# **Appendix**

### Chemical substance of electrophoresis preparation

1. Extraction buffer

Tris-buffer 0.2 M pH 8.4 100 ml.

Preparation method

Stock A: 0.2 M Tris(hydroxymethyl) aminomethane (tris 2.42 g. in 100 ml. of water)

Stock B: 0.2 M HCl (1.7ml. of HCl 37% concentration in 100 ml. of filter water)

Extraction buffer was mixed together between 50 ml. of stock A and 16.5 ml. of stock B in 133.5 ml. of distil water total volume was 200 ml. and adjust pH was 8.4 by 0.2 M HCl 1N. NaOH, after that put this solution in dark brown color bottle and kept in refrigerator

### 2. Gel composite

2.1 Acrylamide / Bis (30% T, 2.67%C)

Preparation

A: 29.2 g of Acrylamide in 50 ml. of distil water

B: N, N- Methylene – bis acrylamide 0.8 g.

Mixed together between A and B in refrigerator

2.2 Tris – HCl (1M) and pH 8.8

Preparation

A: Tris – HCl (1M) (18.15 g. of Tris in 100 ml. of distil water)

B: 1M HCl (8.35ml of HCl (37%) in 100ml. of water

Mixed together between 50 ml. of A and 8 ml. of B and adjust pH was 8.8 by 1.0M HCl put this solution in dark brown color bottle and kept in refrigerator

3. Electrod buffer (pH 8.3)

Tris 3 g

Glycine 14.4 g

Preparation

Mixed together in 500 ml. of distil water

# 4. Plant sample preparation

- 4.1 The leaf was clean and put in plastic bag kept in low temperature
- 4.2 Freeze mortar
- 4.3 The weight of leaf sample was 1g
- 4.4 The leaf was fine grain add 0.05 g. of PVPP(Polyvinyl polypyrrolidon) /sample
- 4.5 Solution of leaf was contained in close plastic tube for centrifuge and kept it in low temperature

Centrifuge solution of leaf

- Turn on centrifuge machine 30 minute and set temperature 2C with 3000 rpm.
- After put solution of leaf in centrifuge machine set 14,000 rpm at 2C for 30 minute

- Separate transparent solution portion on sediment by suction method be careful contact sediment, and kept in low temperature (don't shack)
- 5. 7.5% gel preparation method (2 gel)
  - 5.1 Acrylamide 5 ml
  - 5.2 Distil water 9.7ml.
  - 5.3 Tris HCl pH 8.8 (1M) 5ml
  - 5.4 10% ammonium persulfate 200 ml.(0.1/1ml. of water)
  - 5.5 TEMED 10 µl
- 6. Extract sample + Marker dye.
  - 6.1 Marker dye 10 µl
  - 6.2 Extract sample 90 µl

Two solution mixed together and kept in low temperature

### 7. Enzyme dyestuffs preparation

- 7.1 Peroxidase
  - A: 0.042g of 3-amino-9-ethylcabazole was dissolve by acetone 10 ml (dark condition)
  - B: 0.29g. of \( \beta \)- naphthol was dissolve by acetone 10 ml.(dark condition)

C: Tris- buffer 0.1 M pH 4.0

80 ml.

D:H<sub>2</sub>O<sub>2</sub> 3%

100 ml.

Preparation method: A+B+C mixed together and mixed with D when dye gel.(dark condition)

7.2 Esterase

A: phosphate – buffer pH 6.0

100 ml.

B: Fast blue \(\beta\)-salt

0.15 g.

C: naphthyl acetate 0.003g. dissolve in 3ml. of absolute ethanol

Preparation method: A+B mixed together after that filed it before dye 30 minute.(dark condition)

7.3 Acid-phosphatase

A: Acetate buffer pH 5.0

5 ml

B:Fast ganet GBC disodium salt

50 ml.

C: Disodium naphthyl pospate

25 mg.

Preparation method: A+B+C mixed together and filed it in dark condition, maculate gel in this solution (2-12 hr. in dark condition)

#### 8. Fixing solution

After dye gel until brand appear and clean gel with water after that fixing brand by fixing solution (10% glycerin+7% acetic acid)



สถานบริการวิทยาศาสตร์และเทคโนโลยี มหาวิทยาลัยเชียงใหม่ (สวท-มช.) ชั้น 7 อาคาร 30 ปี คณะวิทยาศาสตร์ มหาวิทยาลัยเชียงใหม่ เชียงใหม่ 50200 โทรศัพท์ : 053-943397, 053-941971 โทรสาร : 053-892275 E-mail : stsc@science.cmu.ac.th

Science and Technology Service Center, Chiang Mai University 7<sup>th</sup> Floor, 30<sup>th</sup> year Science Building, Faculty of Science, Chiang Mai University, Chiang Mai 50200

# รายงานผลการวิเคราะห์

เลขที่รับงาน : 003/006

วันที่รับตัวอย่าง : 3 มีนาคม 2549

วันที่รายงานผล: 25 เมษายน 2549

ตัวอย่าง: สารสกัดจากหางใหล

ชื่อของลูกค้า / หน่วยงาน : นายอรุณ โสตถิกุล

ที่อยู่ : สถาบันวิจัยและฝึกอบรมการเกษตร ลำปาง

**โทรศัพท์** : 04-6081039

โทรสาร: 054-342550

ผลการวิเคราะห์ตัวอย่างดังเอกสารแนบ

ผลการตรวจสอบ/วิเคราะห์ตามเอกสารข้างต้นนี้ รับรองเฉพาะตัวอย่างที่ได้ตรวจวิเคราะห์ เท่านั้น ไม่รับรองวัตถุหรือสินค้าที่ใช้เครื่องหมายเดียวกับตัวอย่างนี้ และห้ามใช้รายงานฉบับนี้ในการ ประกาศ หรือย่อตัดทอน

อนุมัติผลโดย

40-NI

(รองศาสตราจารย์ ดร.นวลศรี รักอริยะธรรม)

ผู้อำนวยการ สถานบริการวิทยาศาสตร์และเทคโนโลยี มหาวิทยาลัยเชียงใหม่

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# ผลการวิเคราะห์ตัวอย่างโดยใช้ GC-MS

เลขที่เอกสารจาก สวท-มช ศร 0515(28)/0137 ลงวันที่ 6 มีนาคม 2549 จำนวนตัวอย่าง: 2 (003/006)

- 1. ข้อมูลของตัวอย่าง ประกอบด้วย
  - 1.1 TIC (Total Ion Chromatogram)
  - 1.2 Area Percent Report
  - 1.3 Summary Library Search Report
    - 1.3 ข้อมูลแมสสเปกตรัมของแต่ละพืกและการเปรียบเทียบกับแมสสเปกตรัมมาตรฐาน
- 2. เครื่องมือ/สภาวะในการวิเคราะห์
  - 2.1 GC 6890 Agilent Technologies

Inlet : 270 °C

ปริมาณในการฉีด 0.5 µL split ratio 40 : 1

Oven : 80 °C---10 °C/min ---> 260°C(42 min)

Carrier: Helium 1.0 ml/min

Column : HP-5MS 30 m x 0.25 mm ID x 0.25 µm film thickness

2.2 MSD 5973(EI) Hewlett Packard

MS Quadrupole

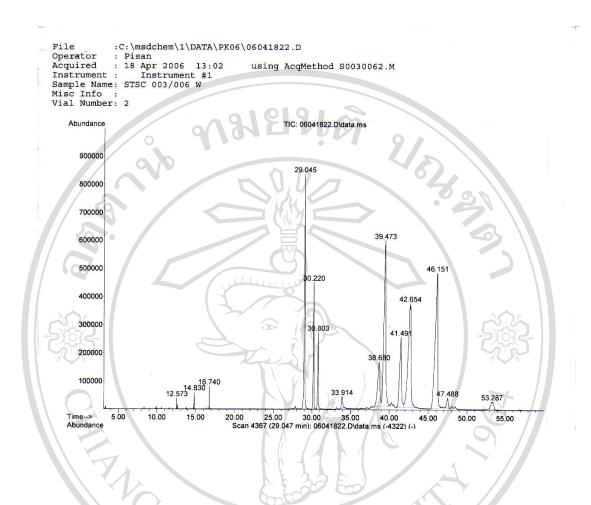
150 °C

MS Source

: 230 °C

3. วันส่งผลการวิเคราะห์ถึง สวท-มช : 25 เมษายน 2549

Copyright by Chian Mai University All right for the fung h t (นายพิสัณห์ คิจสวัสดิ์ใหญูลย์ e r v e d



Chemical components in sediment powder of derris root extracted by water

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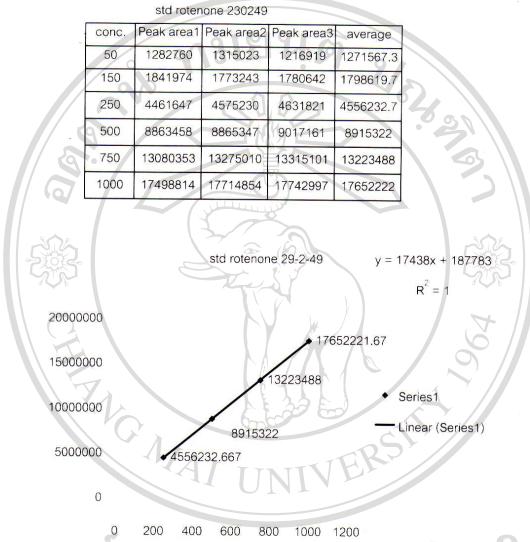
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                                                                           CAS#
                                                                                   Qual
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Oxacyclopentadec-6-en-2-one, (2)-
(CAS) $$ (Z)-5-TETRADECEN-14-QILID
                                                              151483 063958-52-1 92
                    Oxacyclopentadec-6-en-2-one, (Z)-1H-Indene, octahydro-, cis- (CAS)
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      30.801
              3.59 unknown
      33.916 0.86 unknown
      38.679
               6.20 unknown
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                    uro[2,3-h][1]benzopyran-6(6aH)-one
                       1, 2, 12, 12a-tetrahydro-8, 9-dimeth
                    oxy-2-(1-methylethenyl)-, [2R-(2.a
                    lpha., 6a.alpha., 12a.alpha.) ] - $$ C
                    ube-Pulver $$ Dactinol $$ Deril $$
                     Derrin $$ Derris $$ Derris (insec
                    ticide) $$ Dri-ki
                    Rotenone
                                                                 328973 000083-79-4 87
                    Dequelin
                                                                 328971 000522-17-8 78
10 41.492 8.26 C:\Database\wiley7n.1
                    Rotenone
                                                                 328974 000083-79-4 96
                    Rotenone $$ [1]Benzopyrano[3,4-b]f 328973 000083-79-4 96 uro[2,3-h][1]benzopyran-6(6aH)-one
                    nro[2,3-n][1]Benzopyran-o(dan)-one
, 1,2,12,12a-tetrahydro-8,9-dimeth
oxy-2-(1-methylethenyl)-, [2R-(2.a
lpha.,6a.alpha.,12a.alpha.)]- $$ C
ube-Pulver $$ Dactinol $$ Deril $$
                     Derrin $$ Derris $$ Derris (insec
                    ticide) $$ Dri-ki
                    Isorotenone
                                                                 328976 000083-79-4
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                    3.ALPHA., 5-CYCLO-ERGOSTA-7, 22-DIEN 329244 003037-46-5
                   -6-ONE $$ 3,5-Cycloergosta-7,22-di
en-6-one, (3.beta.,5.alpha.,22E)-
```

(CAS) \$\$ 3.alpha.,5-Cyclo-5.alpha. -ergosta-7,22-dien-6-one (CAS) \$\$ Cyclopenta[a]cyclopropa[2,3]cyclopenta[1,2-f]naphthalene, 3,5-cycloe

rgosta-7,22-dien-

```
12 46.153 21.99 C:\Database\wiley7n.1
                Rotenone $$ [1]Benzopyrano[3,4-b]f 328974 000083-79-4 96
                uro[2,3-h][1]benzopyran-6(6aH)-one
                 , 1,2,12,12a-tetrahydro-8,9-dimeth
                 oxy-2-(1-methylethenyl)-, [2R-(2.a)]
                 lpha., 6a.alpha., 12a.alpha.)] - $$ C
                 ube-Pulver $$ Dactinol $$ Deril $$
                Derrin $$ Derris $$ Derris (insecticide) $$ Dri-ki
                Isorotenone
                                                     328976 000083-79-4 90
                Rotenone $$ [1]Benzopyrano[3,4-b]f 328977 000083-79-4 87
                uro[2,3-h][1]benzopyran-6(6aH)-one
                  1,2,12,12a-tetrahydro-8,9-dimeth
                oxy-2-(1-methylethenyl)-, [2R-(2.a)]
                lpha., 6a.alpha., 12a.alpha.) ] - (CAS
                ) $$ Deril $$ Ronone $$ Derris $$
                Derrin $$ Rotenon $$ Noxfish $$ Da
                ctinol $$ Rotocid
            0.99 C:\Database\wiley7n.1
                                                    328973 000083-79-4 91
                Rotenone
                Deguelin $$ 3H-Bis[1]benzopyrano[3 328971 000522-17-8 72
                 ,4-b:6',5'-e]pyran-7(7aH)-one, 13,
                13a-dihydro-9, 10-dimethoxy-3, 3-dim
                ethyl-, (7aS-cis)- (CAS) $$ 3H-Bis
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                -7(7aH)-one, 13,13a-dihydro-9,10-d
                imethoxy-3,3-dimethyl- $$ 3H-Bis[1
                ]benzopyrano[3,4-
                Rotenone $$ [1]Benzopyrano[3,4-b]f 328977 000083-79-4 76
                uro[2,3-h][1]benzopyran-6(6aH)-one
                , 1,2,12,12a-tetrahydro-8,9-dimeth
                oxy-2-(1-methylethenyl)-, [2R-(2.a lpha.,6a.alpha.,12a.alpha.)]- (CAS
                  $$ Deril $$ Ronone $$ Derris $$
                Derrin $$ Rotenon $$ Noxfish $$ Da
                ctinol $$ Rotocid
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14
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                Isorotenone
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                uro[2,3-h][1]benzopyran-6(6aH)-one
                , 1,2,12,12a-tetrahydro-8,9-dimeth
                oxy-2-(1-methylethenyl)-, [2R-(2.a
                lpha., 6a.alpha., 12a.alpha.)] - $$ C
                ube-Pulver $$ Dactinol $$ Deril $$
                 Derrin $$ Derris $$ Derris (insec
               ticide) $$ Dri-ki
                    by Chiang Mai University
```

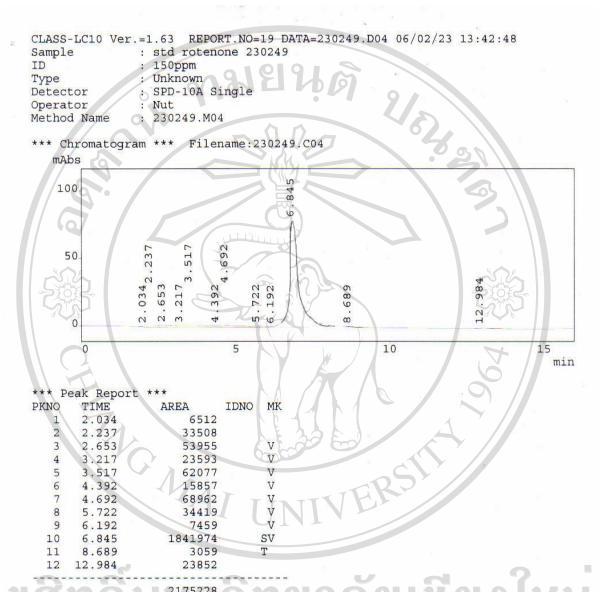
# Relation of peak area and standard rotenone concentration



# ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่

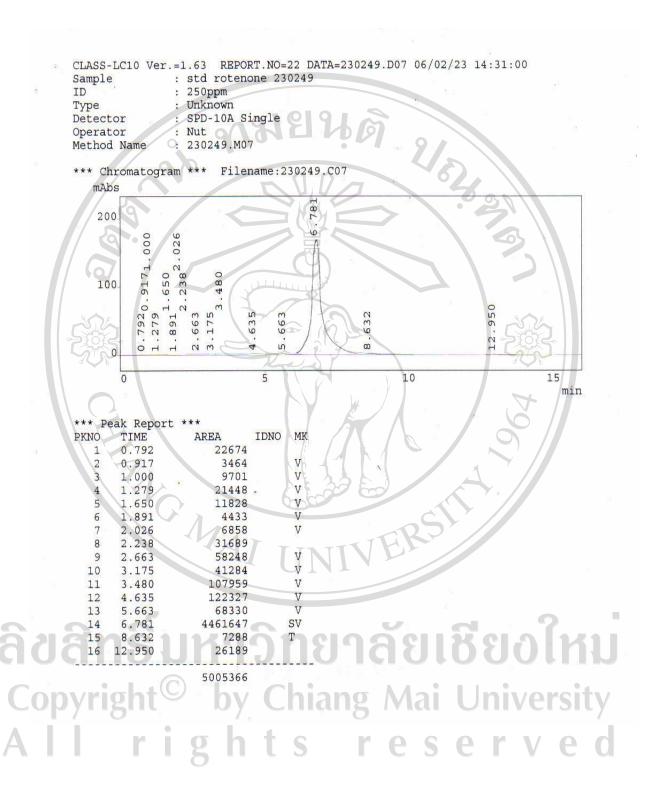
Regression equation between standard rotenone concentration and peak area of HPLC

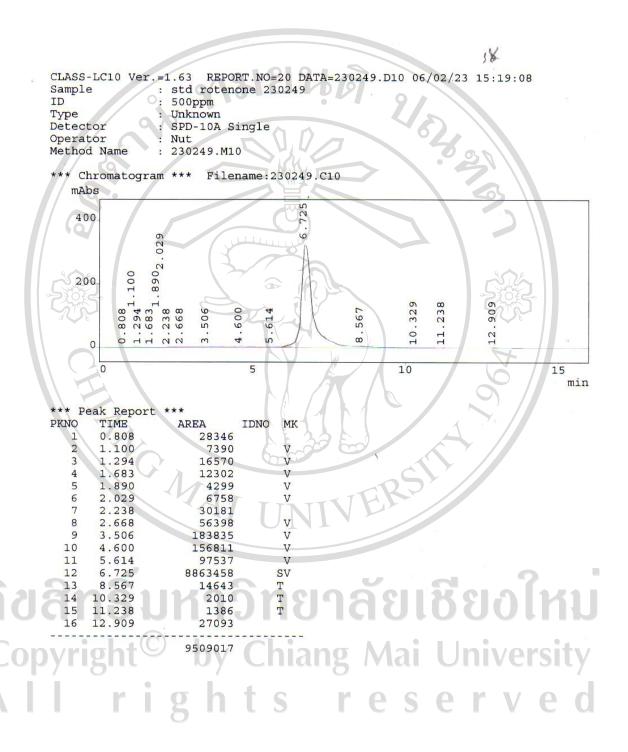
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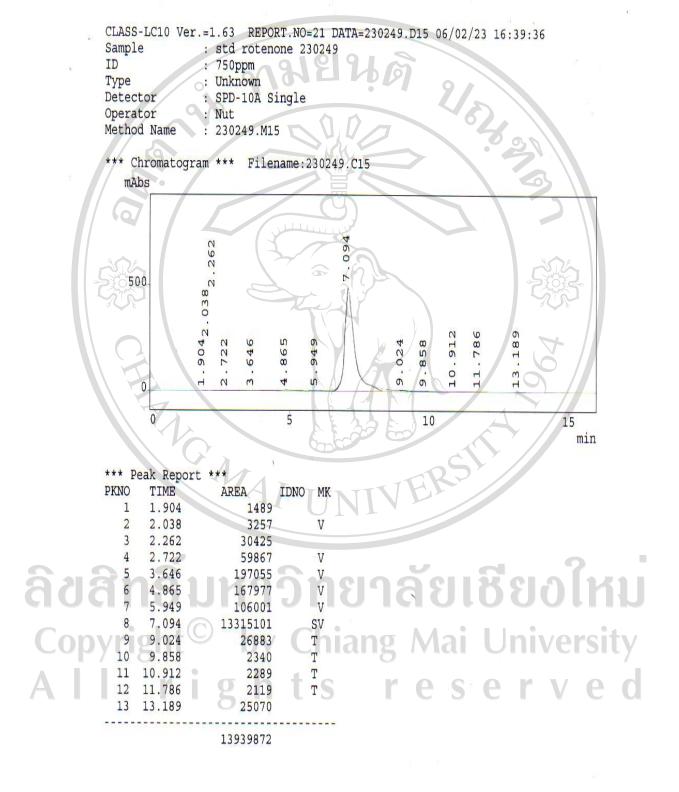


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ID
               : 1000ppm
Type
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                                                 2/52/38
Detector
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Operator
               : Nut
Method Name
               : 230249.M16
*** Chromatogram *** Filename:230249.C16
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   500
                                                      10.936
                                                                203
                      .640
                                5.965
                            878
                             5
                                                  10
                                                                         15
                                                                           min
*** Peak Report ***
PKNO
       TIME
                  AREA
                            IDNO
                                 MK
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       0.843
       1.901
                      1736
                                  V
                      3468
       2.037
       2.263
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       2.748
                      61234
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       4.878
                    198941
       5.965
                    126060
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       7.113
       9.044
                     36294
       9.867
                      2910
      10.936
                      2908
  13
     11.795
                      2395
  14 13.203
                      24849
                  18217067
```

#### Tween 80

Tween 80 (also known as Polysorbate 80) is used in the manufacture of protein solution formulations to help solubilize and stabilize the protein. It is one of a series of materials (including Tween 20, 40 and 60) which are fatty acid esters of sorbitan polyethoxylates. The various Tweens differ in the type of fatty acid present; Tween 80 is an oleate. Tween 80 is a nonionic surfactant and emulsifier derived from sorbitol which is obtained from various types of fruit. Polysorbate 80 is a water-soluble somewhat yellowish amber liquid that is used as a dispersing agent to mix oil and water and to solubilize fragrances and essential oils. It is also a great lubricant and has a pleasant, soothing effect on the skin. Polysorbates are nonirritating and readily biodegradable. Tween 20 - Polysorbate 20 is derived from coconut oil. Other polysobates are derived from palm oil (Polysorbate 40 and 60) and olive oil (polysorbate 80).

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#### **CURRICULUM VITAE**

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**MARRITAL STATUS:** Married

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**POSITION:** Instructor / Head of Plant Protection Branch

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### **EDUCATION:**

Field of	Institute	Qualification	From	To
Study	(3/3)		<b>\</b>	
Entomology	Chiang Mai University, Chiang Mai, THAILAND	B.Sc.	1978	1981
Entomology	Kasetsart University, Bangkok,	M.Sc.	1985	1988
	THAILAND	TVI.Del	7	1700

### **WORK EXPERIENCE:**

**Research** publication more than 20 publications such as

- 1. Determine of Rotenone Degradation after Sprayed Derris Extract by High Performance Liquid Chromatography (HPLC.) Method. (2005-2006)
- 2. Rotenone Quantity and Isozyme Patterns of *Derris malacensis* Prain and *Derris elliptica*. (2005-2006)
- 3. Formulation of derris extract to controlled Cabbage aphid (*Lipaphis erysimi* Kattenbach) (2005-2006)
- 4. Increasing effective of water extracted rotenone from derris root (2005-2006)

**Teaching and Training** (for student, farmers / agricultural and extension officers/secondary school teachers)

Training course: Important Insect Pest and their Control by Botanical Insecticide (1990-at present)