecae* (Syd.) Samuels & M. E. Barr (basionym: *Oplothecium arecae* Syd.; *Ann. Mycol.* 21: 97. 1923) (Trichosphaeriaceae; Trichosphaeriales) based on the similar characteristic to *N. ceratophora* Höhn (type species) which is characterized by setae which are branched at the tips. The partspores of *N. ceratophora* are subglobose, smooth-walled, and numerous; narrow and delicate paraphyses are usually seen interspersed among the asci. *Neorehmia* is also connected to *Tritirachium*-like anamorph by Muller and Samuels (1982) whereas *Trichosphaerella* was linked to *Acremonium*-like anamorph.

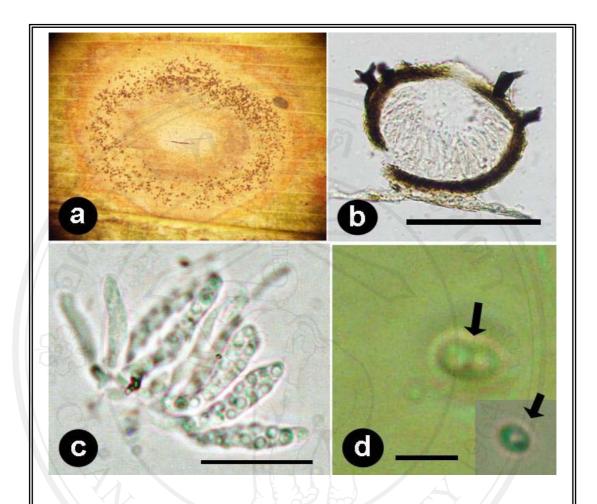


Fig. 2.13 Micrographs of *Neorehmia arecae* on *Wallichia siamensis*. a. Symptom. b. Vertical section of ascoma. c. Asci. d. Ascospore (septum and part of ascospore showed by arrows). Bars: $b=75~\mu m$; $c=25~\mu m$; $d=7~\mu m$.

Neorehmia ceratophora differs to this species in having delicate paraphyses interspersed among the asci and the spinulose ascospores in which this species is lacking of paraphyses and the spores are smooth. The host of *N. ceratophora* is also different being recorded from rotting conifers and hardwoods, possibly fungicolous and also found on small sticks of *Salix* and on rotting wood of *Carpinus betulus*.

Neorehmia arecae is most similar to this species, due to flattened and sphaeroid fruit-bodies with a few short setae branching at the tip and also in being

recorded from a palm (Müller and Dennis, 1965; Samuels and Barr, 1997). This species is only different to N. arecae only in the part of spores' size (3.2 × 2.7 μ m vs 1.5 × 2 μ m of N. arecae) (Müller and Dennis, 1965). Therefore, we report this species as Neorehmia arecae. This is the first report of N. arecae from Thailand and also W. siamensis as the host of the fungus.

Ophioceras tenuisporum Shearer, J. L. Crane & W. Chen, Mycologia 9: 149 (1999)
(Fig. 2.14)

Ascomata 175-300 × 200-350 μm, globose to subglobose, immersed or beneath under host surface, darkened (blackish), solitary to gregarious. Neck composed of textura intricata with hyphae arranged in rows and fanning out obliquely, 125-280 × 45-65 μm, central, cylindrical, pale brown to black and hyaline at the apex. Peridium thin, comprising a few layers of dark brown to black-walled compressed cells, composed of large cells of textura angularis. Paraphyses hyaline, 0.5-0.75 μm, filamenttous, smooth, septate. Asci 90-122.5 x 5-7.5 μm, unitunicate, 8-spored, cylindrical, with rounded apex, short pedicellate, refractive apical ring, J- apical ring, Ascospores 70-95.5 x 1.25-1.5 μm, multiseriate, filliform, multiseptate, hyaline, both ends are slightly broader immediately above and below each septum.



Fig. 2.14 Micrographs of *Ophioceras tenuisporum* on *Wallichia siamensis*. a. Ascomata with necks. b. Asci with short pedicellate (pedicellates and apical ring showed by arrows). c-d. Ascospores. Bars: $b = 50 \mu m$; $c = 25 \mu m$; $d = 15 \mu m$.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on decaying leaflets of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 011); Bahn Phadeng, on *Cocos nucifera* L. (Arecaceae), 13 July 2005, Iman Hidayat (FIH 034).

Host: Saprobic on freshwater decaying woods (Shearer *et al.*, 1999), *Miscanthus* spp. (Lu *et al.*, 2000).

Distribution: Panama (Shearer et al., 1999), Hong Kong (Lu et al., 2000).

Notes: This specimen is the first report of *Ophioceras tenuisproum* associated with *Wallichia siamensis*.

Oxydothis daemonoropsicola J. Fröhl. and K. D. Hyde, Fungal Diversity Research Series 3: 183 (2000)

(Fig. 2.15)

Ascomata forming under weakly raised, blister-like regions, single or clustered in groups 2-3; in section subglobose, long axis horizontal to that of the host surface with neck at one end, ca 135-250 μm diam., 106-127 μm high; Peridium 17.5-20 μm thick, composed of 3 to 5 outer layers of oblong, dark-brown cells and sometimes with an additional inner layer of oblong, hyaline cells; Paraphyses numerous, up to 5 μm diam.; Asci cylindrical, 8-spored, pedicellate, unitunicate, sub apical ring with a rectangular shape, J+ sub apical ring, 225-255 × 12.5-13.75 μm; Ascal ring 2-2.3 high, 2-3 μm diam.; Ascospores 95-105 × 5-6.3 μm, 1-3-seriate,

long fusiform to filiform, hyaline, 1-septate, tapering very gradually from a unconstricted central septum to form rounded ends.

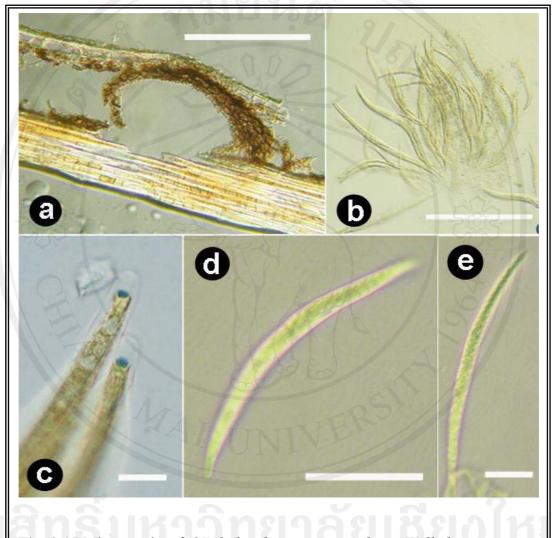


Fig. 2.15 Micrographs of *Oxydothis daemonoropsicola* on *Wallichia siamensis*. a. vertical section of ascoma. b. Asci. c. Ascal ring. d-e. Ascospores. Bars: $a=200~\mu m; b=225~\mu m; c=30~\mu m; e=15~\mu m.$

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on decaying rachis of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 019, 141) (MRC 0005, 0006).

Host: Daemonorops margaritae, Archontophoenix alexandrae, Wallichia siamensis (Taylor and Hyde, 2003; Hidayat et al., 2006).

Distribution: Australia, Hong Kong, Malaysia, and Thailand (Taylor and Hyde, 2003; Hidayat *et al.*, 2006).

Notes: In Thailand, this species was reoprted by Hidayat et al. (2006).

Oxydothis inaequalis Hidayat, To-anun and K.D. Hyde, Fungal Diversity 23: 165 (2006)

(Fig. 2.16)

Stromata 5–40 mm long × 5–10 mm wide, visible as blackened ellipsoidal regions on the host surface, lacking borders. Ascomata 110–120 μm diam. × 22–30 μm high, forming slightly raised domes, singly or clustered in groups up to 10 (mostly 2–3); in section immersed, ellipsoid, long axis horizontal to that of the host surface, papilla at one end curving upwards to the host surface. Stromatic tissue surrounds the ascomata within the host hypodermis. Peridium 10–12.5 μm thick, comprised of 2–3 layers; outer layers of oblong, dark-brown cells and sometimes with an additional inner layer of oblong, hyaline cells. Paraphyses deliquescent early, septate, ca 2.5 μm in diam. Asci 200–285 × 11.25–12.5 μm, 8-spored, unitunicate, cylindrical, short pedicellate, J+, 4–6(–7) μm high, 3–4 μm diam., wedge-shaped, subapical ring, apically truncate. Ascospores 75–100 × 5–7.5 μm, 1–2 seriate, fusiform, 1-septate, hyaline, tapering gradually to form long pointed processes. The ascospores processes are sometimes uneven which may make the septum appear slightly eccentric.

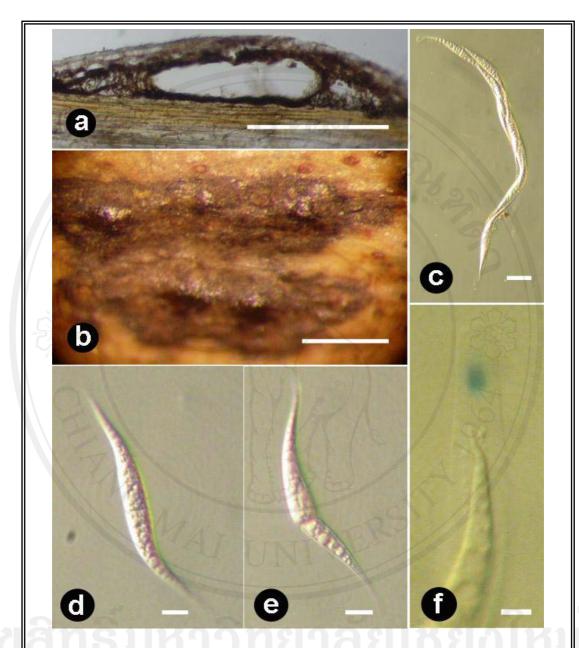


Fig. 2.16 Micrographs of *Oxydothis inaequalis* (from holotype). a. Vertical section through the ascoma. b. Ascomata on the host surface. c. Ascus. d-e. Ascospores. f. Amyloid positive of ascal ring. Bars: $a=110~\mu m$; b=10~mm; $c=12~\mu m$; $d=7~\mu m$; $e=7.5~\mu m$; $f=4~\mu m$.

Anamorph: Unknown.

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Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, decaying rachis of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 018) (**Holotype**: MRC 0004). **Isotype**: *ibid.*, HKU (M) 17169.

Host: Saprobic on the *W. siamensis* fronds.

Distribution: Northern Thailand.

Notes: A detail morphology and molecular elucidation of this species is presented in chapter 3.

Oxydothis wallichianensis Hidayat, To-anun and K.D. Hyde, Fungal Diversity 23: 167 (2006)

(Fig. 2.17)

Stromata 2–3 mm long \times 1.5–2.5 mm wide, surrounded by ellipsoidal, brown borders. Ascomata 70–150 μ m diam. \times 55–100 μ m high, stromata domes on the host surface, mostly clustered in groups of up to 18; in section immersed to erumpent, subglobose, papilla at one end curving upwards to the host surface. Peridium 5–8.75 μ m thick comprised of 2–3 layers outer layers of oblong, dark–brown cells. Asci 87.5–125 \times 10–15 μ m, 8-spored, unitunicate, cylindrical, pedicellate, with a J+, 2–2.5 μ m high \times 3–3.5 μ m diam., wedge-shaped, subapical ring. Ascospores 32.5–55 \times 6.3–7.5 μ m, 1–2-seriate, fusiform, 1-septate, hyaline, tapering abruptly near the ends to form long spine-like processes.

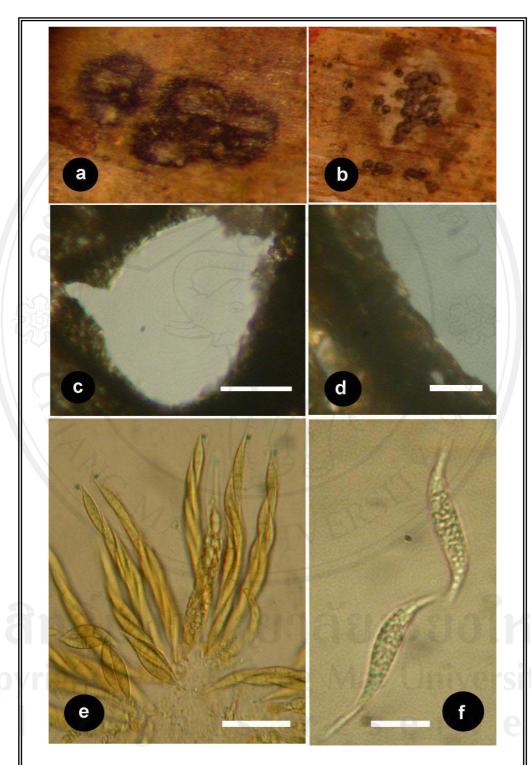


Fig. 2.17 Micrographs of *Oxydothis wallichianensis* (from holotype). a-b. Ascomata on the host surface. c. Vertical section through the ascoma. d. Peridium. e. Asci and ascal ring. f. Ascospores. Bars: $c=50~\mu m;~13,~d=5~\mu m;~e=50~\mu m;~f=10~\mu m.$

100

Anamorph: Unknown.

Material examined: THAILAND, Chiang Mai, Doi Suthep, on decaying

leaflets of Wallichia siamensis Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH

010) (Holotype: MRC 0002). Isotype: ibid., HKU (M) 17174.

Host: Leaflets of *W. siamensis*.

Distribution: Northern Thailand.

Notes: A detail morphology and molecular elucidation of this species is

presented in chapter 3.

Pemphidium rattanicola J. Fröhl. and K.D. Hyde, Fungal Diversity Research Series

3: 225 (2000)

(Fig. 2.18)

Ascomata developing in clusters beneath the uppermost layers of the host

epidermis, darkened, dome-shapped areas, immersed, with central ostiole. *Paraphyses*

numerous, 2.5-7.5 µm wide, tapering towards the tip, hypha-like, filamentous, and

septate. Asci 95-120 × 7.5-11.3 µm, unitunicate, 8-spored, cylindrical to clavate, short

pedicellate, refractive sub apical ring, J- sub apical ring. Ascospores 67.5-87.5 × 1.8-

3.8 µm, filiform, 1-septate, 3-4 seriate, asymmetric, tapering slightly from the broader

apex to a narrower base, hyaline, smooth walled, mucilagenous drop at one of the the

ascospores tip.

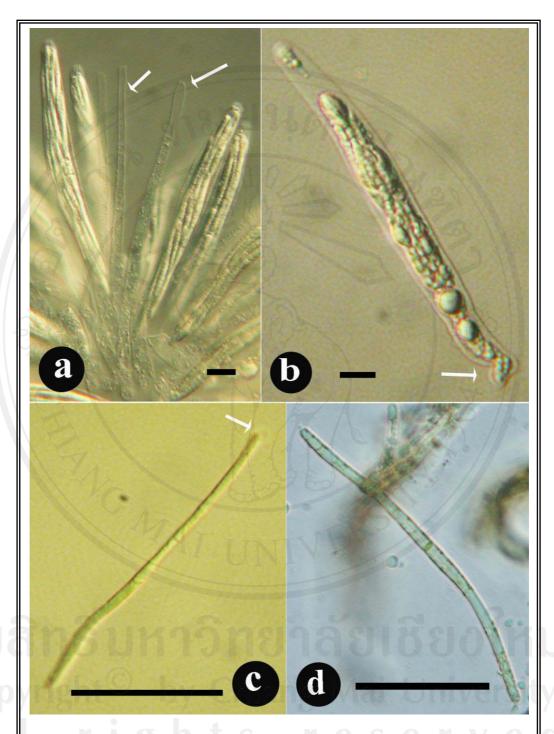


Fig. 2.18 Micrographs of *Pemphidium rattanicola* on *Wallichia siamensis*. a. Asci and paraphyses (showed by arrows). b. A single ascus with short pedicellate (showed by arrow). c-d. Ascospores with a mucilage pad at one end (showed by arrow). Bars: a, b = $10 \mu m$; c, d = $20 \mu m$.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 21 July, 2005, Iman Hidayat (FIH 005).

Host: Calamus caryotoides, C. moti (Fröhlich and K.D. Hyde, 2000).

Distribution: Australia (Fröhlich and K.D. Hyde, 2000).

Notes: This specimen is identified as *P. rattanicola* due to its unusually wide asci and in having ascospores with a mucilage pad at only one end. This specimen is the first record of *P. rattanicola* from Thailand, and *W. siamensis* is reported here as a new host of this fungus.

Pestalosphaeria elaeidis (Booth & Robertson) Aa, Proc. K. Ned. Akad. Wet., Ser. C, Biol. Med. Sci. 67: 87 (1976)

- = Leptosphaeria elaeidis C. Booth & J.S. Robertson, Transactions of the British Mycological Society 44: 24 (1961).
- *Example 1981* = Lepteutypa elaeidis (C. Booth & J.S. Robertson) Arx, The genera of fungi sporulating in pure culture: 176 (1981).

(Fig. 2.19)

Ascomata darkened, immersed, solitary perithecial, gregarious, in section, $150\text{-}250 \times 180\text{-}210 \,\mu\text{m}$, subglobose, with an ostiole in the papilla. *Peridium* 15-25 $\,\mu$ m thick, consisting of two strata; outer stratum composed of 3-5 layers of brown, thick-walled, very compressed cells; inner stratum composed of 1-2 layers of hyaline, thin-walled, very compressed cells. *Paraphyses* 2-4 $\,\mu$ m wide, hyphal-like, septate,

narrow to the ends, not branched, hyaline. *Asci* 55-102.5 \times 8.8-12.5 μ m, unitunicate, 8-spored, cylindrical, pedicellate, with refractive, discoid, J+ sub apical ring, 1-2 μ m high, 2-2.8 μ m diam. *Ascospores* 15-16.3 \times 5-6.3 μ m, uniseriate overlapping, fusiform, mostly 2-septate sometimes 3-septate, brown, smooth.

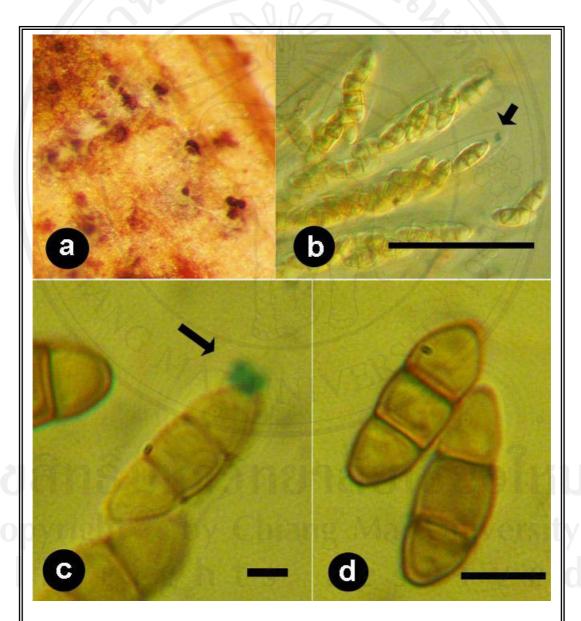


Fig. 2.19 Micrographs of *Pestalosphaeria elaeidis* on *Wallichia siamensis*. a. Ascomata appearance on the host surface. b. Asci (ascus ring showed by arrow). c. Ascus ring (arrow). d. Ascospores. Bars: $b=75~\mu m$; $c=2.5~\mu m$; $d=10~\mu m$.

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Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park,

on leaflets of Wallichia siamensis Becc. (Arecaceae), 21 July 2005, Iman Hidayat

(FIH 027).

Host: Elaeis guineensis (Hyde, 1996).

Distribution: Nigeria (Hyde, 1996).

Pestalotiopsis guepinii (Desm.) Steyaert [as 'guepini'], Bulletin du Jardin Botanique

de l'État à Bruxelles 19: 312 (1949)

Pestalotia guepinii Desm., Annales des Sciences Naturelles, Botanique 13:

181 (1840)

(Fig. 2.20)

Conidiomata acervular, scattered or gregarious, sub-epidermal, remaining

immersed, visible at the surface by dark exuding conidial masses; in section ellipsoid,

 $185-400 \times 175-225 \,\mu\text{m}$. **Peridium** 12.5-18 $\,\mu\text{m}$ thick, consisting of 3 to several layers

of pale brown, moderately thick-walled cells of textura angularis. Conidiophores

arising from the entire periphery of the inside of the conidiomata, reduced to

conidiogenous cells or poorly developed, branched at the base, ampulliform.

Conidiogenous cells 7.5-12.5 × 1-2 µm, annellidic, hyaline, discrete or integrated,

smooth, cylindrical. *Conidia* fusiform, 20-30 × 6.5-8.5 µm, 4-septate; *apical cell*

hyaline and thin-walled; median cells doliiform, brown, with third and fourth cells

from the base darker than the second cell (at times the third cell darker than the fourth

cell), moderately thick-walled; basal cell hyaline to subhyaline, obconical, smooth,

thin-walled. *Apical appendages* 2-3, inserted along the upper half of the apical cell, arising at different points, unbranched, flexuous, $10-20 \times 1 \mu m$, attenuated. *Basal appendage* $2-7.5 \times 1 \mu m$, single, centric, unbranched, attenuated.

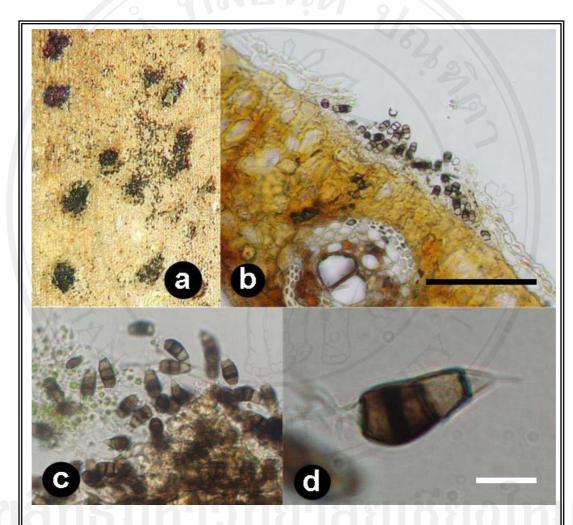


Fig. 2.20 Micrographs of *Pestalotiopsis guepinii* on *Wallichia siamensis*. a. Appearance of conidiomata on the host surface. b. Vertical section of conidioma. c-d. Ascospores. Bars: $b = 150 \, \mu m$, $d = 10 \, \mu m$.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 032).

Host: Camellia japonica, Camelia sinensis, Erica sp., Rhododendron sp., with occasion records on unrelated hosts including Hippocratea sp., Jatropha sp., Laurus sp., Pinus sp., Terminalia sp. (Sutton, 1980).

Distribution: Ghana, Kenya, Togo, India, Bohemia, France, Germany, Netherlands, UK, USA. Reported in literature from Belgium, Denmark, Italy, Sweden, Switzerland, Azores, Indonesia, Japan, New Zealand, Puerto Rico, Dominican Republic (Sutton, 1980).

Notes: This species is commonly recognized as an agent of grey leaf spot, twig dieback, stem canker, petal rot diseases on the plants (Guba, 1961).

Protocreopsis pertusa (Pat.) Samuels & Rossman, Stud. Mycol. 42: 66 (1999)

- ≡ Nectria pertusa Pat., in Patuoillard & Lagerheim, Bull. Soc. Mycol. France 11: 227 (1895).
- = Nectria scitula Bres., Hedwigia 35: 299 (1896).
 - ≡ Protocreopsis scitula (Bres.) Doi, Bull. Natl. Sci. Mus., Tokyo, B. 4: 116 (1978).
- = Nectria aemulans Rehm, Ann. Mycol. 7: 539 (1909).

(Fig. 2.21)

Ascomata superficial on the substratum or immersed in a hyphal subiculum to well developed, without hairs, perithecial, KOH-, white to pale yellow, in section

globose, ostiole central, 125-183 μ m diam, 175-220 μ m high. *Asci* 75–90 × 10–12.5 μ m, unitunicate, 8-spored, cylindrical, truncate at the apices, with refractive, cylindrical, J- apical ring. *Ascospores* 12.5–15 × 3.25–5 μ m, fusiform and /or ellipsoid, 1-septate, striate-punctate, striations visible few (up to 3), biseriate overlapping, hyaline, greenish in mass, with mucilaginous sheath.

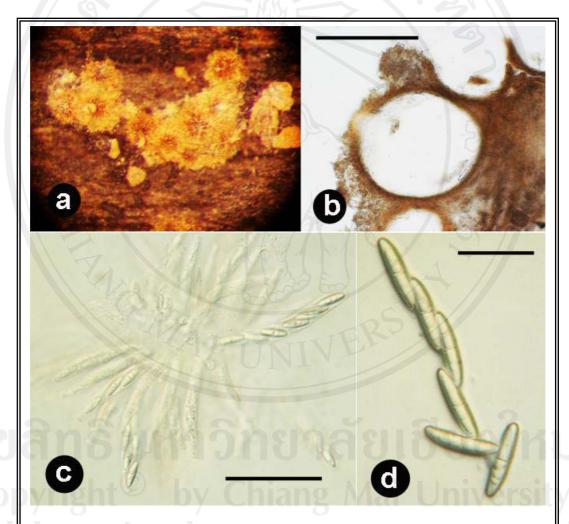


Fig. 2.21 Micrographs of *Protocreopsis pertusa* on *Wallichia siamensis*. a. Appearance of ascomata on the host surface. b. Vertical section of ascoma. c. Asci. d. Ascospores. Bars: $b = 100 \mu m$, $c = 75 \mu m$; $d = 12.5 \mu m$.

Specimens examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, Huay Kog Ma, on rachis of *Wallichia siamensis* Becc. (Arecaceae), 26 November 2006, Iman Hidayat (FIH 067); Bahn Phadeng, on petioles of *Cocos nucifera* L. (Arecaceae), 13 July 2005, Iman Hidayat (FIH 002).

Habitat: On leaves of *Musa*, *Heliconia*, and palms, culms of bamboo, and rachides of tree ferns (Rossman *et.al*, 1999).

Distribution: Probably pantropical and subtropical.

Notes: Rossman *et.al* (1999) noted that this is a common species in the American tropics.

Pseudohalonectria palmicola K. D. Hyde, Joanne E. Taylor & J. Fröhl., Mycologia91: 522 (1999)

(Fig. 2.22)

Ascomata immersed to superficial, clustered, black, coriaceous, in section 350-500 μm diam., 300-400 μm high, subglobose, with a long neck. *Peridium* 20-35 μm thick, composed of 3-4 layers of compressed, dark-brown, angular cells, inner cells hyaline. *Paraphyses* 3-7.5 μm wide, filamentous, hyaline, tapering towards the apex, embedded in a gelatinous matric. *Asci* 75-125 × 8.5-13.5 μm, unitunicate, 8-spored, cylindrical, with a subtruncate apex, apedicellate, with a J- apical ring. *Ascospores* 75-110 × 2.5-3.5 μm, multiseriate, filiform with tapering towards both ends, hyaline, multiseptate, smooth.

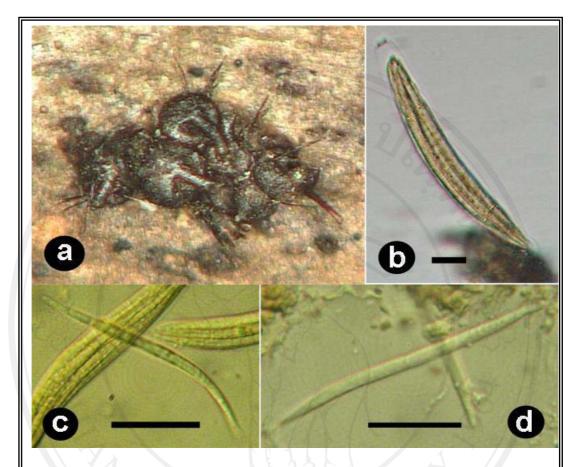


Fig. 2.22 Micrographs of *Pseudohalonectria palmicola* on *Wallichia siamensis*. a. Ascomata appearance on the host surface. b. Ascus. c-d. Ascospores. Bars: $b = 10 \mu m$, c, $d = 25 \mu m$.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on primary rachis of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 142).

Host: Calamus australis (Hyde et al., 1999).

Distribution: Australia (Hyde *et al.*, 1999).

Notes: This specimen is much closed to *Pseudohalonectria palmicola* in ascus size, ascal ring shape, and ascospores shape (Hyde *et al.*, 1999). This is the first report

of *Pseudohalonectria palmicola* from Thailand, and *W. siamensis* is reported here as a new host.

Rachidicola obclavatum Hidayat & To-anun, sp. nov.

(Fig. 2.23)

Differt a Rachidicola palmae asci 75-125 \times 12.5-22 μ m, obclavati et ascospori 22.5-27 \times 5-8 μ m, 2-3-seriati, 6-cellulis.

Etymology: Refers to the shape of ascus, obclavate.

Ascomata immersed beneath a clypeus, visible as raised, darkened, domw-shaped, areas, in section 300-500 μm diam., 185-250 μm high, ellipsoidal, coriaceous, dark brown, solitary, with central ostiole. *Peridium* 15-30 μm thick, inwardly composed of several layers of compressed, angular cells, outwardly composed of angular, dark-brown cells. *Paraphyses* 2.3-4 μm, filamentous, embedded in a gelatinous matrix, hyaline, septate. *Asci* 75-125 × 12.5-22 μm, unitunicate, 8-spored, short-pedicellate, obclavate, apically subtruncate, with a refractive, J- apical ring. *Ascospores* 22.5-27 × 5-8 μm, 2-3-seriate, fusiform, 6-cells, slightly constricted at the septum, hyaline, slightly curved, smooth, lacking a mucilagenous sheath.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (**Holotype**: FIH 140).

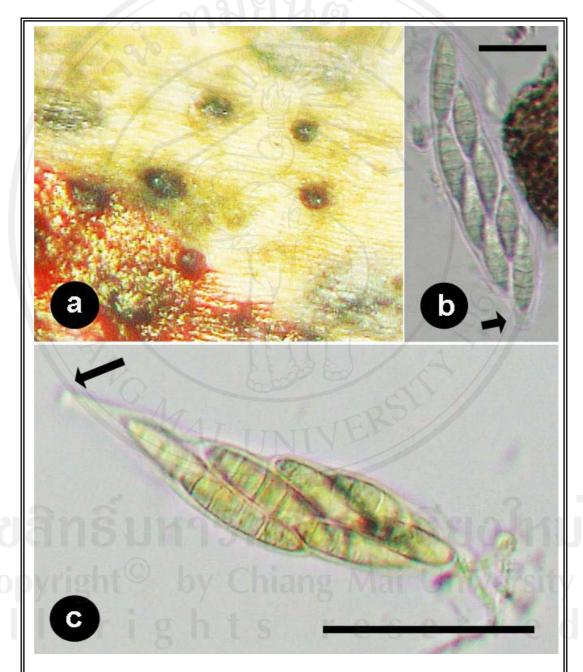


Fig. 2.23 Micrographs of *Rachidicola obclavatum* **sp. nov.** (from holotype). a. Ascomata appearance on the host surface. b. Ascus with short pedicellate (showed by arrow). c. Ascus with J- apical ring (showed by arrow). Bars: $b = 25 \mu m$, $c = 50 \mu m$.

Host: On leaflets of *W. siamensis*.

Distribution: Only known from the type locality.

Notes: This species is distinct from *Rachidicola palmae* K. D. Hyde & J. Fröhl. by having obclavate ascus and the septation of ascospores is more than 2-septate (5-septate) (Hyde and Fröhlich, 1995b).

Rosellinia victoriae Syd. & P. Syd., Annales Mycologici 6: 483 (1908)

- = Hypoxylon victoriae (Syd. & P. Syd.) P. M. D. Martin, South African Journal of Botany 42: 74 (1976).
- = *Hypoxylon victoriae* (Syd. & P. Syd.) P. M. D. Martin, *South African Journal of Botany* **33**: 325 (1967).

(Fig. 2.24)

Ascomata on palm rachides appear as superficial black domes with a central ostiole surrounded by persistent dark brown to black superficial hyphae (subiculum-like) within a well-developed carbonaceous stroma, 1-1.5 \times 0.5-1.5 mm. *Paraphyses* 1-2.5 μ m, not branched, numerous, hypha-like, septate, filiform, gradually tapering toward the apex. *Asci* 105-162.5 \times 8.8-13.8 μ m, unitunicate, cylindrical, long pedicellate, 8-spored, with a cylindrical, J+ sub apical ring, 3.5-4 high, 2-3 μ m diam. *Ascospores* 15-21.3 \times 5-8.8 μ m, fusiform to reniform, dark brown, uniseriate, surrounded by mucilaginous sheath, with a longitudinal straight germ slit in the center.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on primary rachis of *Wallichia siamensis* Becc. (Arecaceae), 21 July 2005, Iman Hidayat (FIH 026).

Host: Decaying wood (Petrini, 2003).

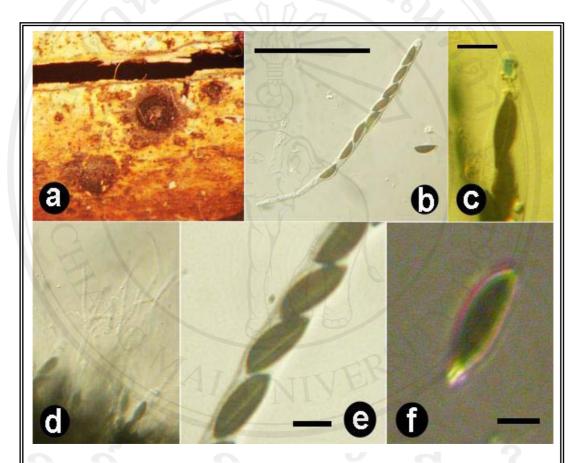


Fig. 2.24 Micrographs of *Rosellinia victoriae* on *Wallichia siamensis*. a. Ascoma. b. Ascus. c. Ascus ring. d. Paraphyses. e-f. Ascospores. Bars: b=75 μm ; c=10 μm ; e,f=7.5 μm .

Distribution: Australia, New Zealand (Petrini, 2003)

Notes: This specimen is the first report of *Rosellinia victoriae* from Thailand, and *W. siamensis* is reported here as a new host.

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Submersisphaeria suthepensis Hidayat & To-anun, sp. nov.

(Fig. 2.25)

Asci 160-165 x 10-10.5 μm, bitunicatis, 8-sporis, cylindricis, ascosporis 22-25

x 7.5-9 μm, tetracellularibus, fusiformis, non-appendiculatum.

Etymology: Named after its collection site, Doi-Suthep.

Ascomata immersed, black, solitary, gregarious, in vertical section, globose,

ostiolate, 120-185 µm diam., 200-250 µm high. Peridium up to 12 µm wide,

composed of inner flattened light brown-walled cells, and outer brown-walled angular

cells. *Paraphyses* 2.5-5 µm wide, hypha-like, septate, numerous, tapering towards the

apex, hyaline,not embedded in a gelatinous matrix. Asci 160-165 × 10-10.6 μm,

unitunicate, 8-spored, long cylindrical, long pedicellate, refractive and massive apical

ring, discoid, J- apical ring. Ascospores 22-25 × 7.5-9 μm, fusiform, 3-septate, dark

green to brown through the cell parts, thick walled, lacking of basal appendages.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park,

Huay Kog Ma, on primary rachis of Wallichia siamensis Becc. (Arecaceae), 21 July

2005, Iman Hidayat (Holotype: FIH 017).

Host: On primary rachis of *W. siamensis*.

Distribution: Only known from the type locality.

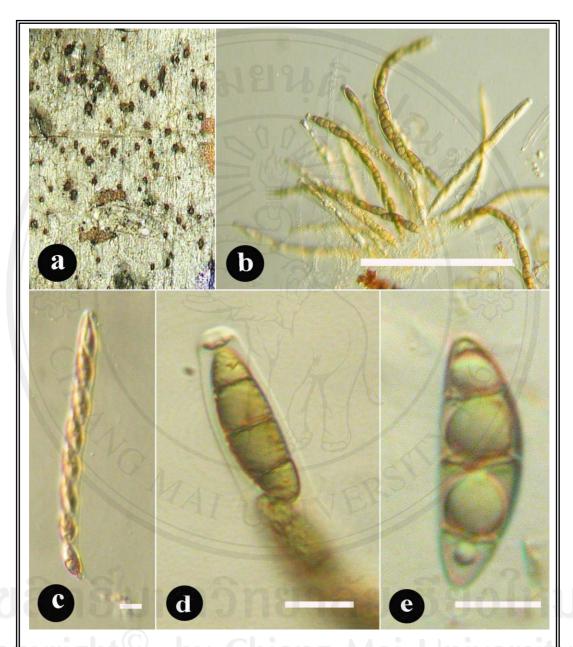


Fig. 2.25 Micrographs of *Submersisphaeria suthepensis* **sp. nov.** (from holotype). a. Ascomata appearance on the host surface. b. Asci. c. Ascus. d. Apical ring. e. Ascospore. Bars: $b = 150 \mu m$; c, $d = 10 \mu m$; $e = 7.5 \mu m$.

Notes: This species is much closed to *Submersisphaeria* K. D. Hyde than *Ascotaiwania* Sivan. & H. S. Chang due to the later species is characterized by having basal small hyaline/lightly cells at both ends and brown colour at the middle cells (Sivanesan and Chang, 1992). Recently, five species of *Submersisphaeria* have been reported, viz, *Submersisphaeria aquatica* K. D. Hyde, *S. bambusicola* D. Q. Zhou & K. D. Hyde, *S. palmae* Pinnoi, *S. rattanicola* J. Fröhl. & K. D. Hyde, and *S. vasicola* You Z. Wang, Aptroot & K. D. Hyde (Cai *et al.*, 2006). The present novel species is distinct to other *Submersisphaeria* species in having 4-cells ascospores, with non-appendages at both ends.

Terriera brevis (Berk.) P. R. Johnst. [as'breve'], Mycol. Pap. 176: 98 (2001)

- = Hysterium breve Berk., Transactions of the Linnaean Society of London 20: 174 (1847).
- Lophodermium breve (Berk.) De Not., Giornale botanico Italiano 2:47 (1847).
- ≡ Hypoderma breve (Berk.) Kuntze, Revisio generum plantarum 3: 487 (1898).

(Fig. 2.26)

Ascomata hysterothecial (lips-like), coriaceous, black, 1-3 mm long, 0.25-0.5 mm high, solitary or clustered. *Peridium* composed of several layers of irregular shaped cells with moderately thick, dark brown walls. *Paraphyses* 1-2.5 μm wide, hypha-like, numerous, branched. *Asci* 110-144 × 6-7.5 μm, unitunicate, clavate, 8-

spored, short pedicellate, with refractive J-, sub apical ring, apically rounded and thick. **Ascospores** 57.5-82.5 \times 1.25-2 μ m, filiform, aseptate, hyaline, lacking a mucilagenous sheath.

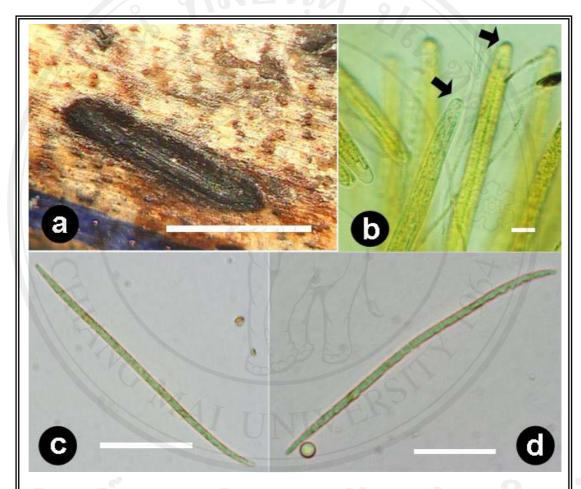


Fig. 2.26 Micrographs of *Terriera brevis* on *Wallichia siamensis*. a. Ascoma appearances on the host surface. b. Asci. c-d. Ascospores. Bars: a=1 mm; b=5 μ m; c=25 μ m; d=10 μ m.

Material examined: THAILAND, Chiang Mai, Doi Suthep-Pui national park, on decaying rachis of *Wallichia siamensis* Becc. (Arecaceae), 16 October 2005, Iman Hidayat (FIH 150).

Host: Carex sp., Uncinia spp., Gahnia sp. (Johnston, 2001), Archontophoenix alexandrae, Cocos nucifera, Trachycarpus fortunei (Taylor and Hyde, 2003).

Distribution: New Zealand (Johsnton, 2001), Australia, China, Hong Kong (Taylor and Hyde, 2003).

Notes: This specimen is the first record of *Terriera brevis* from Thailand, and *W. siamensis* is reported here as a new host.

New taxa from other palms

Apiosporina rhapisicola Hidayat & To-anun, sp. nov.

(Fig. 2.27)

Ascomata 90-98 μ m diam, 65-71 μ m alta, immersa, solitaria, subglobosa, ostiolata. Asci 69-87 \times 11.5-15.5 μ m, bitunicata, 8-spori, cylindrico-clavata, pedunculati, praediti Ascosporae 26.5-28 \times 5-6.5 μ m, fusiformis, brunneus, bicellulares, apiosporae.

Etymology: In reference to the host genus, *Rhapis*.

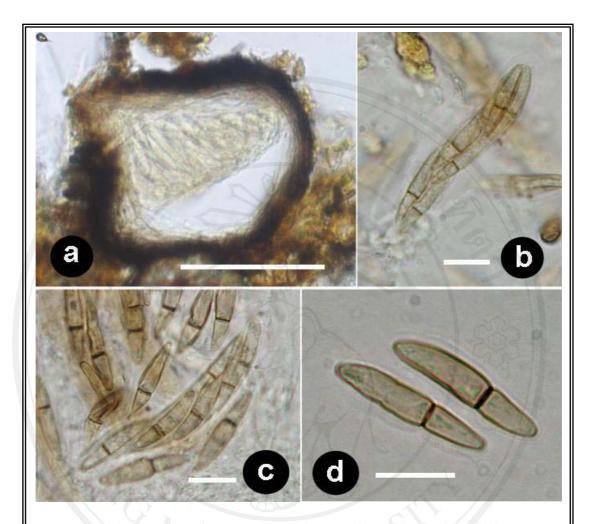


Fig. 2.27 Micrographs of *Apiosporina rhapisicola* **sp. nov.** (from holotype). a. vertical section of ascoma. b-c. Asci. d. Ascospores. Bars: $a=50~\mu m$; b, $c=10~\mu m$; $d=12~\mu m$.

Ascomata immersed, erumpent, solitary or clustered, in section 90-98 μm in diameter, 65-71 μm in height, subglobose, brown to dark-brown, ostiole central, papillate, lacking of setae around ostiole. *Peridium* 9.5-12 μm thick, comprising several layers, compressed, angular to ellipsoidal, dark brown cells. *Pseudoparaphyses* 1.5-2.5 μm wide, numerous, filamentous, cellular, hyaline, embedded in gelatinous matrix. *Asci* 69-87 × 11.5-15.5 μm, bitunicate, 8-spored, cylindrical-clavate, short pedicellate, apically rounded. *Ascospores* 26.5-28 × 5-6.5

μm, irregularly, fusiform, 1-septate, light-brown, septa constricted at the medium or sub-medium, upper cell wider and longer than lower cell.

Material examined: THAILAND, Chiang Mai, Queen Sirikit Botanic Garden, on living leaflets of *Rhapis* sp. (Arecaceae), 25 November 2006, Tatiya Takaew (**Holotype**: FIH 340). **Isotype**: Doi Suthep-Pui national park, Huay Kog Ma, on leaflets of *Wallichia siamensis* Becc. (Arecaceae), 29 April 2007, Iman Hidayat and Jamjan Meeboon (FIH 356).

Host: On living leaflets of *Rhapis* sp.

Distribution: Only known from the type locality.

Notes: This apiosporous fungus belongs to genus *Apiosporina* due to immersed and erumpent ascomata, bitunicate asci and apiosporous ascospores (Barr, 1989; Sivanesan, 1984). This genus is commonly associated with necrosis of leaves and black knot of twigs (Arx, 1987). Five species, namely, *Apiosporina collinsii* (Schwein.) Höhn., *A. coronata* (Speg.) Bat., *A. fallax* Petr., *A. harunganae* Hansf., and *A. morbosa* (Schwein.) Arx, have been recorded worldwide (Index Fungorum database). However, none of them are associated with plant family Arecaceae, therefore, this fungus is proposed as a new species.

Copyright[©] by Chiang Mai University All rights reserved Astrosphaeriella caryotae Hidayat & To-anun, sp. nov.

(Fig. 2.28)

Ascomata 103–130.5 μ m diam, 44-86 μ m alta, immersa, solitaria, subglobosa, ostiolata. Asci 51.5-73.5 \times 6-8.5 μ m, bitunicata, 8-spori, clavata, pedunculati, praediti Ascosporae 14.5-22 \times 3.5-5 μ m, fusiformis, hyalinae, bicellulares et appendicum utrinque.

Etymology: In reference to the host genus, *Caryota*.

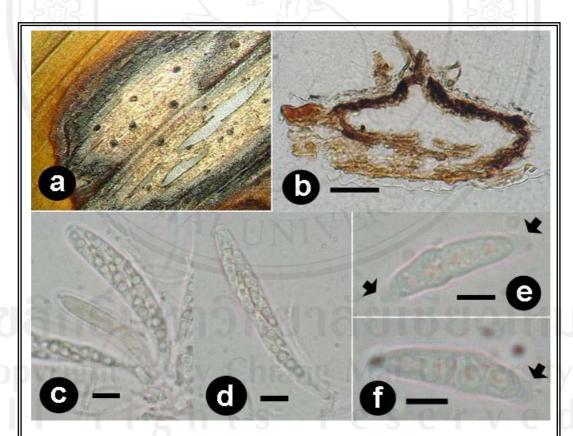


Fig. 2.28 Micrographs of *Astrosphaeriella caryotae* **sp. nov.** (from holotype). a. Symptom. b. Vertical section of ascoma. c-d. Asci. e-f. Ascospores (Appendages showed by arrows). Bars: $b = 40 \mu m$; c, $d = 7.5 \mu m$; e, $f = 4 \mu m$.

Ascomata immersed, solitary, scattered, visible as blackened dots, in section 103-130.5 μm in diameter, 44-86 μm high, subglobose, ostiole central. *Peridium* 4.5-7.5 μm in diameter, comprising of 3-4 layers, comperssed, dark brown, textura angularis cells. *Pseudoparaphyses* 1-2 μm wide, numerous, hypha-like, cellular, anastomosing between asci. *Asci* 51.5-73.5 × 6-8.5 μm, bitunicate, 8-spored, clavate, short pedicellate. *Ascospores* 14.5-22 × 3.5-5 μm, overlapping biseriate, fusiform, 1-septate, euseptate, constricted at the septum, upper cell broader than the lower, hyaline, with pads-like appendages at both ends.

Material examined: THAILAND, Chiang Mai, Chiang Mai University garden, on living leaflets of *Caryota mitis* Lour. (Arecaceae), 29 October 2005, Tatiya Takaew (**Holotype**: FIH 320).

Host: On living leaflets of *Caryota mitis*.

Distribution: Only known from the type locality.

Notes: Species of *Astrosphaeriella* Syd. & P. Syd. are recognized as asaprobes inhabit various decaying plants. On family Arecaceae, *Astrosphaeriella fronsicola* J. Fröhl. & K. D. Hyde on *Oraniopsis appendiculata* (F. M. Bailey) J. Dransf., A. K. Irvine & N. W. Uhl, is the only species reported from leaf spots of palms (Fröhlich and Hyde, 1995a). The novel species is easily recognized and differentiated from *A. fronsicola* by having appendages at both ascospores ends and being recorded from *Caryota mitis*.

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Astrosphaeriella palmicola Hidayat & To-anun, sp. nov.

(Fig. 2.29)

Ascomata 455-725 µm diam., 615-910 µm, superficialis, solitaria, nigra,

conicalia, ostiolata; Asci 75-90×x 7.5-10 μm, clavati, 8-spori, bitunicati, pedicellati;

Ascospori 24-28 \times 2.5-4.5 μ m, biseriati, fusiformis, 1-3-septata, hyalina,

nonappendiculati.

Etymology: Refers to its host common name, palm.

Ascomata superficial, solitary, black, in section 455-725 µm diam., 615-910

μm, conical, base flattened, with rounded and central ostiole. *Peridium* thick and

carbonized, base poorly developed. Pseudoparaphyses ca 0.5 µm, numerous,

filamentous, hyaline, anartomosing between asci. Asci 75-90 × 7.5-10 μm, clavate, 8-

spored, bitunicate, pedicellate, with rounded apex, with an ocular chamber.

Ascospores 24-28 \times 2.5-4.5 µm, overlapping biseriate, narrowly fusiform, 1-3-septate,

hyaline, lacking a mucilagenous sheath.

Material examined: THAILAND, Chiang Mai, A. Pa pae, Bahn Phadeng,

Mushroom Research Centre, on leaflets of Areca catechu L. (Arecaceae), 5

September 2005, Iman Hidayat (Holotype: FIH 084).

Host: On leaflets of Areca catechu.

Distribution: Only known from the type locality.

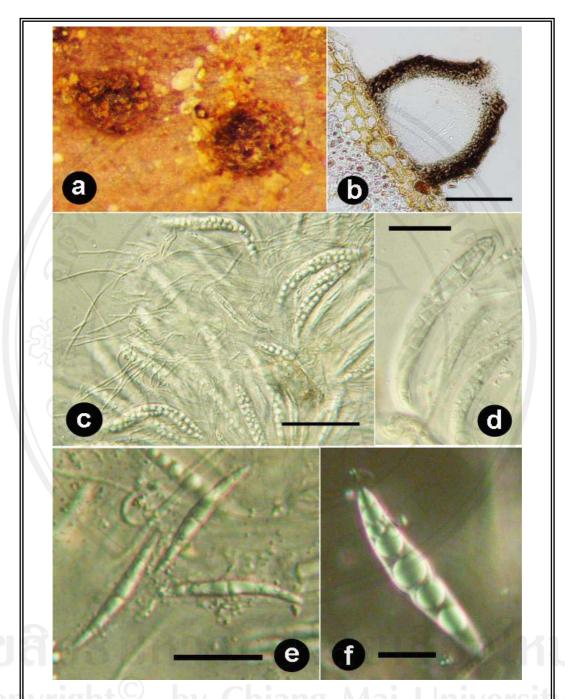


Fig. 2.29 Micrographs of *Astrosphaeriella palmicola* **sp. nov.** (from holotype). a. Ascomata appearance on the host surface. b. Vertical section of ascoma. c. Asci and pseudoparaphyses. d. Ascus. e-f. Ascospores. Bars: $b = 500 \ \mu m$; $c = 50 \ \mu m$; $d = 25 \ \mu m$; $e = 15 \ \mu m$; $f = 5 \ \mu m$.

Notes: According to key of *Astrosphaeriella* Syd. & P. Syd. from palms by Hyde and Fröhlich (1997), *Astrosphaeriella palmicola* sp. nov. is closed to *A. minima*

Aptroot in having ascospores lacking of mucilagenous sheath, hyaline and the size shorter than 30 μ m. However, the novel species is distinct and easily distinguished from *A. minima* by having superficial and carbonaceous ascomata, thin ascospores (2.5-4.5 μ m vs 7-8 μ m of *A. minima*) and being recorded from palm genus *Areca* (*A. minima* is only known from Bamboo) (Hyde and Fröhlich, 1997).

Fasciatispora ujungkulonensis Hidayat, Mycotaxon 102: 350 (2007)

(Fig. 2.30)

Ascomata immersa, circumglobata, clypeata, apparatus leviter elatus et ostiolatus, 115–300 μ m diam,., 50–120 μ m alta, subglobosa, basi planissimus, perithecium, medium ostiolatum. Peridium 7.5–15 μ m crassum. Paraphysae 1–2.5 μ m, numerosae, filamentae, septatae et hyalinae. Asci 43–81 × 4.5–9.5 μ m, unitunicati, 8-spori, cylindrici, apicis rotundi, apicale apparatu discoidae, J+, 2–2.5 μ m diam, I–1.5 μ m alta praediti. Ascosporae 6–12 × 4.5–7.5 μ m, uniseriatae, ellipsoideae, aseptatae, brunneae, fascia aequatoris pallidus vel latus, tunica gelatinosa praeditae.

Etymology: Refers to the place where the specimen was collected.

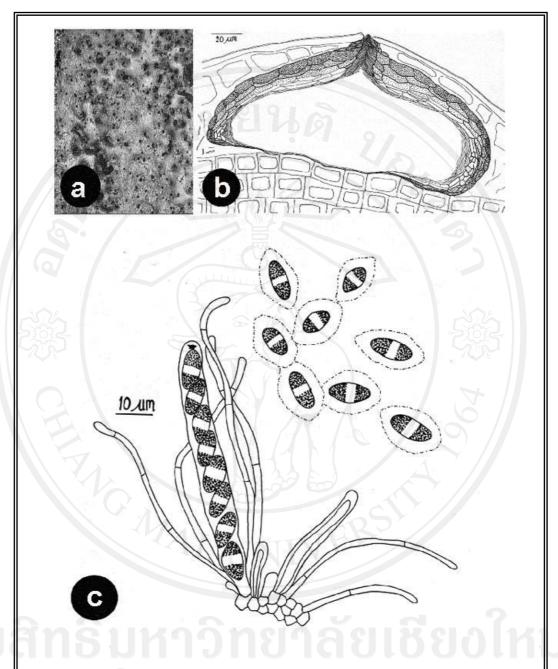


Fig. 2.30 Line drawings of *Fasciatispora ujungkulonensis* (from holotype). a. Ascomata appearance on the host surface. b. Vertical section of ascoma. c. Ascus and ascospores.

Ascomata immersed, clustered, clypeate, visible as a slightly raised, ostiolar, dome-shaped areas on the host surface, in section 115–300 μ m diam., 50–120 μ m high, subglobose, with a flattened base, and only covered by epidermal tissue of the

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host, with a central ostiole. *Peridium* 7.5-15 μ m thick, composed inwardly of 1–2 layers of compressed, hyaline cells, and externally of 2–3 layers of compressed, angular brown-walled cells. *Paraphyses* 1–2.5 μ m wide, numerous, filamentous, septate, hyaline. *Asci* 43–81 \times 4.5–9.5 μ m, unitunicate, 8-spored, cylindrical, short pedicellate, apically rounded, with J+, and non-refractive discoid subapical ring, 2–2.5 μ m diam \times 1–1.5 μ m high. *Ascospores* 6–12 \times 4.5–7.5 μ m, overlapping uniseriate, ellipsoidal, aseptate, brown with a wide equatorial pallid band, and surrounded by a wide mucilaginous sheath, ca 2.5–10 μ m from the ascospore proper.

Specimen examined: INDONESIA, West Java, Tanjung Lame, Ujung Kulon National Park, on decaying fronds of *Caryota* sp. (Arecaceae), 29 March 2006, Dewi and Iman Hidayat (**Holotype**: FIH 195) (MRC Herbarium 00630).

Host: On decaying fronds of *Caryota* sp.

Distribution: Indonesia.

Notes: Fasciatispora ujungkulonensis is similar to F. petrakii in being a terrestrial saprobe on palms and in having ascospores shorter than 14 μ m (Hyde, 1995b). Ascospores of F. ujungkulonensis, however, are distinct from those of F. petrakii in being ellipsoidal with blunt ends (6–12 × 4.5–7.5 μ m); those of F. petrakii (10–14 × 5–5.8 μ m) are ovoid to almost rhomboid with acute ends (Hyde, 1995b). Furthermore, a mucilaginous sheath which surrounds the ascospores of F. ujungkulonensis is relatively wide (ca 2.5–10 μ m from the ascospore proper), compared with that of F. petrakii which is fairly thin. Asci of F. ujungkulonensis are also shorter and narrower than those of F. petrakii (43–81 × 5–9.5 μ m vs 84–94 × 8–10 μ m of F. petrakii) (Hyde, 1995b). Fasciatispora ujungkulonensis is also distinct

from F. bicincta (Syd. & P. Syd.) K. D. Hyde in having a discoid ascus ring; that of F. bicincta is wedge-shaped. Besides F. bicincta has larger ascospores (14–20 \times 8–12 μ m) and larger asci (75–100 \times 12–20 μ m). A synopsis of characters of F. ujungkulonensis, F. petrakii and F. bicincta is given in Table 1. Numerous attempts were made to grow single spore isolates of F. ujungkulonensis in culture, but each time they proved unsuccessful.

Key to Fasciatispora species (modified from Hyde, 1995b):

1a. Ascal ring J F. melanosticta	
1b. Ascal ring J+	2
2a. Marine saprotrophs on palms or other timber	3
2b. Terrestrial saprotrophs on palms	4
3a. Ascospores with polar cap-like appendages	. F. nypae
3b. Ascospores surrounded by a mucilagenous sheath	F. lignicola
4a. Ascospores mostly shorter than 14 μm	S
4b. Ascospores mostly longer than 14 μm	University
5a. Ascospores ellipsoidal, asci 43–81 × 4.5–9.5 μm	F. ujungkulonensis
5b. Ascospores ovoid to rhomboid, asci 84–94 × 8–12 µm	F. petrakii

6a. Ascospores pale brown, ellipsoidal to ovoid, with a swollen equatorial pallid
band
bicincta
6b. Ascospores brown to dark brown, ellipsoidal, without a swollen equatorial pallid
band

7a. Ascospores dark brown, asci $90-118 \times 15-19 \, \mu \text{m} \dots$ *F. pandanicola* 7b. Ascospores brown, asci $120-170 \times 12-16 \, \mu \text{m} \dots$ *F. sabalicola*

Guignardia arengae, Hidayat & To-anun, sp. nov. (Fig. 2.31)

Differt a Guignardia candeloflamma asci decretus, $47.5-65 \times 15-22.5 \mu m$, ascosporae uniappendicum, et G. cocogena asci clavatus et multiseriati.

Etymology: Named after its host generic name, Arenga.

Stromata lacking. Ascomata immersed, black, developing under slightly raised regions, in vertical section 120-165 μ m diam., 120-145 μ m high, subglobose, with a central ostiole. Peridium 20-30 μ m thick, composed of brown-walled textura globulosa, inwardly compressed. Pseudoparaphyses 2-4 μ m wide, cellular, forming chains-like structure, hyaline. Asci 35-55 \times 10-17.5 μ m, bitunicate, 8-spored, broadly clavate, thick-walled, short-pedicellate, with ocular chamber near the apex, apex

subtruncate. *Ascospores* 12-17.5 \times 5-7.5 μ m, overlapping 3-multi-seriate, hyaline-greenish, ovoid, lacking germ pores, with broader part above the middle part of the spores, 1-celled, not smooth, with distinct appendages at the basal ends.

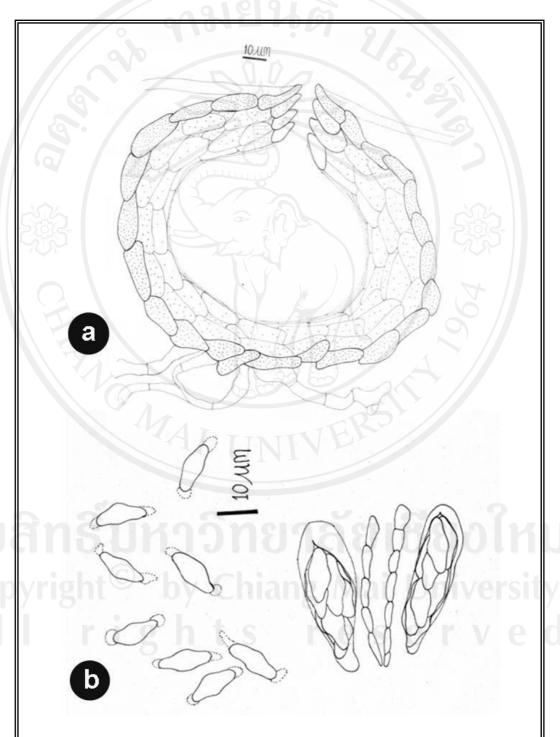


Fig. 2.31 Line drawings of *Guignardia arengae* **sp. nov.** (from holotype). a. Vertical section of ascoma. b. Asci and ascospores.

Material Examined: THAILAND, Chiang Mai Province, Doi Suthep-Pui National Park, Hua Kog Ma, on leaflets of *Arenga hookeriana* (Becc.) Whitmore (Arecaceae), 16 October 2005, Iman Hidayat, FIH 407 (**Holotype**). **Isotype**: *ibid*, on leaflets of *Livistona chinensis* R.Br. (Arecaceae), 26 october 2006, Iman Hidayat (FIH 327).

Host: On leaflets of *W. siamensis* and *L. chinensis*.

Distribution: Only known from the type locality.

Notes: Two species of *Guignardia* that having bipolar appendages have been reported from palms, viz, *G. cocogena* (Cooke) Punith. and *G. candeloflamma* J. Fröhl. & K. D. Hyde (Hyde, 1995a). However, this specimen is distinct from *G. candeloflamma* based on the characteristics of appendages and smaller asci (35-55 × 10-17.5 μm vs 91-140 × 17.5-25 μm *G. candeloflamma*) and ascospores (12-17.5 × 5-7.5 μm vs 17.5-25 × 7.5-11 μm of *G. candeloflamma*) (Hyde, 1995a). The present novel species also differs from *G. cocogena* in having broadly clavate asci and multiseriate ascospores arrangements inside the asci while the later species having cylindric-clavate asci and uni to 2-seriate ascospores arrangement inside the asci (Punithalingam, 1974).

Linocarpon nonappendiculatum Hidayat & To-anun, sp. nov.

(Fig. 2.32)

Differt a Linocarpon elaeidis, L. carinisporium et L. livistonae, ascospori non appendiculati et implicatus cum Arecca catechu

Etymology: Refer to the absence of appendages on its ascospores.

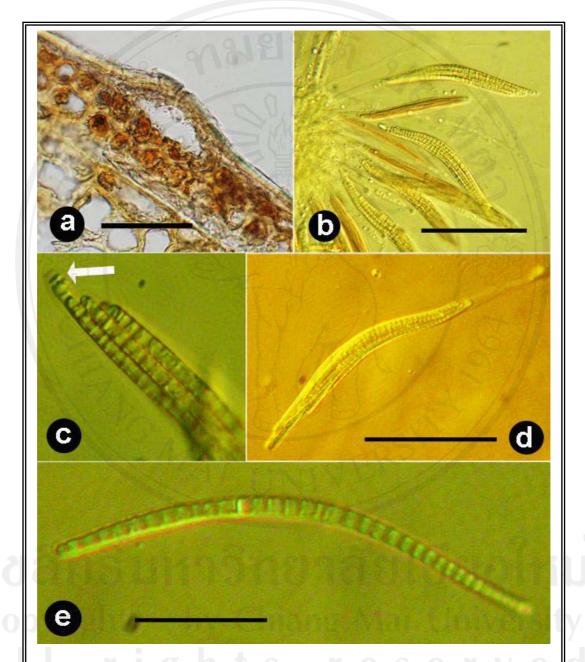


Fig. 2.32 Micrographs of *Linocarpon nonappendiculatum* **sp. nov.** (from holotype). a. Vertical section of ascoma. b. Asci and paraphyses. c. Ascus ring (showed by arrow). d. Ascus. e. Ascospore. Bars: $a=250~\mu m$; $b=125~\mu m$; $d=75~\mu m$; $e=25~\mu m$.

Ascomata immersed beneath a clypeus, solitary, in section 262.5-300 μm diam., 75-125 μm high, subglobose to lenticular, with a central ostiole. *Peridium* 12.5-17.5 μm thick, outwardly comprising of several layers of compressed, ellipsoidal, brown-walled cells, inwardly composed of 1-2 layers of hyaline cells,. *Paraphyses* 0.5-2.5 μm, numerous, filamentous, hyaline, anastomosing between asci. *Asci* 137.5-187.5 \times 12.5-13.5 μm, unitunicate, 8-spored, clavate, long pedicellate, J- subapical ring. *Ascospores* 85-97.5 \times 2.5-3 μm, multiseriate, filiform, with refringent bandslike septum, hyaline, lacking of appendages at both ends.

Material examined: THAILAND, Chiang Mai, A. Pa pae, Bahn Phadeng, Mushroom Research Centre, on rachis of *Areca catechu* L. (Arecaceae), 5 September 2005, Iman Hidayat FIH 082 (Holotype). Isotype: *ibid.*, FIH 083.

Host: On rachis of *Areca catechu*.

Distribution: Only known from the type locality.

Notes: Linocarpon nonappendiculatum sp. nov. has a similar characteristics with Linocarpon elaeidis Petr, L. carinisporium K. D. Hyde and L. livistonae (Henn.) K. D. Hyde, in asci, and ascospores appearance and size (Taylor and Hyde, 2003). However, the recent novel species is easily distinguishable from those similar species due to lacking of ascospores appendages and being associated with Areca catechu. Because of the present or absent of ascospores appendages as well as the type of appendages are important characteristics in differentiating species within genus Linocarpon, therefore, this fungus is proposed as a new species.

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Lophiostoma livistonicola Hidayat & To-anun, sp. nov.

(Fig. 2.33)

Ascomata 110.5-184.5 μm diam., 123-164.5 μm, immersa, solitaria,

ostiolata; Asci 72-97 × 12.5-15 µm, clavati, 8-spori, bitunicati, pedicellati; Ascospori

 $14.5-17 \times 4-5 \ \mu m$, biseriati, fusiformis, 1-septata, hyalina,cum utrinque apicale

appendiculati.

Etymology: Refers to the generic name of the host, Livistona.

Ascomata immersed, solitary, erumpent, aclypeus, in section 110.5-184.5 µm

in diameter, 123-164.5 µm high, subgloblose, with a slot-like and central ostiole.

Peridium 5-12 µm thick, composed of compressed, brown-walled, ellipsoidal and

angular cells. *Pseudoparaphyses* 1-2.5 µm, filamentous, cellular, hyaline, embedded

in gelatinous matrix. Asci 72-97 × 12.5-15 μm, bitunicate, 8-spored, clavate, short-

pedicellate, Ascospores 14.5-17 \times 4-5 μ m, overlapping biseriate, fusiform, hyaline, 1-

septate, euseptate, constricted at the septum, smooth, with distinct appendages at both

Material examined: THAILAND, Chiang Mai, Chiang Mai University, Palm

Garden, on leaflets of Livistona chinensis R. Br. (Arecaceae), 29 October 2006,

Tatiya takaew (Holotype: FIH 323).

Host: On leaflets of *Livistona chinensis*.

Distribution: Only known from the type locality.

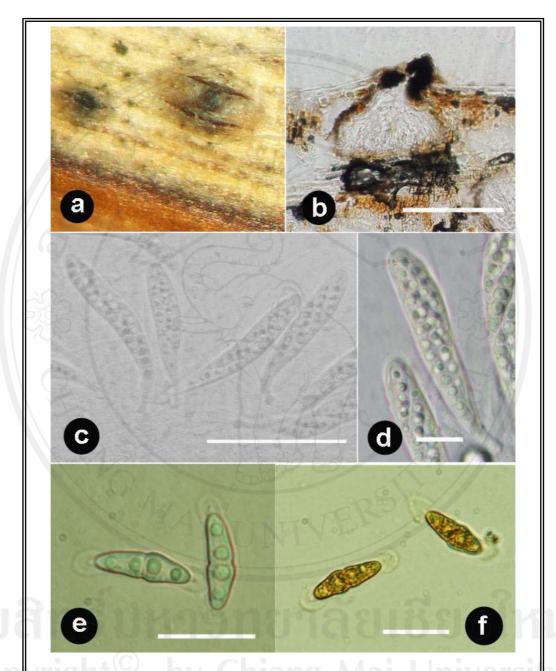


Fig. 2.33 Micrographs of *Lophiostoma livistonicola* **sp. nov.** (from holotype). a. Ascomata appearance on the host surface. b. Vertical section of ascoma. c. Asci and pseudoparaphyses. d. Ascus. e. Ascospores. f. Ascospores under iodine solution. Bars: $b = 125 \ \mu m$; $c = 75 \ \mu m$; $d = 10 \ \mu m$; $e, f = 15 \ \mu m$.

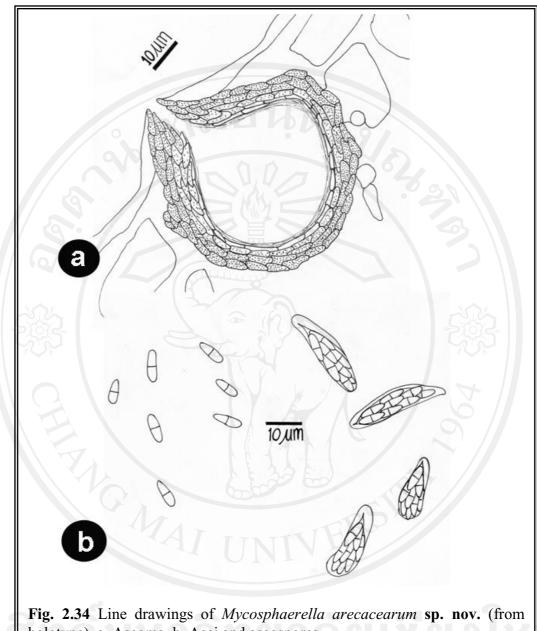
Notes: This specimen belongs to genus *Lophiostoma* Ces. & De Not. due to bitunicate asci and slit-like ostiole of ascomata (Holm and Holm, 1988). *Lophiostoma rubi* (Fuckel) Liew, Aptroot & K. D. Hyde and *L. palmettum* (Cooke) Aptroot & K.D. Hyde are very similar in asci and ascospores dimension (Hyde and Aptroot, 1997; Aptroot, 1998). However, *L. palmettum* is distinct from this novel species due to its multilocular ascomata and very wide asci (74-104 × 28-32 μm) (Hyde and Aptroot, 1997). A new species proposed in this study, *Lophiostoma livistonicola*, is also distinct from *L. rubii* due to the ascospores are ornamented with very distinct appendages at both ends (Aptroot, 1998).

Mycosphaerella arecacearum Hidayat and To-anun, sp. nov.

(Fig. 2.34)

Ascomata 45-70 μ m diam., 43-75 μ m alta, immersa, aggregata, globosa, ostiolata. Asci 15-25 \times 3.75-6 μ m, bitunicati, 8-spori, obclavati, apedunculati, praediti Ascosporae 5-8.5 x 1.5-3 μ m, ellipsoidae, bicellularis, asymmetrica, hyalinae, non-appendicum.

Etymology: Named after its genus name of host. *Areca*.



holotype). a. Ascoma. b. Asci and ascospores.

Leaf spots amphigenous, irregular, lesions brown. .Ascomata visible as black ostiolar dots on the host surface, immersed, becoming erumpent, 6-8 aggregated or in groups, amphigenous, subepidermal, in section very small, 45-70 µm diam., 43-75 µm high, globose, pseudothecial, with central ostiole, periphysate. Peridium 7.5-10 μm

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thick, composed of 4-6 layers of medium brown, polygonal cells, become hyaline inwardly. *Pseudoparaphyses* absent. *Asci* 15-25 \times 3.75-6 μ m, bitunicate, 8-spored, obclavate, apically rounded, apedicellate. *Ascospores* 5-8.5 \times 1.5-3 μ m, biseriate overlapping, 1-septate, slightly constricted at the septum, ellipsoidal, unequally bicelled, with obtuse ends, constricted at the septum, hyaline, thin-walled, straight to slightly curved, tapering towards the lower end, lacking a mucilagenous sheath.

Material examined: THAILAND, Payao, on leaflets of *Areca catechu* L. (Arecaceae), 17 September 2007, Iman Hidayat (**Holotype**: FIH 102).

Host: Leaflets of Areca catechu.

Distribution: Only known from the type locality.

Notes: Recently, none of *Mycosphaerella* species have been reported associated with plant genus *Areca* (Aptroot, 2006). The new species, *Mycosphaerella* arecacearum, is easily recognised by having obclavate, apedicellate asci with small ascospores as well as being associated with leaflets of *Areca catechu*.

Mycosphaerella borassi Hidayat and To-anun, sp. nov.

(Fig. 2.35)

Ascomata 10-60 μ m diam., 70-120 μ m alta, immersa, aggregata, subglobosa, ostiolata. Asci 40-50 \times 3.75-6 μ m, bitunicati, 8-spori, obclavati vel saccati, pedunculati, praediti Ascosporis 8.5-12 \times 2-3 μ m, ellipsoidis, bicellularis, asymmetrica, hyalinae, cum uni appendicum.

Etymology: Refers to generic name of its host, Borassus.

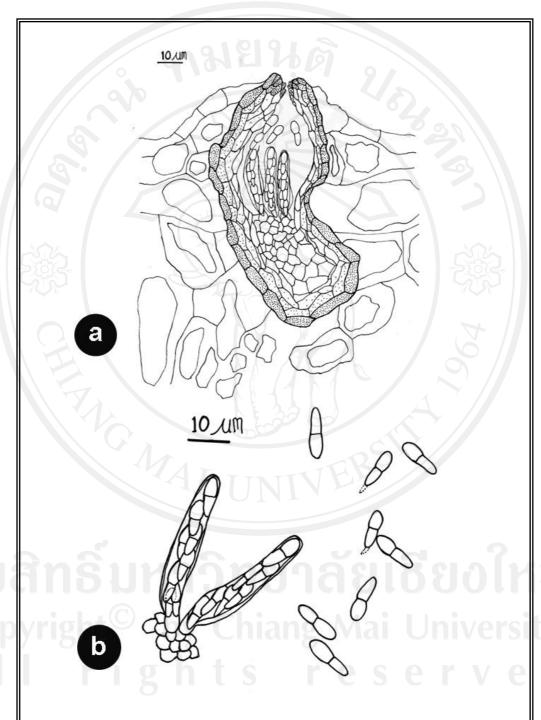


Fig. 2.35 Line drawings of *Mycosphaerella borassi* **sp. nov.** (from holotype). a. vertical section of ascoma. b. Asci and ascospores.

Leaf spots amphigenous, irregular, lesions brown. Ascomata visible as black ostiolar dots on the host surface, immersed, becoming erumpent, 6-8 aggregated or in groups, amphigenous, subepidermal, in section very small, 10-60 μm diam., 70-120 μm high, subglobose, pseudothecial, with central ostiole, periphysate. Peridium 5-14 μm thick, composed of 2-3 layers of medium brown, polygonal cells, become hyaline inwardly. Pseudoparaphyses absent. Asci 40-50 × 3.75-6 μm, bitunicate, 8-spored, obclavate to saccate, apically rounded, short pedicellate. Ascospores 8.5-12 x 2-3 μm, biseriate overlapping, 1-septate, slightly constricted at the septum, ellipsoidal, unequally bicelled, with obtuse ends, constricted at the septum, hyaline, thin-walled, straight to slightly curved, tapering towards the lower end, surrounding by a thin mucilagenous sheath, often with basal appendage.

Material examined: THAILAND, Payao, on lamina of *Borassus flabellifer* L. (Arecaceae), 27 May 2007, Iman Hidayat and Jamjan Meeboon (**Holotype**: FIH 059).

Host: On lamina of Borassus flabellifer.

Distribution: Only known from the type locality.

Notes: The genus *Mycosphaerella* is widely recognized as plant pathogen, although some species are also recognized as saprobes on various plants (Aptroot, 2006). Presently, none of *Mycosphaerella* species have been reported associated with plant genus *Borassus*. According to the Aptroot's key (2006), *Mycosphaerella borassi* belongs to section *Fusispora*, however, this novel species is distinct from the species within the section *Fusispora* by having very small ascospores (< 12 μm long). Therefore, we proposed this specimen as a new species.

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Oedocephalum formosus Hidayat & To-anun, sp. nov.

(Fig. 2.36)

Coloniae brunneae; Mycelia plerumque immersa, ramosa, hyalina, septata; Conidiophora $105-160 \times 2-7 \, \mu m$, solitaria, cylindricum, simplicia, hyalina; Ampulla $25-30 \, \mu m$ in diameter, globosa, in pagina integra conidiifera, basi septata; Cellulae conidiogenae $1.5-2 \, \mu m$ diam., $1-1.5 \, \mu m$ high, denticulatae, holoblasticae, schizolyticae; Conidia $32-40 \times 14-16 \, \mu m$, solitaria, ellispoida, aseptata, laevia, brunnea.

Etymology: Refers to its beautiful appearances.

Colonies on the natural substratum scattered, hyaline with dark-brown conidial mass at the apical part. *Mycelium* immersed, branched, hyaline, septate. Conidiophores $105-160 \times 2-7 \mu m$, solitary, cylindrical, erect, simple, hyaline, septate only present between ampulla and conidiophore. Ampulla 25-30 μm in diameter, globose, hyaline, solitary, terminal, with a septate at the basal part. Conidiogenous cells $1.5-2 \mu m$ diam., $1-1.5 \mu m$ high, denticulate, with pimple-shape denticles. Conidia secession blastic, holoblastic, schizolytic. Conidia $32-40 \times 14-16 \mu m$, solitary, ellipsoidal, 1-celled, brown, smooth.

Material examined: THAILAND, Payao, on seed of *Borassus flabellifer* L. (Arecaceae), 27 May 2007, Iman Hidayat and Jamjan Meeboon (**Holotype**: FIH 127).

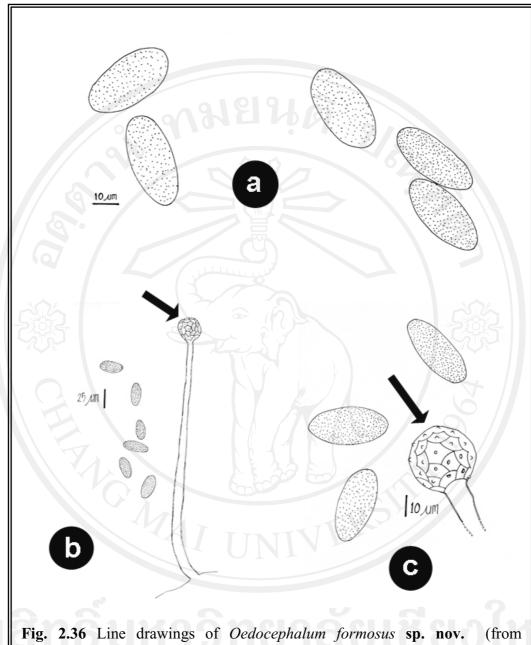


Fig. 2.36 Line drawings of *Oedocephalum formosus* **sp. nov.** (from holotype). a. Conidia. b. Conidia and conidiophores (ampula showed by arrow). c. Ampula (showed by arrow) and conidia.

Host: On seed of *Borassus flabellifer*.

Distribution: Only known from its type locality.

Notes: The novel species is easily distinguished from other species of Oedocephalum by having pimple-shape denticles, aseptate conidiophores, as well as brown and large conidia (> 30 μ m).

Oxydothis cyrtostachicola Hidayat & To-anun, Fungal Diversity 23: 164 (2006)

(Fig. 2.37)

Ascomata 130–155 μ m diam, 20–34 μ m alta, immersa, subglobosa, ostiolata. Asci 102–120 × 12–13 μ m, 8-spori, pedunculati, aparatu apicale J–, praediti Ascosporae 48–52 × 5–6 μ m, hyalinae, fusiformis, bicellulares.

Etymology: In reference to the host genus, Cyrtostachys.

Ascomata forming under slightly raised, ellipsoidal regions on the host surface, black border, solitary or in groups 2–3; in section immersed, subglobose, ostiole eccentric, long axis horizontal to that of the host surface with neck at one end, forming ca 130–155 μm diam. × 20–34 μm high. Peridium comprised of 2–3 layers outer layers of oblong, dark-brown cells and with an additional inner layer of oblong, hyaline cells. Paraphyses deliquesce early. Asci 102–120 × 12–13 μm, 8-spored, unitunicate, clavate, short pedicellate, J-, refractive subapical ring, has a canal leading to the apex. Ascospores 48–52 × 5–6 μm, fusiform, 1-septate, hyaline, tapering gradually from the central septum to pointed processes, without spine-like form.

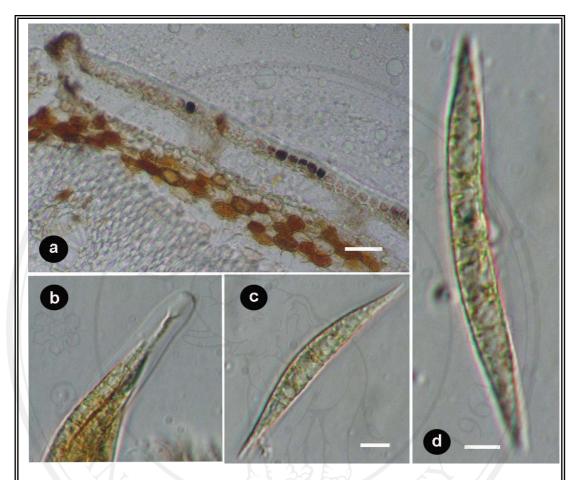


Fig. 2.37 Micrographs of *Oxydothis cyrtostachicola* (from holotype). a. Vertical section through the ascoma. b. Asci and ascal ring. c-d. Ascospores. Bars: a=20 μm ; c-d = 5 μm .

Material examined: THAILAND, Chiang Mai, Chiang Mai University garden, on petioles of *Cyrtostachys renda* Blume (Arecaceae), 30 October 2005, Iman Hidayat, FIH 151 (**Holotype**: MRC 0007). **Isotype**: *ibid*., HKU (M) 17170.

Host: Saprobic on petioles of *Cyrtostachys renda*.

Distribution: Northern Thailand.

Notes: The morphology and molecular elucidation of this species is presented in chapter 3.

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Terriera livistonae Hidayat & To-anun, sp. nov.

(Fig. 2.38)

Differt a Terriera nematoidea et T. asteliae paraphysae ramosa et asci

amplius (78.5-92 \times 3.5-5 μ m).

Etymology: Refers to its generic host name, Livistona.

Ascomata 2-4 mm long, 0.25-0.75 mm high, superficial, lip-like cells, lacking

of zones border surrounded the ascomata, in section triangular in shape, with central

ostiole, 110-160 µm in diameter, 95-160 µm in height. Peridium 14.5-29.5 µm,

composed of several layers, slight thick walls, dark brown walls. *Paraphyses* 1–1.5

um in diameter, hypha-like, branched near apex slightly sowllen, hyaline, septate.

Asci 78.5-92 × 3.5-5 μm, unitunicate, 8-spored, long cylindrical with subtruncate

apex, short pedicellate, with a J- apical ring. Ascospores 66-73 × 1 μm, filiform,

aseptate, hyaline, lack of gelatinous sheath.

Material examined: THAILAND, Chiang Mai, Chiang Mai University

garden, on living leaflets of Livistona chinensis R.Br. (Arecaceae), 16 October 2005,

Iman Hidayat (Holotype: FIH 133).

Host: On living leaflets of Livistona chinensis.

Distribution: Only known from the type locality.

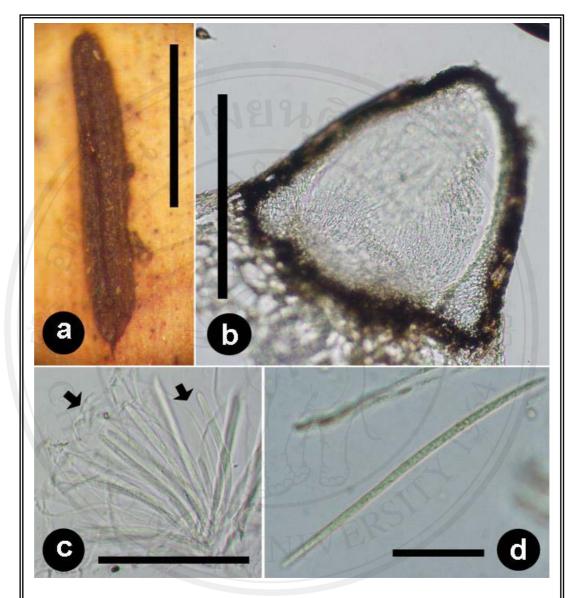


Fig. 2.38 Micrographs of *Terriera livistonae* **sp. nov.** (from holotype). a. Ascoma appearances on the host surface. b. Vertical section of ascoma. Asci and paraphyses (arrows). d. Ascospore. Bars: a=2 mm; b=100 μ m; c=75 μ m; d=25 μ m.

Notes: This species belongs to genus *Terriera* due to lip cells of ascomata do not lining with the ascomata opening, well differentiated paraphyses, and ascospores lack of gelatinous sheats. *Terriera nematoidea* (P. R. Johnst.) P. R. Johnst., and *T. asteliae* (P. R. Johnst.) P. R. Johnst.) P. R. Johnst. were reported as *Terriera* species that

characterized by subtruncate apex of asci (Johnston, 2001). However, this species differs from T. nematoidea and T. asteliae in having branched paraphyses near the apex and longer asci (> $70\mu m$).

Venturia frondicola Hidayat & To-anun, sp. nov.

(Fig. 2.39)

Differt a Venturia populina ascosporae hyalinae, verruculosae et implicare cum palmae.

Etymology: In reference to the substrate where this species was collected.

Ascomata superficial, erumpent, black, solitary, in section 94 μm in diameter, 78 μm in height, subglobose, brown to dark-brown, ostiole central, papillate, ornamented with setae around ostiole. Peridium 10-15 μm thick, comprising several layers, compressed, angular to ellipsoidal, brown to dark brown cells. Pseudoparaphyses ca 1.25 μm, trabeculate, numerous, cellular, hyaline. Asci 85-115 × 12.5-16.25 μm, bitunicate, 8-spored, cylindrical, with ocular chamber near the apex, short pedicellate, apically rounded. Ascospores 17.5-22.5 × 7.5-10 μm, apiosporous, 1-septate, verrucose, hyaline, biseriate, constricted at the medium or submedium septum, upper cell wider and longer than lower cell.

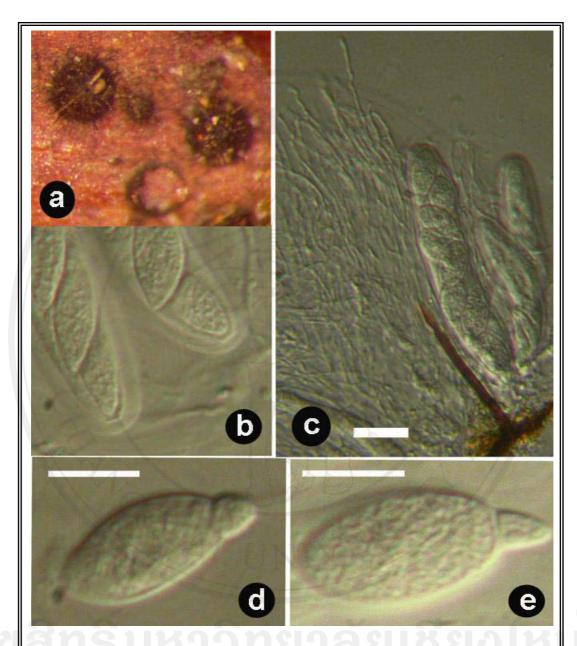


Fig. 2.39 Micrographs of *Venturia frondicola* **sp. nov.** (from holotype). a. Ascomata appearance on the host surface. b-c. Asci and pseudoparaphyses. d-e. Ascospores Bars: $c = 15 \mu m$; d, $e = 8 \mu m$.

Material examined: THAILAND, Chiang Mai, T. Pa Pae, Bahn Pha Deng, Mushroom Research Centre, on rachis of *Areca catechu* L. (Arecaceae), 5 September 2005, Iman Hidayat (**Holotype**: FIH 082).

Host: On petioles of Areca catechu.

Distribution: Only known from the type locality.

Notes: This novel apiosporous fungus belongs to family Venturiaceae. The superficial ascomata, apiosporous cell, and bitunicate asci of this specimen are closed to genus *Apiosporina* (Barr, 1989; Sivanesan, 1984). However, the setose ascomata of this specimen is ditinct from the genus *Apiosporina* which is not lacking of setae. Barr (1989) considered setose and superficial ascomata as important characters in differentiating between genera of *Apiosporina* and *Venturia*. The later genus belongs to family Venturiaceae that characterized by having superficial setose ascomata and apiosporous ascospores (Sivanesan, 1984). The present new species, *Venturia frondicola*, is closed to *V. populina* (Vuill.) Fabric. based on the key to *Venturia* species (Sivanesan, 1984), however, the novel species differs in having hyaline and verruculose ascospores as well as being recorded from palms petioles.

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