

APPENDIX

Appendix 1 Wild rice survey sheet

- Surveyor.....  
Date of survey.....
1. Location no..... Local name of wild rice.....  
2. Lat/Lon or UTM.....  
3. Address..... city..... province.....  
4. Map.....

5. Habitat  
 In side rice field     Edge rice field     By road side     Abandoned
6. Area.....m<sup>2</sup>/km<sup>2</sup>     patchy     all covered
7. Awn color     white     red
8. Awn length     0-5 cm.     5-10 cm.     >10 cm.
9. Growth stage     anthesis     milking stage     grain filling     seed maturity
10. Plant type     erect     prostrate
11. Stigma color     white     dark purple     other.....
12. Anther length     1/3 of spikelet     1/2 of spikelet     same of spikelet
13. Type     annual     perennial     annual-perennial intermediate
14. Hull color     black     black with stripe     straw     other.....
15. Special traits.....

**Appendix 2 Descriptor state and Meaning of morphological and other characters of common wild and cultivated rice**

<b>Character No. and Name</b>	<b>Descriptor state and Meaning</b>
1. G.S. No.	
2. Species name :	
3. Designation :	
4. Location :	
5. Growth habit : 1= erect; 3= semi-erect; 5= intermediate; 7= decumbent	
6. Leaf sheath color : 1= green; 2= dark green; 3= light purple; 4= purple	
7. Leaf blade color : 1= green; 2= dark green; 3= purple at margin; 4= purple line; 5= all purple	
8. Tillers: number of tiller per plant at full vegetative stage	
9. Ligule color : 1= colorless 2= purple	
10. Ligule shape : 1= 1clip 2= 2 clips 3= truncate	
11. Auricle color : 1= colorless 2= purple	
12. Internode color : 1= green 2= light green 3= purple line 4= purple	
13. Flowering date (50%) or Maturity day	
14. Spikelet color : 1= straw 2= brown 3= yellow 4= red 5= black	
15. Apiculus color : 1= colorless 2= brown 3= yellow 4= red 5= black	
16. Anther length : 1= $\frac{1}{4}$ spikelet 2= $\frac{1}{2}$ spikelet 3= $\frac{3}{4}$ spikelet 4= full or equal to spikelet.	
17. Stigma color : 1= colorless 2= red 3= purple 4= black	
18. Stigma exertion : 1= enclosed 2= red 3= well exerted	
19. Panicle type : 1= compact 2= intermediate 3= open	
20. Awn color : 0= no awn 2= white 3= red	
21. Awn length : 0= no awn 1= 0-5cm 2= 5-10cm 3= >10cm	
22. Pericarp color : 1= white 2= red	

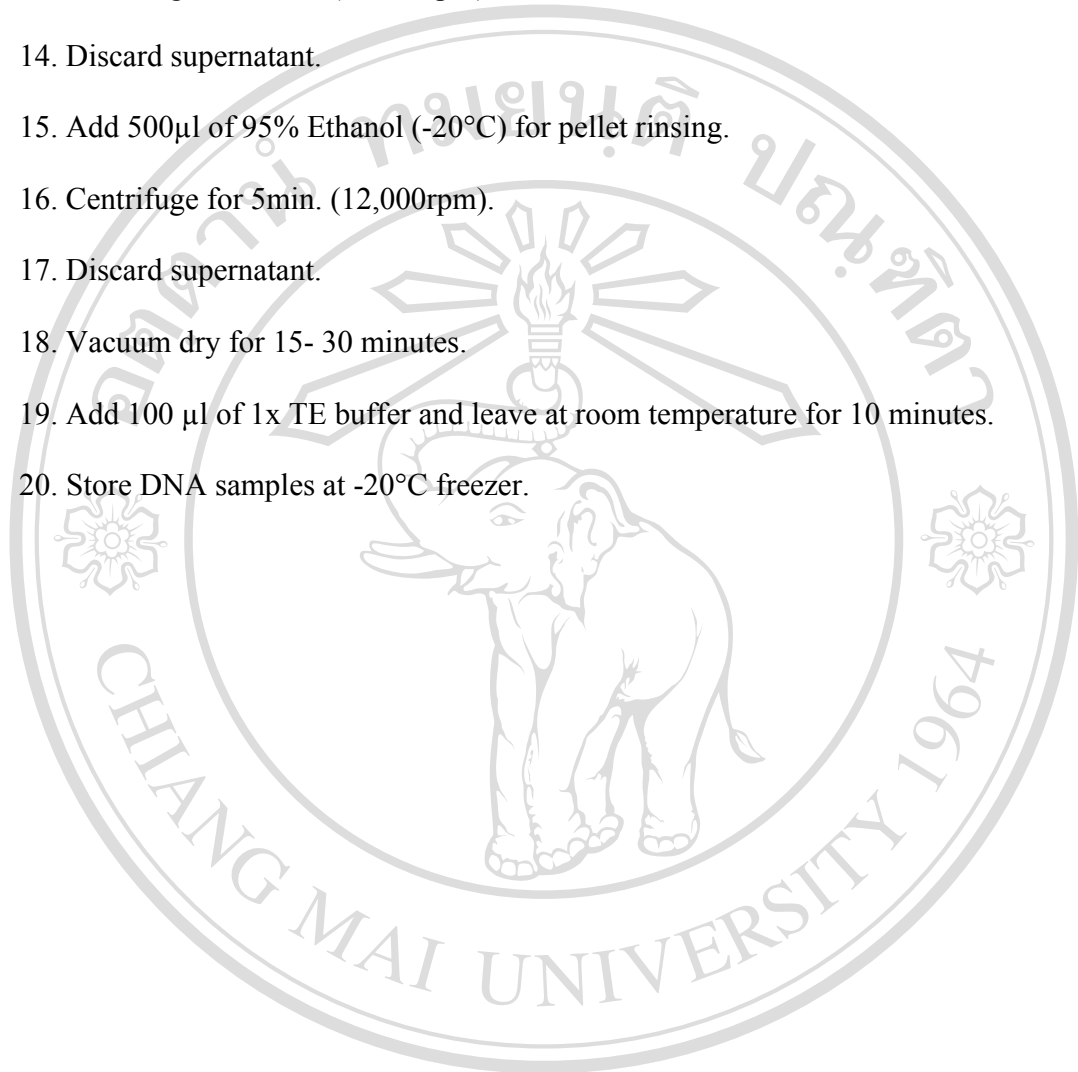
### Appendix 3 DNA extraction

#### 1. Extraction buffer:

DNA extraction	Stock	10 ml
4% CTAB	100%	0.4g
100 mM Tris – Hcl pH 8.0	1M	1ml
20 mM EDTA pH 8.0	0.5 M	0.4ml
1.4 M NaCl	5 M	2.8 ml
0.4% β-mercapto-ethanol	100%	40 µl
H <sub>2</sub> O	100%	6ml

2. Extraction buffer 1ml into 0.1g of grinded leaf sample.
3. Vortex mixed ~ 15 – 30 (second) each tube.
4. Incubate 3 hours in 65°C of water bath and inverting every 30 min.
5. Centrifuge 5 min at 12000 rpm.
6. Supernatant transferring to the new tube and put Chloloform-isoamyl alcohol (24:1) (500µl). Gently inverting about 5min, and centrifuge at 12000 rpm for 20min.
7. Carefully transfer 500µl of the top aqueous phase (3 layer) into the new tube and then add (100 µl) with Sodium-acetate (3M NaoAC).
8. Add 600µl ice-cold Isopropanal Alcohol (-20°C) into the Supernatant and inverting every tube until homogeneous together.
9. Keep in the freezer (-20°C) overnight or (-70°C) at least 20 minutes.
10. Centrifuge for 10 minutes at (12,000 rpm).
11. Carefully discard supernatant. Keep only DNA in the bottom tube.
12. Add 500µl of 70% Ethanol (-20°C) for pellet rinsing.

13. Centrifuge for 5min. (12,000rpm).
14. Discard supernatant.
15. Add 500 $\mu$ l of 95% Ethanol (-20°C) for pellet rinsing.
16. Centrifuge for 5min. (12,000rpm).
17. Discard supernatant.
18. Vacuum dry for 15- 30 minutes.
19. Add 100  $\mu$ l of 1x TE buffer and leave at room temperature for 10 minutes.
20. Store DNA samples at -20°C freezer.



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#### Appendix 4 PCR Processing

1. Ice preparing in the box
2. Tube preparing (clean your hand with alcohol).
3. Make a cocktail for 20 $\mu$ l PCR per reaction.

PCR extraction	1 Reaction	25 Reactions
H <sub>2</sub> O	15.34 $\mu$ l	383.5 $\mu$ l
10 X Buffer	2 $\mu$ l	50 $\mu$ l
50 mM MgCL	1 $\mu$ l	25 $\mu$ l
Primer (F)	0.2 $\mu$ l	5 $\mu$ l
Primer (R)	0.2 $\mu$ l	5 $\mu$ l
25 mM dNTP	0.16 $\mu$ l	4 $\mu$ l
Taq DNA polymerase	0.1 $\mu$ l	2.5 $\mu$ l

4. Cocktail tube vortex mix ~ 5 – 10 (second)
5. Take 19 $\mu$ l from cocktail tube to PCR tube.
6. Take 1 $\mu$ l from DNA tube (concentrate) to every reaction tube.
7. Put in PCR processing machine until machine shown forever and keep in freezer (-20°C).

### Appendix 5 Polyacrylamine Gel Electrophoresis Preparing (PAGE)

1. Glass rinsing with (75% alcohol).
2. Electrophoresis installing.
3. Take agarose 1% or 1g mixed with 1xTBE (100ml) into microwave until homogeneous and then put it on the bottom layer of electrophoresis (2-3ml).
4. 10% of polyacrylamine gel in lower layer:

Raw material	One gel
Water	15ml
5xTBE	6.5ml
Acrylamide: Bis (29:1)	11ml
APS	224 $\mu$ l
TEMED	11.5 $\mu$ l
<b>Total</b>	<b>32.5ml</b>

5. 4% of acrylamine gel in upper layer:

Raw material	One gel
Water	4.3ml
5xTBE	0.6ml
Mixed from lower	2ml
<b>kept in freezer (4°C) until use</b>	
APS	35 $\mu$ l
TEMED	4 $\mu$ l

6. Mixed lower together and take it into electrophoresis and quickly put ~ 2ml of Buthanol into upper of electrophoresis glass.

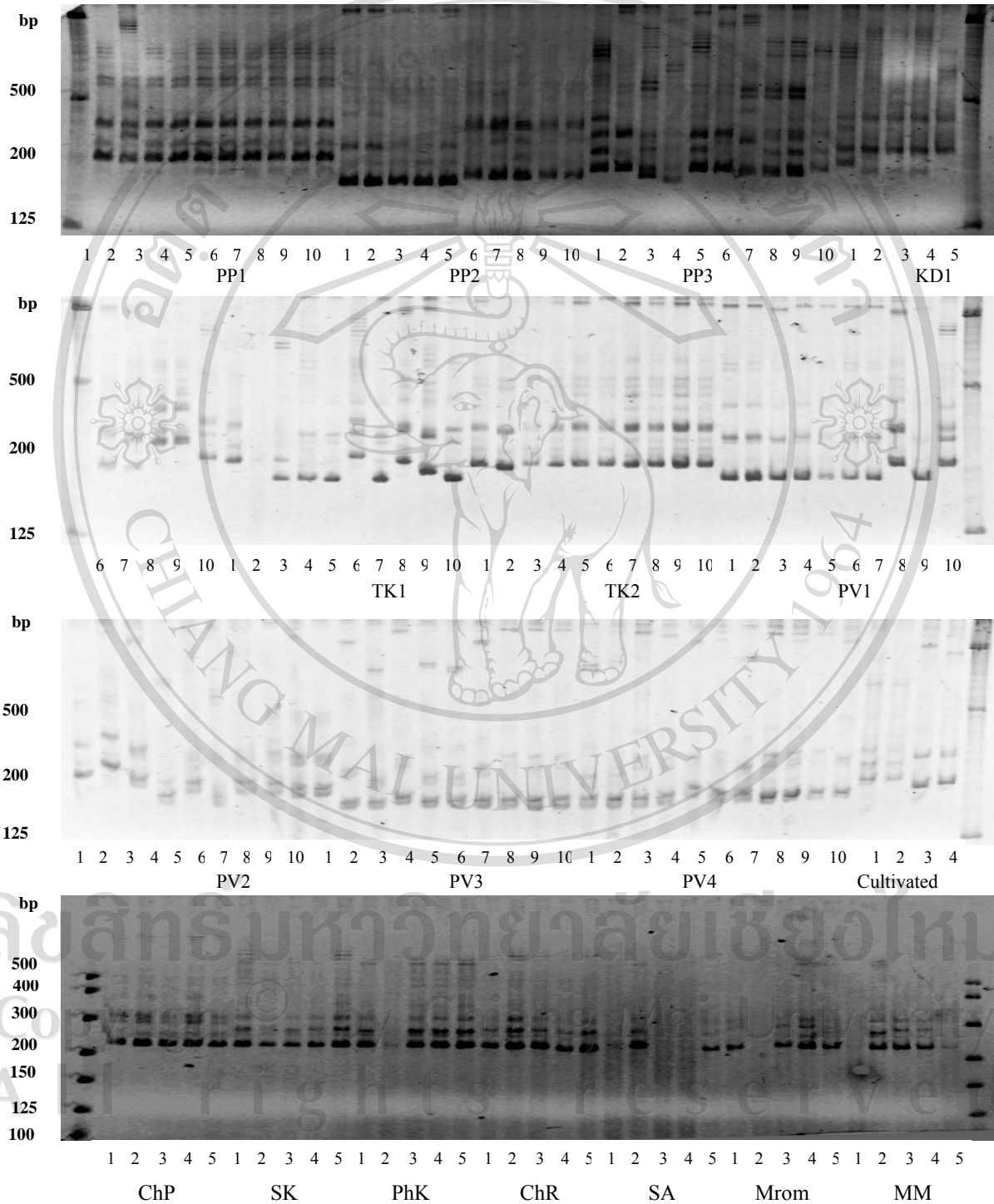
7. Waiting about 30 min or until the gel were polymerized.

8. Butanol water pouring out from electrophoresis and than it's cleaning with the water.
9. Comb molding.
10. Put the upper concentration on the lower layer and waiting may be 30 min until to become coagulated again.
11. To prepare box electrophoresis with water (1xTBE).
12. Carefully to take the electrophoresis glass into a box electrophoresis.
13. Comb drawing out.
14. PCR (freezer) mixed loading dye (12-13 $\mu$ l) per each PCR tube and put in the hole.



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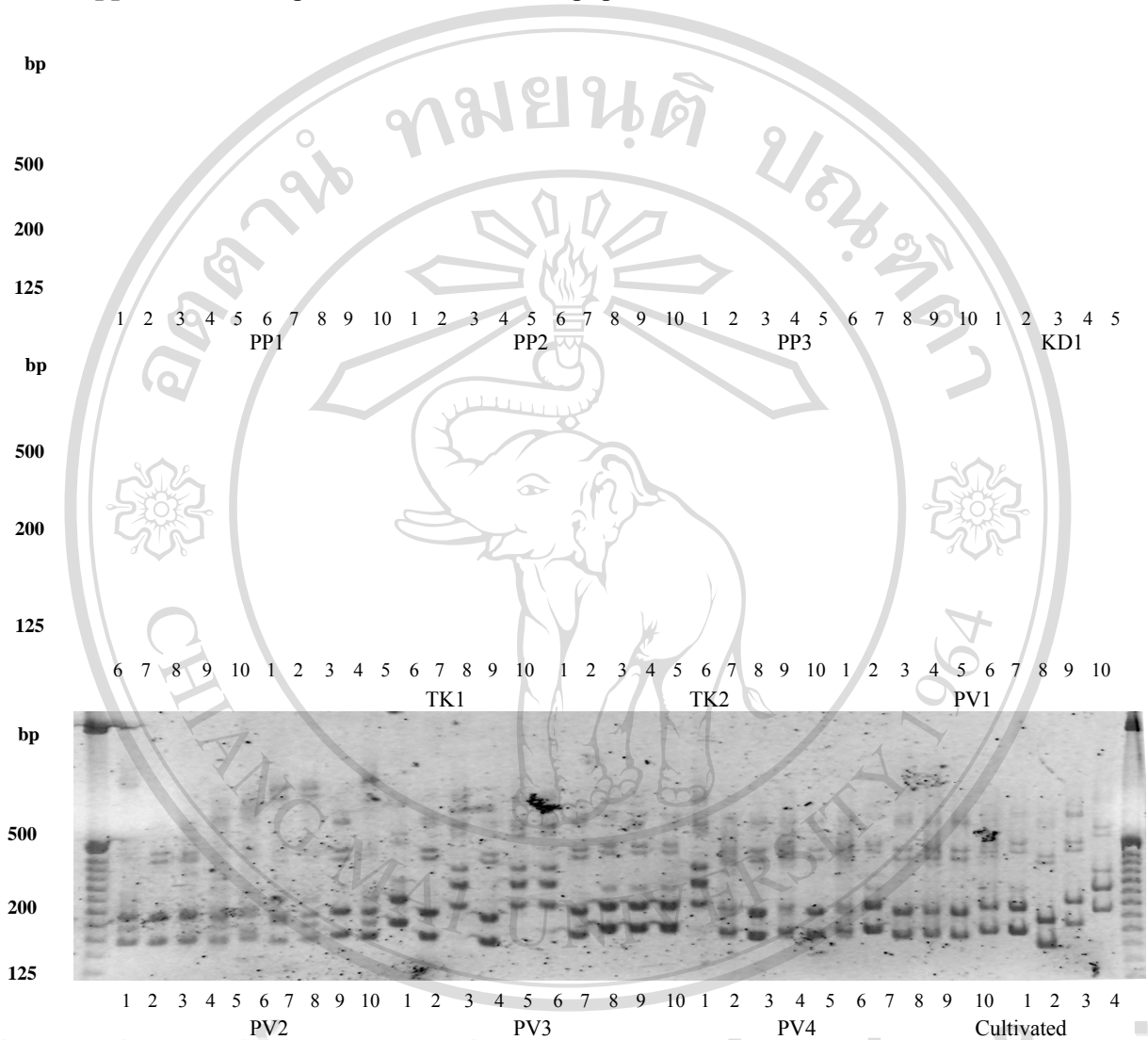
**Appendix 6** Band patterns of 10 wild rice populations and 7 local rices at RM20 locus.



PP = Phnom Penh, KD = Kandal, TK = Takeo, PV = Prey Veng, ChP = Chhmar Prom, SK = Srau Krahorm, PhK = Phkar Khgney, ChR = Chomkoum Rumpak, SA = Sombok Angkrorong, MM = Mong Mang.



**Appendix 7** Band patterns of 10 wild rice populations and 7 local rices at RM164 locus.

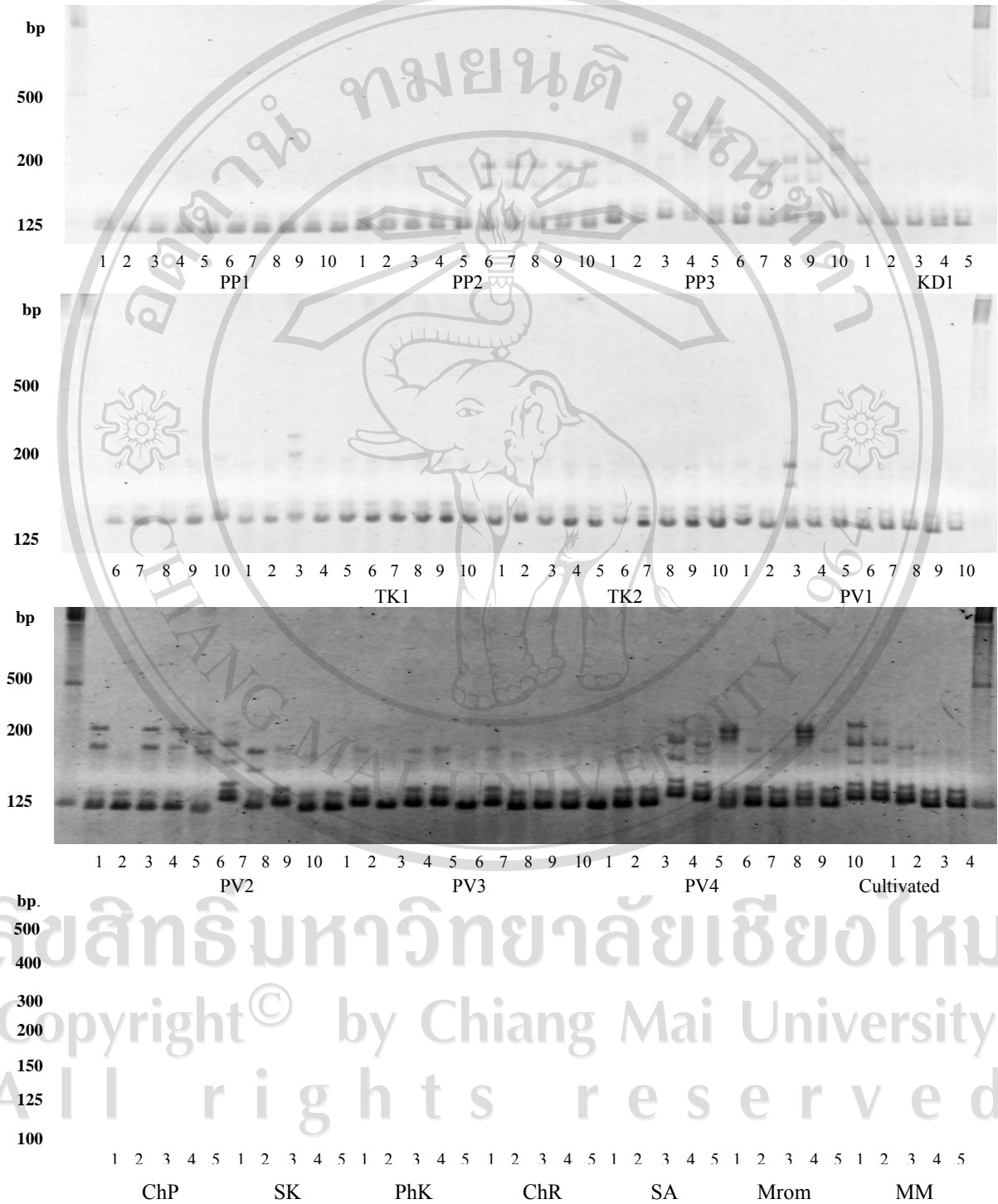


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1 2 3 4 5 1 2 3 4 5 1 2 3 4 5 1 2 3 4 5 1 2 3 4 5 1 2 3 4 5 1 2 3 4 5  
 ChP SK PhK ChR SA Mrom MM

PP = Phnom Penh, KD = Kandal, TK = Takeo, PV = Prey Veng, ChP = Chhmar Prom, SK = Srau Krahorm, PhK = Phkar Khgney, ChR = Chomkoum Rumpak, SA = Sombok Angkrorong, MM = Mong Mang.

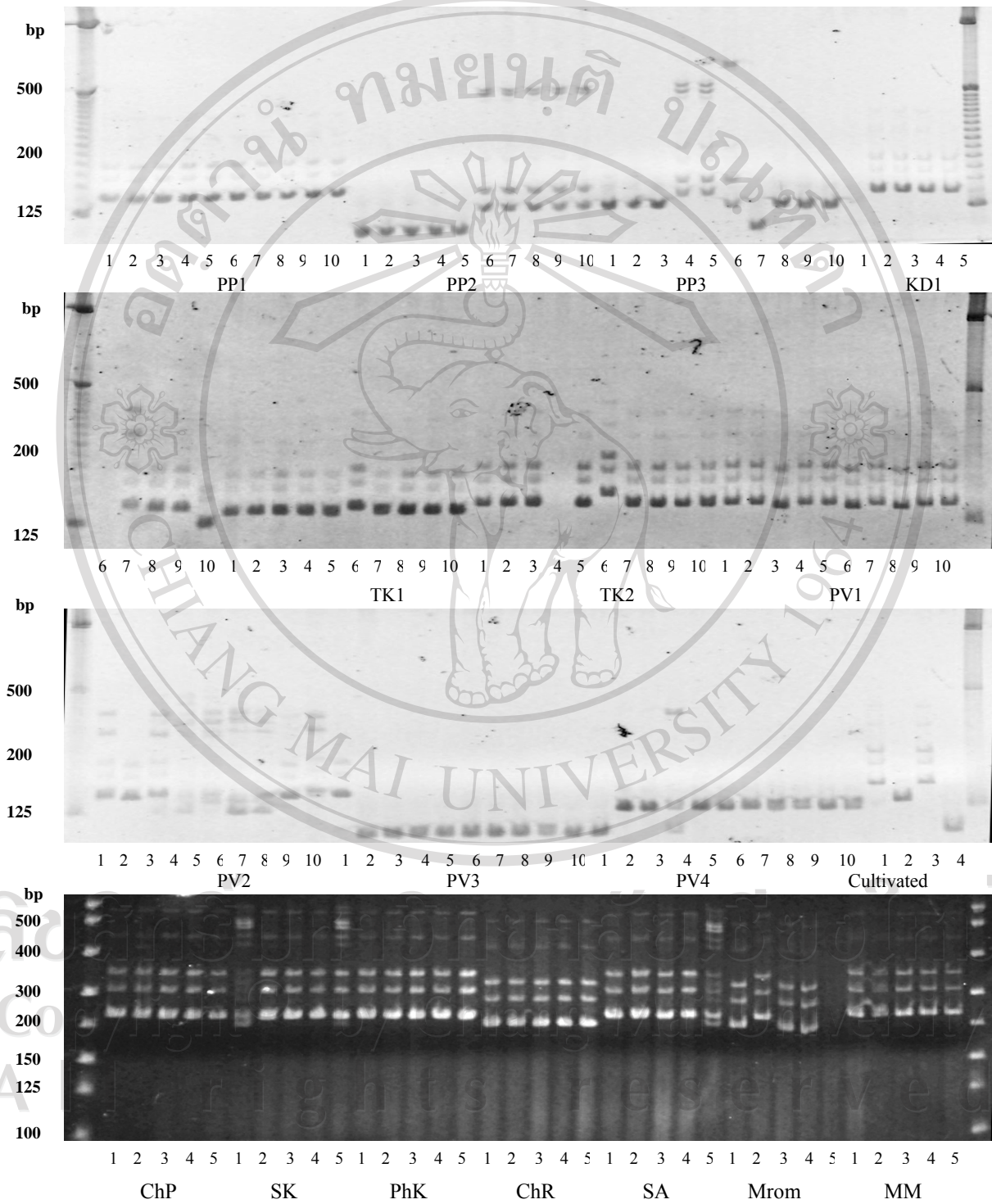
**Appendix 8** Band patterns of 10 wild rice populations and 7 local rices at RM225 locus.



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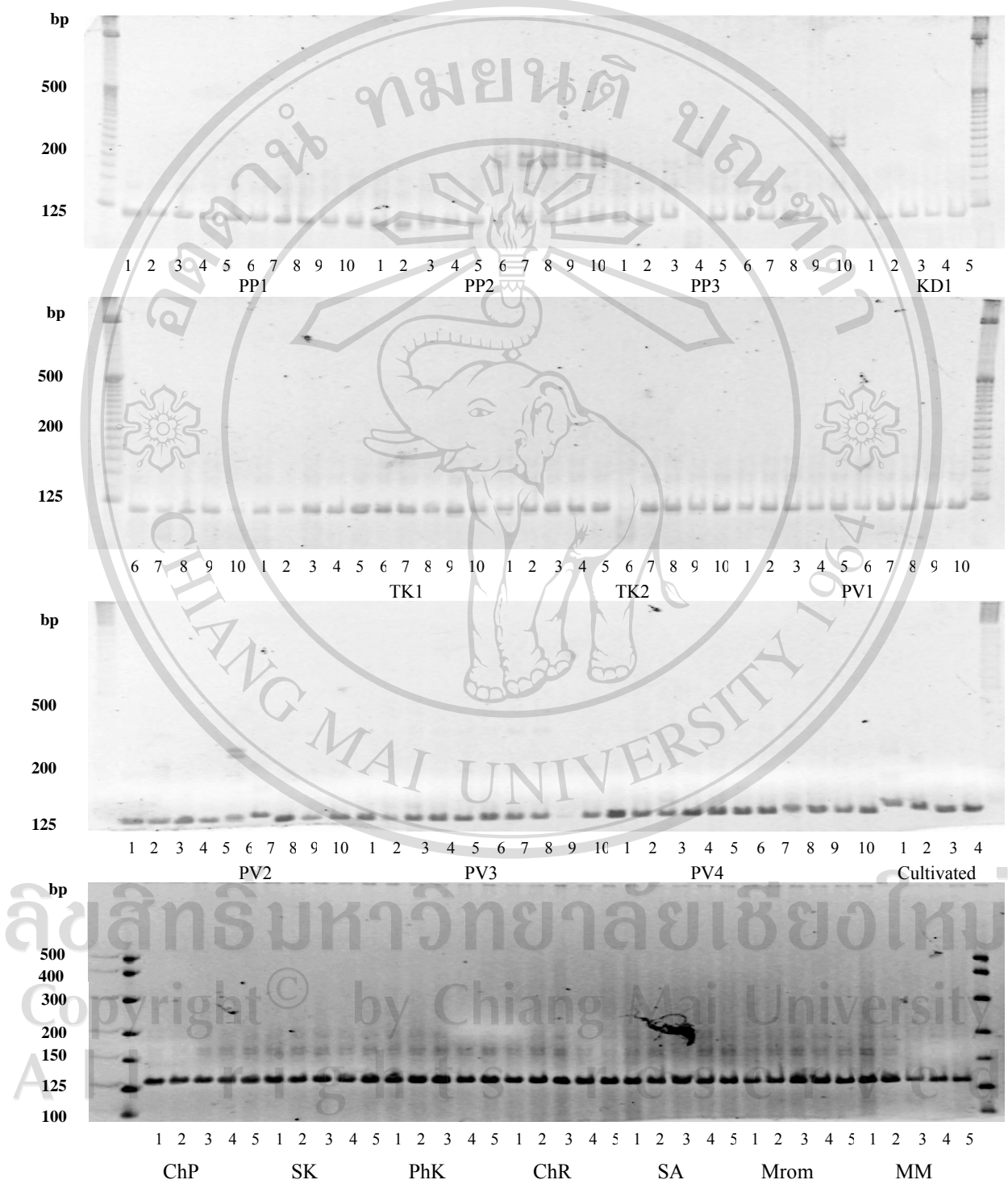
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**Appendix 9** Band patterns of 10 wild rice populations and 7 local rices at RM341 locus.



PP = Phnom Penh, KD = Kandal, TK = Takeo, PV = Prey Veng, ChP = Chhmar Prom, SK = Srau Krahorm, PhK = Phkar Khgney, ChR = Chomkoum Rumpak, SA = Sombok Angkrorong, MM = Mong Mang.

**Appendix 10** Band patterns of 10 wild rice populations and 7 local rices at RM588 locus.



PP = Phnom Penh, KD = Kandal, TK = Takeo, PV = Prey Veng, ChP = Chhmar Prom, SK = Srau Krahorm, PhK = Phkar Khgney, ChR = Chomkoum Rumpak, SA = Sombok Angkrorong, MM = Mong Mang.

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